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Evolving paradigms in biological carbon cycling in the ocean

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44 **ABSTRACT**

45

46 Carbon is a keystone element in global biogeochemical cycles. It plays a fundamental role in
47 biotic and abiotic processes in the ocean, which intertwine to mediate the chemistry and redox
48 status of carbon in the ocean and the atmosphere. The interactions between abiotic and
49 biogenic carbon (e.g., CO₂, CaCO₃, organic matter) in the ocean are complex, and there is a
50 half-century-old enigma about the existence of a huge reservoir of recalcitrant dissolved
51 organic carbon (RDOC) that equates to the magnitude of the pool of atmospheric CO₂. The
52 concepts of the biological carbon pump (BCP) and the microbial loop (ML) shaped our
53 understanding of the marine carbon cycle. The more recent concept of the microbial carbon
54 pump (MCP), which is closely connected to those of the BCP and the ML, explicitly considers
55 the significance of the ocean's RDOC reservoir and provides a mechanistic framework for the
56 exploration of its formation and persistence. Understanding of the MCP has benefited from
57 advanced “omics”, and novel research in biological oceanography and microbial
58 biogeochemistry. The need to predict the ocean’s response to climate change makes an
59 integrative understanding of the BCP, ML and MCP a high priority. In this review, we
60 summarize and discuss progress since the proposal of the MCP in 2010 and formulate research
61 questions for the future.

62

63

64 **Keywords:** biological carbon pump, microbial loop, microbial carbon pump, ocean carbon
65 cycle, global climate change

66

67 **INTRODUCTION**

68

69 The modern ocean accounts for ~50% of global photosynthesis, with its primary production of
70 organic matter forming the core of the ocean carbon cycle. Thus the ocean has a major
71 influence on the chemistry and redox status of the atmosphere through the net uptake of
72 atmospheric CO₂ and net release of molecular oxygen. An early estimate showed that about 25%
73 of the ocean's primary production was transported to the interior of the ocean (below the
74 euphotic zone) via the biological carbon pump (BCP) (Falkowski et al., 2000); later on this
75 number was changed to 10-15% for gravitational sinking with another 5% each for passive
76 transport by water motion and active transport by vertical migrators (Siegel et al., 2016).
77 Carbon transported to the deep ocean (> 1000 m) is sequestered on timescales of > 100 years
78 up to 1000 years (i.e. the residence time of deep waters). About 0.3% of the ocean's primary
79 production is buried in marine sediments (Dunne et al., 2007; Ridgwell and Arndt, 2015),
80 some of which eventually forms a major reservoir of organic matter that persists for hundreds
81 of millions of years in rock formations (Fig. 1).

82

83 **【insert Figure 1 here】**

84

85 Since the industrial revolution, the ocean is estimated to have taken up approximately 25% of
86 the anthropogenic CO₂ (Le Quéré et al., 2018), resulting in ocean acidification with
87 consequences for biogeochemical and climatological processes, and the ocean carbon cycle
88 (Falkowski et al., 2000; Cai, 2011; Bauer et al., 2013; Laruelle et al., 2018). Global warming
89 and ocean acidification and their respective consequences influence the functioning of the BCP,
90 a major pathway for sequestering atmospheric CO₂ in the ocean. The microbial carbon pump
91 (MCP) (Jiao et al. 2010) provides an additional path for carbon sequestration within the
92 marine ocean carbon cycle (Stone, 2010), which is intimately linked to climate change.

93

94 The BCP is the mechanism by which carbon-containing compounds are exported via
95 biological processes from the surface to the deep ocean (Sarmiento and Gruber, 2006),
96 whereas the MCP addresses the dissolved organic carbon (DOC) pool, specifically the
97 recalcitrant (R) DOC (Fig. 1), which constitutes the majority of DOC and persists in the ocean
98 for up to 4000–6000 years (Bauer et al., 1992; Hansell, 2013). Hansell (2013) defines RDOC
99 as “DOC that is resistant to rapid microbial degradation and so has accumulated and is

100 observable in the ocean”. Concentrations of DOC in the open ocean range from 360–960

101 µg/kg (or 30-80 µmol/kg) (Hansell et al., 2009) with significant seasonal variation often seen
102 in surface waters (Copin-Montégut and Avril, 1993). Accounting for a global ocean inventory
103 of 662 Gt C, the huge DOC pool is almost equal to the carbon dioxide pool (750 Gt C) in the
104 atmosphere. Therefore, the biogeochemical behavior of the DOC pool has important
105 implications for the ocean carbon cycle and climate.

106

107 The MCP mediates the transformation of labile carbon to RDOC, which builds on elements of
108 the previously recognized processes involved in ocean carbon cycling and storage (Benner and

109 Amon, 2015), namely the BCP, microbial loop (ML) and viral shunt (VS). The functioning of
110 the MCP also impacts nutrient stoichiometry when preferentially remineralizing N and P from
111 dissolved organic matter (DOM). This DOM is produced via the VS (Wilhelm and Suttle,
112 1999; Suttle, 2007) and other processes such as phytoplankton excretion and zooplankton
113 sloppy feeding (Roy et al., 1989; Ducklow et al., 1995; Biddanda and Benner, 1997; Strom et
114 al., 1997; Arrigo, 2007; Moller 2007). This recycling of nutrients enhances local primary
115 production while enriching the remaining DOM in carbon, thus lowering its nutritional value.

116

117 The detailed processes of the MCP are currently not well understood. This is largely due to
118 microbial complexity and the vast unresolved chemical structures of DOM compounds.
119 Growing efforts have been devoted to use microbiological and geochemical tools to bridge the
120 gap between microbial omics and organic carbon composition (Kujawinski, 2011; Lechtenfeld
121 et al., 2015; Moran et al., 2016). In this review, we discuss important aspects of the BCP, the
122 ML and the MCP, and summarize progress that has been made concerning the MCP since Jiao
123 et al. (2010).

124

125

126 **EVOLUTION OF OUR UNDERSTANDING OF THE MICROBIAL ROLE IN** 127 **DOC GENERATION AND DEGRADATION**

128

129 Understanding of the ocean's carbon cycle in the late 20th century was largely promoted by the
130 biological carbon pump (called "soft tissue pump" in Volk and Hoffert, 1985) and the
131 microbial loop (Azam et al., 1983). The term "pump" was initially used to refer to the
132 movement of carbon against a concentration gradient between the surface ocean and the deep
133 ocean (Volk and Hoffert, 1985). Both concepts find their roots in Dugdale and Goering (1967),
134 who recognized new (BCP) and regenerated (ML) production in the ocean.

135

136 The BCP begins in the euphotic zone where photoautotrophic organisms fix dissolved CO₂ to
137 produce particulate organic carbon (POC) (Fig. 2). Particulate organic matter (POM) consists
138 of both living and nonliving components, and most of it is respired to CO₂ by metabolic
139 processes in the epipelagic ecosystem. The subsequent export of a small fraction of the POM
140 is carried out by gravitational flux, vertical migrations of zooplankton and physical subduction
141 of water masses, which remove the organic matter to deeper regions where it accumulates or is
142 respired. The respiratory CO₂ at depth is removed from contact with the atmosphere for a
143 period corresponding to the residence time of deep waters, i.e. tens to hundreds of years below
144 100 m and thousands of years below 1000 m (Fig. 2). In addition, organic matter in particulate
145 or dissolved form reaching the latter depth via the BCP should be considered as sequestered at
146 the time scale of climate change.

147

148 **【insert Figure 2 here】**

149

150 Increasing atmospheric CO₂ concentration raises several questions: "(1) Will the ocean
151 continue to take up carbon? (2) At what rate? (3) For how long will the exported carbon

152 *remain removed from the atmosphere?* These questions address the functioning and efficiency
153 of the future BCP. Global warming and past carbon sequestration (ocean acidification) will
154 also change the BCP leading to the next question: *(4) How will the biological pump respond to*
155 *the consequences of increased carbon input combined with warming?"* (Passow and Carlson,
156 2012). One scenario suggests that in the coming decades decreasing phytoplankton cell size
157 will decrease the downward POC flux from the surface ocean, while changes in zooplankton
158 community structure will decrease the downward POC flux in subsurface waters (Boyd, 2015).
159 However, other predictions suggest alternative outcomes and the answers to these questions
160 are still discussed controversially in the scientific community. In a recent report on a
161 transformative understanding of the ocean's BCP to the US National Science Foundation,
162 Burd et al. (2016) recommended three major research directions addressing "*(i) food web*
163 *regulation of export, (ii) the dissolved-particulate continuum, and (iii) variability of organic*
164 *transport in space and time*". Several large programs, e.g. the ongoing US -EXPORTS (Siegel
165 et al., 2016) and the UK COMICS (Sanders et al., 2016) programs, as well as many other
166 efforts are currently focusing on the BCP.

167

168 **Though many forms of vertical export can be related to BCP, it mainly** focuses on particles
169 that move downward through physical and biological forces (i.e., by gravity and transport by
170 vertically migrating zooplankton). The ML, on the other hand, intimately links intricate
171 interactions between microorganisms and their physical and chemical surroundings (Azam et
172 al., 1983; Azam, 1998). **The ML focuses on carbon cycling in the water column where bacteria**
173 **(actually referring to both bacteria and archaea), protozoa, and viruses determine the fate of**
174 **dissolved organic matter (Azam, 1998).** It was estimated that bacteria could channel up to 50%
175 of marine primary production into the microbial loop, highlighting their importance in the
176 ocean's carbon cycle (Azam, 1998; Fenchel, 2008). Similarly, Legendre and Rivkin (2008)
177 found that heterotrophic microbes always dominate respiration in the euphotic zone, even
178 when most particulate primary production is grazed by metazoans. The ML intertwines with
179 the grazing food web and provides a mechanism to retain nutrients such as N and P in the
180 highly stratified upper oligotrophic oceans by recycling them through pico-phytoplankton,
181 bacteria and microzooplankton (Azam et al. 1983) (Fig. 2).

182

183 **The MCP complements and connects the concepts of BCP and ML, additionally including the**
184 **idea of the viral shunt, into a more integrated concept of the cycling of biogenic carbon in the**
185 **ocean. The viral shunt, which refers to the release of carbon and nutrients back into the**
186 **environment due to cell lysis, is tightly connected to the BCP, the ML and the MCP because**
187 **cell lysis transforms living particulate organic matter (POM) into DOM and non-living POM**
188 **(Wilhelm and Suttle 1999; Suttle, 2005).** As much as a quarter of the C fixed by phytoplankton
189 is estimated to flow through the VS (Wilhelm and Suttle, 1999), thereby promoting ecosystem
190 respiration (Fuhrman, 1999). **The released DOM and POM are largely of bacterial origin, and**
191 **hence, relative to bacterial requirements (because of the carbon required for respiration) have**
192 **too little carbon relative to other nutrients. This shortage of carbon is exacerbated because of**
193 **the recalcitrant nature (e.g. cell-wall material) of some of the carbon released by cell**
194 **lysis.** Therefore, as the lysis products are processed by the ML, the more accessible DOM is
195 metabolized, releasing inorganic nutrients, altering pathways of nutrient cycling (Weitz et al.,

196 2015; Shelford and Suttle, 2018), and enriching the pool of less labile DOC. This process
197 directly couples the VS to the ML and MCP, and has been termed the ‘shunt and pump’ (Suttle,
198 2007).

199

200 The BCP, ML and MCP have distinct ecological or biogeochemical meanings (Table 1), and
201 each has influenced multiple research disciplines (Table 2). These three concepts are
202 fundamental in developing global biogeochemical and ecological models that rely on
203 understanding organismal biology and the interactions between the POC and DOC pools (Fig.
204 3).

205

206 **【insert Table 1 here】**

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209 **【insert Table 2 here】**

210

211

212 **【insert Figure 3 here】**

213

214 Several reviews provide thorough descriptions of the BCP and the ML (e.g. Kirchman, 2000;
215 [Sarmiento and Grüber, 2006](#); Fenchel, 2008; Honjo et al., 2008; Passow and Carlson, 2012).
216 Here we focus on recent progress concerning the MCP in the context of the BCP and ML.

217

218 **PROGRESS ON THE MCP DURING THE LAST EIGHT YEARS**

219

220 During the last eight years our understanding of the MCP has advanced appreciably (e.g. Jiao
221 and Zheng, 2011; Jiao et al., 2014, 2015; Lechtenfeld et al., 2014, 2015; Legendre et al., 2015;
222 Zhang, 2016; Chen et al., 2018; Jiao et al., 2018a), specifically addressing some of the
223 questions raised in Jiao et al. (2010). In particular, substantial progress has been made on
224 composition of recalcitrant DOM, the mechanisms of its formation, the nature of its
225 interactions with microbial loop biogeochemistry, and the associated community shifts and
226 trophic dynamics. There were also gains in our understanding of the microbial processing of
227 DOM at various taxonomic and functional group levels (e.g., Dang and Jiao, 2014;
228 Kujawinski et al., 2016; Sarmiento et al., 2016) (Table 3). The state of the art of these topics
229 will be discussed in the remainder of this review.

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231 **【insert Table 3 here】**

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236 IDENTIFICATION AND QUANTIFICATION OF THE COMPOSITION OF RDOM

237

238 According to Hansell et al. (2009) less than 1% of the DOC in the ocean is labile and 94% is
239 refractory, while the remaining 5% is classified as semi-labile (note: Hansell, 2013 divided the
240 DOC into labile, semi-labile, semi-refractory, refractory and ultra-refractory). **Much of the**
241 **RDOC production in the ocean can be attributed to microbial activities (e.g., Ogawa et al.,**
242 **2001).** Kaiser and Benner (2008) estimated that 25% of the total organic carbon (including
243 both POC and DOC) was of bacterial origin. Based on the estimates of Hansell et al. (2009)
244 and Kaiser and Benner (2008), Benner and Herndl (2011) calculated that about 10 Pg of
245 semi-labile DOC and 155 Pg of refractory DOC are of bacterial origin. Hansell (2013)
246 calculated rates of DOC production for different fractions based on meridional DOC
247 concentration gradients, with the production of RDOC having a rate of 0.043 Pg C/year, which
248 is comparable to the higher end of the RDOC production estimated by Benner and Herndl
249 (2011). Other authors have estimated RDOC production using different criteria. **Legendre et al.**
250 **(2015) estimated a rate of 0.2 Pg C/year for production of RDOC in the world's oceans at all**
251 **depths using the constraint of RDOC lifetime of > 100 years, which is the minimum residence**
252 **time for the ocean sequestration of carbon in the literature (the origin of the 100-year threshold**
253 **is explained in Legendre et al., 2015).** Walker et al. (2016a) calculated production rates of
254 low-molecular-weight DOM in the range of 0.11–0.14 Pg C/year as a proxy for RDOC
255 production in the deep ocean. These numbers interestingly are comparable to earlier estimates
256 from microbial incubation experiments (0.5–0.6 Pg C/year) (Brophy and Carlson, 1989).

257

258 Recent efforts to quantify the RDOC pool have been accompanied by progress in the
259 identification of the molecular composition of RDOC and the microbial populations that are
260 responsible for its production in the ocean water column. Microbial RDOC production will be
261 the focus of the following sections, whereas **RDOC turnover at deep-sea hydrothermal vents**
262 **(Hawkes et al., 2015; Lang et al., 2006; Walter et al., 2018) and other processes will not be**
263 **discussed.**

264

265 **Characterization of specific biochemicals in RDOM**

266

267 **Carbohydrates, amino acids and amino sugars.** Early studies examined the composition of
268 RDOC based on measurements of common biochemicals, such as carbohydrates, amino acids
269 and lipids. Ogawa et al. (2001) reported the transformation of labile substrates (D-glucose and
270 D-glutamate) into refractory forms of hydrolysable neutral sugars, amino sugars and amino
271 acids that persisted after one year in bioassay experiments. The concentrations of these
272 compounds were later confirmed to be similar to those reported for natural deep ocean waters
273 (Kaiser and Benner, 2009) and represented less than 2% of the total RDOC in
274 low-molecular-weight DOC (Benner and Amon, 2015). In particular, D-enantiomers of amino
275 acids have been observed to contribute to the RDOC pool and are predominantly derived from
276 bacterial sources (Kaiser and Benner, 2008, 2009). The ratio of the D-amino acids vs. L-amino
277 acids has been used as a proxy for the degree of recalcitrance, which increases dramatically
278 from bulk POM to the refractory low-molecular-weight DOM (Benner and Amon, 2015)
279 (Table 4).

280

281 【insert Table 4 here】

282

283 **Microbial lipids.** Microbial lipids may be important compounds contributing to the RDOC
284 pool in the ocean (Hwang et al., 2003). Some lipids are much more resistant to degradation
285 than carbohydrates or proteins (hydrolyzed to amino acids) (Benner and Amon, 2015) and can
286 be preserved in sediments or rocks for hundreds of millions or billions of years (Logan et al.,
287 1995; Summons et al., 1999; Brocks et al., 2003). Most studies of microbial lipids have been
288 conducted in sediments or POM (e.g., Zhang et al., 2002, 2003; Pancost and Sinninghe
289 Damsté, 2003; Wakeham et al., 2003) because of the requirement for a large amount of
290 organic material for lipid analysis. Selective accumulation of the refractory lipid-like material
291 in the water column has been demonstrated by the increasing alkanes in the pyrolyzates of
292 sinking POC as depth increased in the Mediterranean Sea (Peulvé et al., 1996). Alkanes from
293 Proterozoic rocks were also identified as biomarkers of heterotrophic bacteria (Logan et al.,
294 1995). These biomarkers might have been derived from MCP activity that contributed to the
295 large DOC pool that may have been 100-1000 times greater than in the modern ocean
296 (Rothman et al., 2003; Ridgwell, 2011; Tziperman et al., 2011). Lipid-like macromolecules in
297 the deep ocean have similar radiocarbon ages and $\delta^{13}\text{C}$ values as the majority (~70%) of the
298 uncharacterized acid-insoluble fraction, indicating that the bulk POC may be compositionally
299 similar to the lipid-like macromolecules (Hwang et al., 2003) (Table 4).

300

301 The greater ages of lipid-like material than carbohydrate- and protein-like substances were
302 also observed in the DOM pool of the open Atlantic and Pacific Oceans (Loh et al., 2004). In
303 particular, the deep-water lipid extract was 13-14 kyr older than the corresponding protein-
304 and carbohydrate-like components in the DOM. This lipid extract was also up to 1 kyr older
305 than the high molecular weight DOM. However, the $\delta^{13}\text{C}$ values of the high-molecular-weight
306 DOC were more similar to the carbohydrate- and protein-like substances than to the lipid
307 extracts, in contrast to the observations of POC (Hwang et al., 2003). This suggests that deep
308 ocean POM and DOM have different origins, with the latter having undergone more extensive
309 recycling (Loh et al., 2004) (Table 4).

310

311 Hwang et al. (2003) and Loh et al. (2004) did not identify specific lipid compositions in either
312 the POM or DOM fractions. However, numerous studies focusing on POM have shown
313 diverse lipid biomarkers from planktonic archaea, bacteria and phytoplankton (Sinninghe
314 Damsté et al., 2002; Wakeham et al., 2003, 2007; Ingalls et al., 2006; Turich et al., 2007;
315 Schubotz et al., 2009; Wei et al., 2011; Schouten et al., 2012). In particular, crenarchaeol was
316 identified as a major glycerol dialkyl glycerol tetraether (GDGT) biomarker for planktonic
317 *Thaumarchaeota* that are present in the global ocean at a total inventory of 10^{28} cells (Karner
318 et al., 2001). GDGTs can be preserved in sediments for millions of years (Kuypers et al., 2001)
319 and can be a significant component of the lipids in the RDOC pool (Table 4). Because
320 *Thaumarchaeota* cell size is small, they are more abundant in the dissolved organic matter
321 fraction (operationally defined as the fraction passing through a ~0.7 μm filter) than the
322 particulate organic fraction (Ingalls et al., 2012). Measurements of the dissolved phases of

323 lipids give total GDGT abundance in the tens of nanograms per liter range (Ingalls et al.,
324 2012); however, once the organisms die, their core lipids may be incorporated into larger
325 particles (0.7- to 60- μ m size fraction) that can be more quickly transported into the deeper
326 ocean and buried in marine sediments (Table 4). The same mechanism may apply to bacterial
327 lipid accumulation in the POM fraction that is preserved in marine sediments. It is unknown,
328 however, how much bacterial or archaeal lipids are **actually** present in the uncharacterized
329 fraction of the RDOM because the uncharacterized RDOM is largely acid-insoluble and
330 cannot be identified by regular gas chromatography- or liquid chromatography mass
331 spectrometry.

332

333 **Carotenoid degradation products.** A recent report by Arakawa et al. (2017) identified
334 carotenoid degradation products (CDP) to be a significant component of the aged DOM using
335 solid phase extraction and comprehensive gas chromatography coupled to mass spectrometry.
336 The CDP are a subset of carboxyl-rich alicyclic molecules (CRAM) and have similar nuclear
337 magnetic resonance spectra as CRAM (Hertkorn et al. 2006). However, the cyclic head groups
338 and branched methyl side chains, with conjugated double bonds, are defining features of
339 isoprenoids characteristic of numerous unique carotenoids that can be produced by plankton
340 (Arakawa et al., 2017). The CDP-rich DOM fraction was depleted in radiocarbon (14 C age >
341 1500 years), indicating a possible long-term accumulation of CDP in the ocean. This was the
342 first direct confirmation of these terpenoids accumulating in refractory DOM and may provide
343 a distinct pathway for a single class of biosynthetic precursors to transform to refractory DOM
344 (Arakawa et al., 2017) (Table 4). **However, this pathway can be either biotic or abiotic and the
345 role that microorganisms play in the transformation of carotenoids to RDOM is unknown.**

346

347 **Characterization of RDOM using proxies**

348

349 **DOC:DON ratio, TDAA (%DOC), and fluorescent DOM.** Microorganisms preferentially
350 utilize nitrogen-containing molecules. Thus the ratio DOC:DON could be used to indicate the
351 bioavailability of DOM (Fellman et al., 2008). Jiao et al. (2010) noted that DOC:DON (molar
352 ratio) increased from 10.0 in surface labile DOM to 17.4 in deep sea refractory DOM
353 (Hopkinson and Vallino, 2000). Similarly, DOC-normalized total dissolved amino acid (TDAA
354 (%DOC)) may be an indicator of DOC lability (Davis and Benner, 2007; Shen et al., 2015).
355 Davis and Benner (2007) observed that TDAA (%DOC) decreased from >20% in labile DOM
356 to 0.7% in deep ocean refractory DOM. Humic-like fluorescent DOM was also thought to be
357 bio-refractory as revealed by its good correlation with apparent oxygen utilization in deep
358 ocean water. This relationship is explained as the production of RDOC from in situ microbial
359 degradation of more labile DOC at the expense of oxygen (Yamashita and Tanoue, 2008;
360 Martínez-Pérez et al., 2017). In addition to fluorescence, absorbance could also be used to
361 infer DOM lability. Specific ultraviolet absorbance has been demonstrated to be a good
362 indicator of aromaticity (Weishaar et al., 2003), which negatively correlates to the lability of
363 DOM (or positively correlates to DOM recalcitrance) (Fellman et al., 2008; Fellman et al.,
364 2009a,b) (Table 4).

365

366 **Coupling between molecular size and radiocarbon age of DOC.** It has been observed that
367 the distribution of total organic carbon in the global ocean is heavily skewed toward the
368 nanometer size range (Benner and Amon, 2015). A hypothesis is that bioavailability of the
369 organic matter decreases with decreasing size and alteration of the organic molecules (Fig. 3
370 insert), meaning that smaller size classes of organic molecules are more slowly remineralized
371 by microorganisms (Amon and Benner, 1996; Benner and Amon, 2015). This has been
372 confirmed by approaches coupling the chemical composition and radiocarbon content of
373 marine organic matter in different size fractions (Loh et al., 2004; Walker et al., 2016a, b). In
374 Loh et al. (2004), seawater from different depths of the central North Pacific and the Sargasso
375 Sea region of the North Atlantic showed that the $\Delta^{14}\text{C}$ values ranged from -5‰ to -434‰ for
376 high-molecular-weight DOM and from -210‰ to -539‰ for low-molecular-weight DOM,
377 with the latter being older than the former by 1650-1850 kyr. The low-molecular-weight DOM
378 was also the most abundant (77-95%) fraction of total DOM, consistent with the overall
379 dominance of RDOM in the ocean (Hansell, 2009). Walker et al. (2016a) examined the C:N
380 ratio and ^{14}C age of organic matter in different size classes from the coastal, surface and deep
381 waters of the Pacific Ocean. In all three environments, larger particles were characterized by
382 young ages and nitrogen enrichment and smaller molecules by older ages and nitrogen
383 depletion. The size-age-composition relationship was also observed in marine sediments with
384 pore water DOC being dominated by low-molecular-weight DOM (Burdige and Gardner,
385 1998).

386

387 In addition to the relationships between size, age and composition, a recent study observed
388 declining concentrations of high-molecular-weight DOM correlated with increasing apparent
389 oxygen utilization along the shallow overturning circulation cell of the Mediterranean Sea
390 (Martínez-Pérez et al., 2017). Decreases in high-molecular-weight DOM accounted for about
391 30% of DOM mineralization. The apparent low-molecular-weight DOM experienced little
392 mineralization, indicating microbes primarily utilized high-molecular-weight molecules,
393 whereas the smaller size classes resisted degradation and were the primary source of
394 recalcitrant DOM in the deep ocean (Martínez-Pérez et al., 2017).

395

396 **Characterization of RDOM composition using FT-ICR MS**

397

398 It is well established that RDOM is composed of less than 10% of common biomolecules such
399 as carbohydrates, amino acids or lipids (see discussion above). Proxies such as the DOC:DON
400 ratio, TDAA (%DOC), fluorescent DOM or the size-age relationship provide insights about
401 the composition and reactivity of DOM, but additional analytical approaches are needed to
402 understand RDOM composition. One approach, Fourier transform ion cyclotron resonance
403 mass spectrometry (FT-ICR MS), has gained popularity in recent years because it identifies
404 thousands of molecular formulae, which can be further analyzed in detail. FT-ICR MS was
405 proposed over 20 years ago (Kujawinski et al., 2002) and has been increasingly applied in the
406 characterization of changes in DOM composition in both terrestrial and marine environments
407 and along environmental gradients (Koch et al., 2005; Hertkorn et al., 2006; Sleighter et al.,
408 2008; Kujawinski et al., 2009; Flerus et al., 2012; Lechtenfeld et al., 2014; D'Andrilli et al.,
409 2015; Medeiros et al., 2017).

410

411 A number of proxies have been developed based on characterization of DOM using FT-ICR
412 MS. CRAM are commonly believed to be refractory and occur as the most abundant
413 components of DOM in the deep ocean. Using the FT-ICR MS technique, Hertkorn et al.
414 (2006) identified over 613 CRAM (Table 4), which can be constrained by the double bond
415 equivalent (DBE) normalized to C (DBE/C = 0.30–0.68), H (DBE/H = 0.20–0.95) or O
416 (DBE/O = 0.77–1.75) within the van Krevelen diagram. These compounds are characterized
417 by abundant carboxyl groups and alicyclic rings commonly found in terpenoids that occur as
418 membrane constituents or secondary metabolites in diverse prokaryotic and eukaryotic
419 organisms (Ourisson et al., 1987). Such findings can be linked to the GC/GC MS analysis of
420 the carotenoid degradation products that can account for 4% of the RDOM component
421 (Arakawa et al., 2017), which agrees with the estimate that CRAM account for 8% of the
422 DOC (Benner, 2002; Hertkorn et al., 2006). Lechtenfeld et al. (2014) further identified 361
423 most stable molecular formulae, called the “island of stability” (IOS) (Table 4) within the
424 CRAM domain (Figure 8 of Lechtenfeld et al., 2014) in the Atlantic and Southern Ocean
425 waters. These molecules are deemed potential indicators of refractory DOM in the Southern
426 Ocean; however, it is unknown whether the same IOS compounds exist in other oceanic
427 environments.

428

429 Another proxy called the degradation index (I_{DEG}) was developed by Flerus et al. (2012) to
430 describe the degradation status of marine DOM analyzed with FT-ICR MS from solid phase
431 extraction (SPE) samples (Table 4). I_{DEG} was calculated using 10 mass peak magnitudes that
432 have either significant linear positive or negative correlation with the $\Delta^{14}\text{C}$ values of the
433 SPE-DOM. The value of I_{DEG} ranges between 0–1 with higher I_{DEG} indicating older age and
434 greater recalcitrance of the DOM. Analysis of seawater at 37° N and 14° W from the eastern
435 Atlantic Ocean showed that I_{DEG} values increased from 0.756 at 400–500 m to 0.808 at
436 4000–5000 m, consistent with the notion that DOM from deeper water is more refractory than
437 shallower water. Likewise, the I_{DEX} was developed based on the SPE DOM samples from the
438 Atlantic Ocean, which needs to be verified in other oceanic regions (Flerus et al., 2012).

439

440 Lastly, Medeiros et al. (2017) identified 184 molecular formulae (Table 4) using FT-ICR MS
441 and used them to indicate riverine inputs in the deep North Atlantic and North Pacific Oceans.
442 These compounds are most enriched in river water and correlated well with known terrigenous
443 tracers in the deep ocean waters, based on which the authors concluded that terrigenous
444 organic matter can be preserved in the deep ocean (Medeiros et al., 2017). This observation is
445 consistent with the deep-ocean distributions of dissolved lignin phenols, biomarkers derived
446 from terrestrial plants (Hernes and Benner, 2006).

447

448 **FT-ICR MS and nuclear magnetic resonance spectroscopy have been used together to trace** the
449 source of deep ocean RDOC from surface primary production. Zhao et al. (2017) observed
450 that cultured picocyanobacteria, *Synechococcus* and *Prochlorococcus*, released fluorescent
451 DOM that underwent similar photo-degradation behavior when compared with deep-ocean
452 fluorescent DOM (Table 4). Ultrahigh-resolution mass spectrometry and nuclear magnetic
453 resonance spectroscopy revealed abundant nitrogen-containing compounds in *Synechococcus*

454 DOM, which may originate from degradation products of the fluorescent phycobilin pigments.
455 Their results suggested that picocyanobacteria are likely to be important sources of marine
456 autochthonous fluorescent DOM, which may accumulate in the deep ocean as RDOC (Zhao et
457 al., 2017).

458

459 Proxies of RDOM in carbon cycle studies must be used with caution given the current
460 constraints in defining the composition and reactivity of RDOC. Jiao et al. (2014) used the
461 term RDOct to describe RDOC compounds maintaining recalcitrance in a specific
462 environmental context and used RDOcc to describe RDOC compounds being inaccessible to
463 microbes due to their extremely low concentrations. It was debated whether low concentration
464 of any DOC compound is the predominant reason for RDOC to remain recalcitrant in the
465 ocean (Arrieta et al., 2015; Jiao et al., 2015). Recent evidence indicates that only a small
466 fraction of RDOC molecules are too dilute for microbial utilization and that environmental
467 conditions, including exposure to photochemical alterations in surface waters and varying
468 microbial communities, are critical for the removal of RDOC from the ocean (Shen and
469 Benner, 2018). The size-age-composition relationship that organic matter size is negatively
470 correlated with radiocarbon age and carbon:nitrogen ratios also supports the dominant role of
471 chemical composition (RDOct) in determining the long persistence of the RDOC pool
472 (Walker et al., 2016a; Amon et al., 2016).

473

474 In addition, if the majority of deep oceanic DOC is RDOcc, i.e., the dilution hypothesis
475 dominates deep oceanic DOC persistence, the $\Delta^{14}\text{C}$ in the deep-ocean calculated from a mass
476 balance model of deep oceanic diluted DOC would be difficult to reconcile with the observed
477 $\Delta^{14}\text{C}$ (4000-6000 years) for deep oceanic DOC (Wilson and Arndt, 2017). **This is because with
478 this observed age constraint, the box model of diluted DOC in the deep ocean would result in
479 either (1) labile DOC comprising a relatively large fraction of bulk DOC but with radiocarbon
480 ages similar to or older than bulk radiocarbon ages or (2) a smaller labile DOC pool with
481 much younger radiocarbon ages; the latter would be most consistent with a variety of other
482 observations (Hansell et al., 2012).**

483

484 **MECHANISMS AND PROCESSES OF RDOC PRODUCTION**

485

486 Studies on the MCP have attempted to address the grand challenges of dissecting the
487 composition of the bulk RDOM and identifying the diverse microbial populations responsible
488 for the fate and complexity of RDOM; both are still largely ‘black boxes’. The research
489 community has reached a consensus that in-depth and integrative characterization of both
490 complex DOM compounds and microbial communities are prerequisites for exploring the
491 relationship between microbial community composition and the processing of DOM (Logue et
492 al., 2016; Moran et al., 2016). Hopes are high to unveil the intimate linkages between the two
493 black boxes by using the advanced technologies provided by both genomics and
494 bioinformatics, and by mass spectrometry capabilities (Kujawinski, 2011; Worden et al., 2015;
495 Moran et al., 2016; Zhang et al., 2016). Here we present some of the latest advances on
496 focused groups of marine organisms as well as community shifts and trophic dynamics
497 associated with RDOM production.

498 **Carbon metabolism of known organisms.** Bacterial metabolism of organic matter is
499 constrained by their physiological capability and biochemical pathways for processing organic
500 molecules. The most studied marine bacteria have been the “eutrophic” *Roseobacter* clade and
501 the “oligotrophic” SAR11 clade of marine alphaproteobacteria (Giovannoni, 2017); both are
502 numerically dominant and functionally important groups of marine bacteria (Dang and Jiao,
503 2014). These clades have distinct patterns of DOC utilization, with *Roseobacter* clade strains
504 mostly taking up carbohydrates and SAR11 preferring nitrogen-containing DOC such as
505 amino acids, which are attributed to different capabilities of ATP binding cassette transporters
506 among these organisms (Jiao and Zheng, 2011; Tang et al., 2012; Dang and Jiao, 2014). Two
507 other studied groups of marine bacteria are the *Gammaproteobacteria* and the
508 *Cytophaga-Flavobacterium-Bacteroides*, which are known to be capable of metabolizing
509 macromolecules through the TonB-dependent transporter proteins (Tang et al., 2012; Dang
510 and Jiao, 2014). Cottrel and Kirchman (2000) observed in estuarine and coastal environments
511 that the *Cytophaga-Flavobacter* cluster showed overrepresentation in the assemblage
512 consuming chitin, *N*-acetylglucosamine, and protein but underrepresentation in the assemblage
513 consuming amino acids. Tang et al. (2017) demonstrated through multi-omics analysis and
514 cultivation experiments that the *Bacteroidetes* strain *Gramella flava* JLT2011 (*Flavobacteria*)
515 has the ability to grow on a wide range of polysaccharides such as xylan and
516 homogalacturonan from pectin, which are operated by different polysaccharide utilization loci
517 (PUL) or PUL-like systems. *Flavobacteria* have also been demonstrated to be a major
518 contributor for the utilization of exopolysaccharides that represent an important source of
519 organic carbon in marine ecosystems (Zhang et al., 2015). However, *Flavobacteria* could not
520 completely utilize exopolysaccharides and fluorescent DOM (e.g., humic acid-like substances)
521 produced during metabolism of exopolysaccharides, which may be refractory and may
522 contribute to the carbon storage in the oceans (Zhang et al., 2015). While these model
523 organisms provide specific knowledge of carbon compounds they metabolize, it is uncertain
524 how these compounds can be identified in natural environments where complex community
525 interactions occur (see below).

526

527 **Carbon metabolism of natural populations.** Studies using individual organisms under
528 laboratory conditions often focus on limited substrates of known compositions. However, the
529 situation is much more complex for natural populations regarding which bacteria may utilize
530 which carbon compounds and whether such compounds in turn may affect specific bacterial
531 community composition (Gómez-Consarnau et al., 2012). Multiple reports demonstrate that
532 specific carbon compounds can select for particular species or groups of organisms under
533 different environmental conditions (Rosselló-Mora et al., 2008). For example,
534 low-molecular-weight molecules (e.g., monomers amino acids, sugars, short chain fatty acids)
535 can be easily transported across cell membranes and may be utilized by most heterotrophic
536 Bacteria or Archaea. However, it has been demonstrated that different low-molecular-weight
537 organic compounds stimulated growth of different types of bacteria, leading to the suggestion
538 that changing composition of the DOC pool can selectively alter the community structure of
539 bacterioplankton (Gómez-Consarnau et al., 2012). This is consistent with observations of the
540 distribution of *Roseobacter* or SAR11 types of organisms selecting for different types of
541 organic substrates (see above). However, it also has been demonstrated that it is the quantity

542 and not the quality of phytoplankton-derived dissolved organic carbon that selects for different
543 types of bacteria in a given range (10-100 μM) of substrate concentrations (Sarmiento et al.,
544 2016).

545

546 The importance of community composition for the fate of DOM has also been shown
547 (Kujawinski et al., 2016; Logue et al., 2016). For example, in incubation experiments using
548 only <1.0- μm microbial populations, DOM composition was dominated by compounds with
549 lipid and peptide characteristics; whereas in incubations with the presence of organisms larger
550 than 1.0- μm , the DOM composition from the culture experiment was nearly identical to that in
551 the natural water, indicating the role of larger microorganisms in constraining DOM
552 composition in the marine environment (Kujawinski et al., 2016). These studies highlight the
553 importance of both microbial community structure and composition or abundance of DOM in
554 the marine system, which should allow distinction between RDOct and RDOcc to better
555 understand the MCP framework (see above).

556

557 The interplay between bacterial community and DOM composition is also examined by
558 comparing particle-attached vs. free-living organisms using genomic tools (Crump et al., 1998;
559 DeLong et al., 1993; Moeseneder et al., 2001; Ghiglione et al., 2007; Eloe et al., 2011; Tarn et
560 al., 2016). Despite our awareness of the different ecological strategies of particle-associated
561 and free-living microbes (e.g., Dang and Lovell, 2016), we know little about the principles
562 behind the phylogenetic differences and life strategies between free-living and
563 particle-attached microbes in the marine environment (Moeseneder et al., 2001; Herndl and
564 Reinthaler, 2013). Particle-associated microbes are capable of utilizing a variety of substrates
565 under nutrient-rich conditions. Free-living heterotrophs, on the other hand, often face a
566 massive pool of refractory dissolved organic molecules under oligotrophic conditions (Lauro
567 et al., 2009; Herndl and Reinthaler, 2013). However, Zhang et al. (2016) observed that the
568 composition of POM was more strongly related to the free-living than to the particle-attached
569 bacterial community, which indicates that POM composition may significantly influence the
570 free-living bacterial community through the release of labile or semilabile organic matter from
571 particles contributing to the bioavailability of dissolved organic carbon (Zhang et al., 2016).
572 The nutritional status of the environment may also affect the difference between
573 particle-attached and free-living populations. For example, in the deep ocean when substrates
574 (ammonia, for example) are scarce, particles provide concentrated life-supporting
575 microenvironments. Microorganisms adapted to a particle-attached lifestyle show the
576 dominance of extracellular hydrolytic enzymes; free-living bacteria, on the other hand, are
577 characterized by hydrolytic enzymes typically bound to the cell surface (Herndl and Reinthaler,
578 2013). In the eutrophic surface ocean and estuaries, substrates or nutrients are abundant and
579 organisms were found to be similar between particle-attached and free-living populations
580 (Dang and Lovell, 2016; Xie et al., 2018).

581

582 **Microbes-DOM interaction at the ecosystem level.** The finding of Kujawinski et al. (2016)
583 that incubation experiments using the whole water community resulted in DOM composition
584 similar to the natural water composition highlights the need to examine the microbes-DOM
585 interaction at the ecosystem scale (Fig. 3). This is convincingly demonstrated by a long term

586 large volume (>100 tons) water column (12 m in depth) incubation, which showed solid
587 evidence of the effective microbial transformation of organic matter from labile to refractory
588 states (Jiao et al., 2018a). A another study provides metagenomic evidence of system level
589 dynamics of microbes-DOM interactions, utilizing the Tara Ocean data that included
590 comprehensive sequences of eukaryotic, prokaryotic and viral lineages from samples collected
591 within the euphotic zone of ocean waters (Guidi et al., 2016). The increased carbon export in
592 this water column was found to correlate not only with bacteria, particularly *Synechococcus*,
593 but also several unicellular eukaryotic microorganisms including three *Rhizaria* lineages and
594 three dinoflagellate lineages that have previously not been believed to play important roles for
595 carbon flux. Also important is the finding of a correlation between the abundance of
596 *Synechococcus* phages and increased carbon export at depth, indicating that phage induced cell
597 lysis promotes particle sinking through enhanced aggregate formation (Suttle, 2007), thus
598 increasing carbon export to the deep ocean (Guidi et al., 2016). The importance of viruses in
599 deeper water is also highlighted in Zhang et al. (2014), who considered viral particles as
600 ‘bottom-up’ agents fueling the microbial loop in the deep ocean.

601

602 Another comprehensive study (McCarren et al., 2010) examined the genomic and
603 transcriptional responses of microbial communities to high-molecular-weight DOM addition
604 in samples from the surface ocean. These authors observed specific resource partitioning of
605 DOM by the bacterial species *Idiomarina* and *Alteromonas* spp. that were most highly
606 represented at the early time points and *Methylophaga* at the final point of the experiment.
607 Their results demonstrated a temporal succession of taxa, metabolic pathways, and chemical
608 transformations associated with high-molecular-weight DOM turnover, suggesting that the
609 cycling of marine DOM may require a coordinated and cooperative effort between different
610 bacterial “specialists”.

611

612 **CASE STUDIES OF INTERACTIONS BETWEEN BCP, ML, AND MCP**

613

614 **Case 1. MCP dynamics associated with upwelling activities**

615

616 Jiao et al. (2014) hypothesized that microbial activity plays a significant role in mediating the
617 source and/or sink of CO₂ in a productive upwelling region. This hypothesis was tested by
618 measuring multiple biogeochemical parameters at two cyclonic-eddy-induced upwelling sites
619 in the western South China Sea, which allowed the formulation of a scenario model of MCP
620 processes under different upwelling conditions.

621

622 In the western South China Sea, satellite altimetric data identified intensification of two
623 cold-core cyclonic eddies, CE1 (decaying) and CE2 (growing), during sample collection (Jiao
624 et al., 2014). In the case of the decaying eddy CE1 (modeling scenario 1, Fig. 4), no
625 phytoplankton bloom occurred and *Prochlorococcus* dominated. The small-sized non-sinking
626 organic particles favored the transfer of energy and organic matter through the ML pathway
627 rather than through the BCP. The enhanced production of labile organic carbon due to
628 upwelled nutrients and phytoplankton growth stimulated microbial respiration (e.g., net
629 community respiration) and decreased POC flux, which suggested that the MCP is the

630 prevailing mechanism for carbon sequestration. In the case of a growing eddy, CE2 (modeling
631 scenario 2, Fig. 4), the rapid growth of phytoplankton caused enhancement of POC downward
632 export flux, where the BCP was the prevailing mechanism for carbon sequestration. Further
633 research is needed to validate these models for general applications.
634

635 【insert Figure 4 here】

636

637 **Case Study 2. Modeling the MCP functions**

638

639 Lu et al. (2018) made an attempt to analyze the MCP-related variables and processes using a
640 coupled physical-ecosystem model that used data collected in the South China Sea and
641 assumed a constant annual production of RDOC of ~ 0.2 Pg C for global oceans (Legendre et
642 al. 2015). They also ran the model with different scenarios simulating rising sea surface
643 temperature and compared the BCP and MCP rates and their relative contributions to carbon
644 sequestration.

645

646 The model coupled a physical model from the operational Taiwan Strait Nowcast\Forecast
647 system (Jiang et al., 2011; Lin et al., 2016) and a biogeochemistry model based on the Carbon,
648 Silicon, Nitrogen Ecosystem module (Xiu and Chai, 2014), which was modified to incorporate
649 an explicit RDOC pool and the MCP processes (Fig. 5). With the constraint of a bulk RDOC
650 concentration of $40 \mu\text{M}$ (Hansell, 2013), and the satellite-based value of primary production,
651 this model estimated the ratio of MCP to BCP (at the depth of 1,000 m) to be 1:6.08 in the
652 South China Sea. The annual production rate of RDOC by the MCP averaged over the whole
653 South China Sea domain was estimated to be $1.55 \text{ mg C m}^{-2} \text{ d}^{-1}$. The BCP, on the other hand,
654 sequestered $9.43 \text{ mg C m}^{-2} \text{ d}^{-1}$.

655

656 【insert Figure 5 here】

657

658 **FUTURE RESEARCH FOCI AND PROSPECTS**

659

660 Jiao et al. (2010) highlighted nine major questions regarding MCP processes, which have been
661 addressed at different levels over the past eight years (Table 3). There is an urgent need to
662 better understand the impacts of global-scale environmental change, including ocean warming
663 and acidification and related deoxygenation and changes in nutrients availability on carbon
664 cycling in the ocean (Jiao et al., 2018b). A central question is how microbial processes
665 contribute to the transformation of organic carbon in the ocean. We advocate three approaches
666 to promote future research in this direction in accordance with Jiao et al. (2018a).

667

668 Firstly we recommend increased investigation of microbiomes in different natural
669 environments, including a much better coverage of the deep ocean. These studies should
670 integrate various omics approaches (i.e., metagenomics, metatranscriptomics, metaproteomics,
671 and metabolomics) at all levels of the microbial community (i.e. virus, bacteria, archaea,

672 phytoplankton, and zooplankton), as well as at selected time-series locations in the coastal and
673 open ocean to identify how the metabolic capacity of the ocean's microbiome responds to
674 spatial and temporal changes in an environmental context (e.g., Moran et al., 2016; Xie et al.,
675 2018).

676

677 The second proposed approach is to strengthen the understanding of the connections between
678 microbial metabolism and the chemical structure of DOC compounds (e.g., Zhang et al., 2016).
679 Bioassays of DOC composition coupled with changes in bacterial communities can now be
680 conducted integrating omics and FT-ICR-MS and NMR technologies, which offers the
681 potential for new insights into mechanisms responsible for the formation of RDOCt and
682 RDOCc. In particular, efforts are needed to fully examine the fate of DOM under different
683 trophic conditions and at the ecosystem level (Guidi et al., 2016; Kujawinski et al., 2016;
684 Osterholz et al., 2016).

685

686 The third proposed approach is to establish and expand long-term incubation studies
687 employing large-scale facilities, such as the existing Aquatron Tower Tank (Dalhousie
688 University, Canada) and the planned Marine Environmental Chamber System (Shandong
689 University, China) under controlled environmental conditions. Using such facilities provides a
690 unique complement to field studies by seeking to mimic ocean-relevant physical, chemical,
691 and biological environmental conditions (e.g. vertical stratification) and their variations for
692 long-term experiments. Such experiments are required to provide unique data and insight for
693 testing hypotheses regarding the effects of global environmental change on the ocean carbon
694 cycle (Legendre et al., 2017; Robinson et al., 2018).

695

696 We also highlight the need to examine the role of planktonic archaea in the carbon cycle.
697 These archaea, such as *Thaumarchaeota*, have been recognized to play an important role in the
698 ocean carbon cycle (Dang and Chen, 2017). Yet, the claim made 7 years ago that “..we are
699 woefully unaware of DOM production (or assimilation) mechanisms in the Archaea”
700 (Kujawinski, 2011) still holds true. The study of archaea is largely hampered by the difficulty
701 of isolating strains from the ocean (e.g., MGII and MGIII). Hence future efforts should include
702 the development of new technologies for enrichment and isolation of these and other
703 organisms, guided by genomic information (Zhang et al., 2015; Xie et al., 2018).

704

705 The MCP has stimulated provocative and constructive discussions and studies on the processes
706 and mechanisms of RDOC formation and preservation (Jiao et al., 2015; Lechtenfeld et al.,
707 2015; Wilson and Arndt, 2017; Zark et al., 2017; Lønborg et al., 2018; Shen and Benner, 2018).
708 Increasing and synergistic efforts will continue to be made to gain further understanding of the
709 ocean carbon cycle through an integration of the concepts of the BCP, ML, VS and MCP,
710 particularly in the context of global ocean circulation (e.g., Shen and Benner, 2018).

711

712

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714

715

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1231 Figure legend

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1233 Figure 1. Cycling of biologically produced organic carbon (POC and DOC) in the ocean and
1234 links between the seafloor and the atmosphere: The BCP, which transports organic matter from
1235 the surface to the interior and floor of the ocean; the MCP, which converts parts of labile
1236 organic carbon into RDOC via microbial activities, mainly by heterotrophic archaea and
1237 bacteria, and associated viruses.

1238

1239 Figure 2. Schematic depiction of the BCP, the ML, and the MCP. The remineralization length
1240 scale in the left part of the figure shows the return of respired CO₂ back to the surface, from
1241 three depth zones (modified from Passow and Carlson, 2012).

1242

1243 Figure 3. Global biogeochemical and ecological models rely on the present understanding of
1244 organismal biology and the interactions between the POM and DOM carbon pools. Modified
1245 from Worden et al. (2015). The inset panel is from Benner and Amon (2015), showing a
1246 decreasing size and reactivity and an increasing complexity and age of organic molecules
1247 along the decomposition pathway. Small dissolved molecules comprise the bulk of RDOC.

1248

1249 Figure 4. Scenario models for the effects of upwelling on ocean carbon uptake/outgassing
1250 dynamics (adopted and modified from Fig. 7 of Jiao et al., 2014). (1) Functioning of the BCP
1251 and the MCP in a non-upwelling region of the ocean. (2) Dominance of the MCP in scenario 1
1252 where the total upward CO₂ flux exceeds downward POC export flux: nutrients are injected
1253 only into the lower layer of the euphotic zone; *Prochlorococcus* is dominant; microbial
1254 respiration is enhanced; CO₂ outgassing exceeds POC export; the MCP is the prevailing
1255 mechanism for carbon sequestration. (3) Dominance of the BCP in scenario 2 where the
1256 downward POC flux exceeds the total upward CO₂ flux: nutrients are injected into the upper
1257 layer of the euphotic zone; diatoms are dominant; POC export exceeds CO₂ outgassing; the
1258 BCP is the prevailing mechanism for carbon sequestration.

1259

1260 Figure 5. Schematic diagram of the MCP module (from Lu et al., 2018). The RDOC in the
1261 model is produced via two bacteria-related pathways: (1) direct exudation by bacteria, and (2)
1262 passive release from viral lysis of microbial cells. The additional POC degradation pathway
1263 (Jiao et al. 2010) is implicitly included by transforming from POC to labile/semi-labile organic
1264 carbon and then to RDOC via aforementioned two pathways (see Lu et al., 2018 for detailed
1265 explanation).

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