



# Ecological roles, climate-driven responses, and critical knowledge gaps of krill in the global ocean

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## Abstract

Human-induced global climate change and other anthropogenic stressors are fundamentally altering our oceans. Understanding the ecological and societal implications of these changes is critical for developing mitigation strategies and conservation measures. However, major components of the marine pelagic ecosystem remain poorly understood. This is true for euphausiids (“krill”), which are a crucial part of marine food webs and play an important role in elemental cycling, including in the biological carbon pump, but for which we know surprisingly little. In this review, we first provide an overview of the ecological and socio-economic value of krill, highlighting their function in marine food webs and biogeochemical cycling. Next, we describe what is currently known regarding the response of krill to climate change and other anthropogenic stressors, focusing on changes in their biogeography, physiology, life history, as well as the impacts of krill fishing and their association with pathogens and parasites. We identify five key gaps in our current knowledge of krill: (1) the effects of krill on food web dynamics and stability, (2) the effects of changing predator and/or prey communities on krill populations, (3) the identification of important krill habitats, (4) the understanding of vertical and horizontal range shifts, and (5) the combined effects of multiple climate change and other anthropogenic stressors on krill. We also highlight the krill species, regions, and habitats that are understudied. Finally, we propose strategies to improve our understanding of this ecologically important taxonomic group, including the sustained funding for time series; implementation of novel research technologies; expanding research on understudied species and regions; and creating a global community of krill researchers.

**Keywords** euphausiids, krill, climate change, anthropogenic stressors, fisheries, carbon pump, ecosystem health

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## Introduction

Krill (Euphausiacea; Dana 1850), hereafter referred to collectively as “krill,” are a fundamental component of global marine ecosystems supporting extensive fish stocks and populations of marine megafauna (Everson 2000). Krill can be found in every ocean, from the poles to the equator and from the surface ocean to the deep sea (Baker et al. 1990, Brinton et al. 2015). A recent functional size spectrum model of zooplankton in marine ecosystems suggests that if krill were removed, fish biomass may decrease by up to 80% (Heneghan et al. 2020). Healthy krill populations are thus critical to both global food security and economies. The ecological importance of krill in global marine ecosystems is not limited to their fundamental role in food webs. Due to their relatively large body size and tendency to form dense aggregations, many krill species are significant mediators of carbon sequestration and are important components in global biogeochemical cycles, including the biological carbon pump and the cycling of other elements necessary for primary production, such as nitrogen and iron (Fortier et al. 1994, Robinson et al. 2010, Schmidt et al. 2011, 2016, Cavan et al. 2019, Franco-Cisterna et al. 2022, 2024).

The combined effects of human-induced global climate change (including ocean warming, ocean acidification, and deoxygenation) and other anthropogenic activities, such as overfishing and pollution, exert a suite of stressors on marine pelagic ecosystems. These stressors are causing cascading effects and reducing energy transfer efficiency through marine food webs with the potential to disrupt fisheries and destabilize global food security through trophic amplification (Heneghan et al. 2023, Atkinson et al. 2024). Warming oceans favor smaller organisms (Campbell et al. 2021) and there is evidence of shifts in the composition of pelagic zooplankton communities from those dominated by larger crustaceans, like large calanoid copepods and krill, to those dominated by smaller taxa, primarily small calanoid and cyclopoid copepods (Heneghan et al. 2023). In this shifting seascape, krill emerge as potential early-warning indicators of broader ecological changes due to their pivotal role in global marine ecosystems and their sensitivity to changes in ocean temperature and chemistry (Díaz-Astudillo et al. 2022a, Werner 2013, Fiechter et al. 2020, Dorman et al. 2023).

Even if CO<sub>2</sub> emissions are reduced in the near future, the global oceans are predicted to warm by ~1.42°C by the end of the century (Kwiatkowski et al. 2020). Marine heatwaves (MHWs) will likely become more frequent, more intense, and of longer duration. For example, recent Coupled Model Intercomparison Project Phase 6 (CMIP6) models predict the Indian Ocean to experience a marine heatwave-state for over 6 months of the year by 2050 (Dalpadado et al. 2024b, Roxy et al. 2024). The current worst-case scenario predicts a 41-fold increase in the number of MHWs from present-day values by the end of the century (Frolicher and Laufkotter 2018, Frolicher et al. 2018). In addition, a general deoxygenation of marine systems is predicted (Bakun 2017, Shepherd et al. 2017, Kwiatkowski et al. 2020), and oxygen minimum zones (OMZs) are expected to expand, shoaling into surface waters (Stramma et al. 2008, 2012). Furthermore, with continued anthropogenic emissions of CO<sub>2</sub>, we can expect to see increased ocean acidification (OA; Doney et al. 2020). Compounding the impacts of human-induced climate change, other anthropogenic stressors like pollution and overfishing affect marine life in a myriad of ways that are overall detrimental to the health of marine organisms. There

is thus a pressing need to understand the implications of human-induced climate change and other anthropogenic stressors on marine ecosystems.

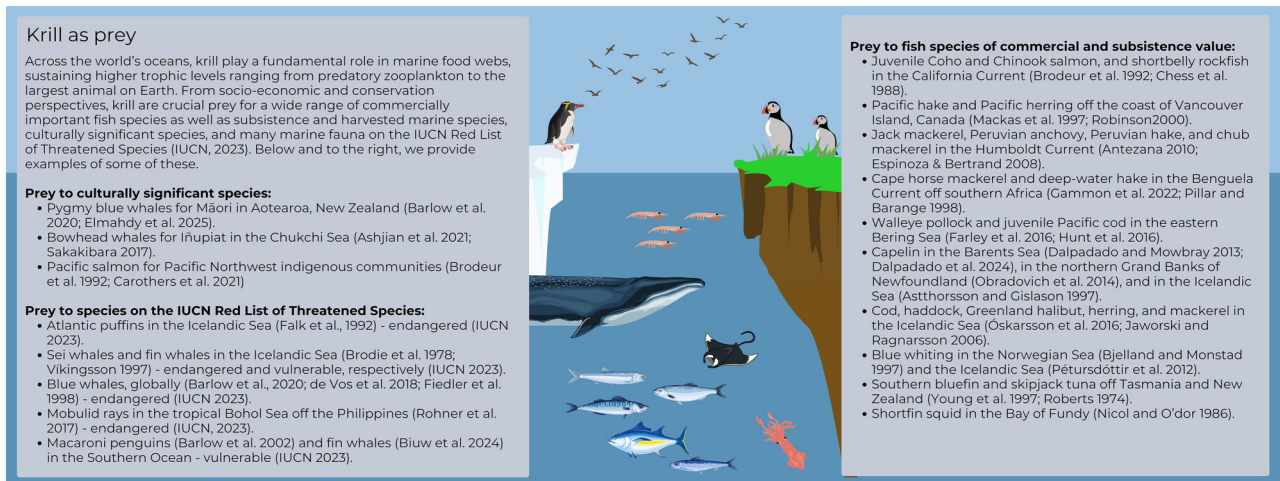
Despite the critical role that krill play in global marine food webs and biogeochemical cycles, and the clear direct impacts climate-induced declines in their populations have on marine ecosystems, very few of the 87 known krill species [86 in Baker et al. (1990), plus the addition of *Hansarsia einarssoni* by Kula-gin et al. (2024)] have been studied in depth, and krill are rarely considered adequately in Earth system models. In the present review, we first highlight the ecological and socio-economic significance of krill in their role as vital trophic intermediaries supporting commercially and culturally significant higher trophic level predators and their contribution to global biogeochemical cycles. Next, we synthesize current understanding of the responses of global krill to human-induced climate change and other anthropogenic stressors, focusing on changes in species biogeography, physiology, life history, the impacts of krill fisheries, and interactions with pathogens and parasites. Using a Delphi-style approach (Linstone and Turoff 1975) to reach academic consensus, we identified the top five gaps in our current knowledge on global krill and provide an overview of the future research directions necessary to address them.

## The importance of krill in global marine ecosystems

### Trophic intermediaries

Krill provide a crucial link between primary producers at the base of the food web and top predators in many ocean habitats from the Antarctic to the Arctic and in between (Dalpadado et al. 2024a, Laws 1977, Everson 2000, Antezana 2010, Savenkoff et al. 2013, Hunt et al. 2016, Trathan and Hill 2016). As consumers, krill exhibit versatile feeding behaviors reflected in the morphological characteristics of their different feeding apparatus, with most species being considered omnivorous (reviewed in Mauchline 1989). The diets and trophic levels of krill are dynamic, varying with life stage, time of day, season, and region (Schmidt 2010, Park et al. 2011, Schmidt and Atkinson 2016, Färber-Lorda and Murcia-Riaño 2021). This versatility in feeding strategies enables krill to optimally use their seasonally or regionally variable food resources. For instance, the diet of dominant krill species differs between the California Current and eastern tropical North Pacific, with individuals of *Thysanoessa spinifera* and *Euphausia pacifica* in California occupying lower trophic positions than *Euphausia diomedea* and *Euphausia distinguenda* in the eastern tropical Pacific (Färber-Lorda and Murcia-Riaño 2021).

The variability in krill diets, and consequently their dynamic trophic position within the food web, alters their quality as prey to higher trophic levels (Färber-Lorda et al. 2009, Färber-Lorda and Mayzaud 2010, Bernard et al. 2022) and modifies the efficiency of energy transfer through marine food webs. Those krill species for which diatoms represent a large component of their diet, like *Euphausia superba* in the Southern Ocean (Stowasser et al. 2012, Cleary et al. 2018) or *Thysanoessa inermis* in the Barents Sea (Dalpadado and Mowbray 2013), provide an efficient energy transfer from primary producers to higher trophic levels. Krill species that occupy comparatively higher trophic positions (i.e. are predomi-



**Figure 1.** The ecological importance of krill as prey in global marine food webs. Depiction of predator guilds (fish, squid, whales, penguins, and seabirds) and the central role of krill in sustaining global marine biodiversity and fisheries productivity.

nantly carnivorous), such as *Meganyctiphanes norvegica*, also play an important role in marine food webs and can be the food of a wide range of fish species, squid, shrimp, whales, and even brittle stars (Simard and Harvey 2010).

The nutritional quality of krill is linked to their diet and physiology and varies by species, season, and life stage (Riquelme-Bugueño et al. 2020a, Hagen et al. 2007, Ju et al. 2009, Färber-Lorda and Mayzaud 2010, Harvey et al. 2012, Hellessey et al. 2018, Fisher et al. 2020, Bernard et al. 2022). Those species and/or life stages that have higher lipid contents tend to be the preferred prey of top predators. For instance, in the California Current, *T. spinifera* has a higher average lipid content than *E. pacifica* (Fisher et al. 2020, Färber-Lorda and Murcia-Riaño 2021) and is preferentially preyed upon by blue whales (Nickels et al. 2018) and humpback whales (Rockwood et al. 2020, Kaplan et al. 2025) despite being less abundant than *E. pacifica*. In Southwest Alaska and Antarctica, humpback whales target adult krill rather than immature individuals that have reduced lipid contents (Szabo 2015, Cade et al. 2022).

Globally, krill are key prey for some of the largest and most lucrative commercially fished species as well as many subsistence fisheries (Fig. 1). In addition, numerous seabirds and marine megafauna, many of which are currently listed on the IUCN Red List of Threatened Species (a comprehensive, science-based information source on the global extinction risk of plants, animals, and fungi) or have cultural significance (either as a subsistence food source or as a spiritual symbol) feed directly on krill (Fig. 1). Although total annual consumption of krill by higher trophic levels has not been assessed globally, it has been estimated at the regional scale. For example, in the Lower St. Lawrence Estuary, Canada, a community of higher trophic level predators ranging from carnivorous macrozooplankton to blue whales consumed an estimated  $14.2 \text{ tonnes km}^{-2} \text{ year}^{-1}$  of *M. norvegica* and  $19.3 \text{ tonnes km}^{-2} \text{ year}^{-1}$  of *Thysanoessa raschii*, which equated to an average of 35% of the krill biomass estimated between 2008 and 2010 for that region (Savenkoff et al. 2013). In Icelandic waters, dominated by *M. norvegica*, *Thysanoessa longicaudata*, and *T. inermis* (Astthorsson and Gislason 1997), annual consumption of krill by cetaceans was estimated to be 3.4 million tonnes (Sigur-

jónsson and Víkingsson 1997), while seabirds consumed  $\sim 34,000$  tonnes each year (Lilliendahl and Solmundsson 1997). In the eastern Bering Sea, where *T. raschii* and *T. inermis* are the main krill species, walleye pollock alone consumed between 17% and 29% of the summer krill production each year between 1999 and 2009 (Hunt et al. 2016). Off the coast of Vancouver Island, Canada, the fish community (including Coho and Chinook salmon, dogfish, sablefish, herring, hake and Pacific cod) consumed an annual average of 268,400 metric tonnes of krill—which primarily consist of *E. pacifica* and *T. spinifera* (Evans et al. 2021)—between 1985 and 1989 (Robinson 2000). Marine mammals and seabirds in the Scotia Sea, Southern Ocean, consume at least 55 million tonnes of *E. superba* annually (Trathan and Hill 2016). Recent estimates of annual summer *E. superba* consumption by fin whales alone amount to  $\sim 8$  million tonnes (Biuw et al. 2024). Consequently, the health of global krill populations is vital to sustaining thriving top predator populations with the potential for serious ramifications for commercial fisheries and conservation efforts when krill abundances decrease.

## Role in biogeochemical cycles

Krill play an important role in the biological carbon pump, the suite of biologically driven processes that are responsible for transferring photosynthetic carbon from surface waters to the ocean interior (Sarmiento and Gruber 2013, Boyd et al. 2019). Although traditionally described in terms of the gravitational pump—the sinking of particulate organic carbon in the form of faecal pellets and detritus—the biological carbon pump is now recognized as a composite of several interconnected processes, including the mesopelagic migrant pump, which involves active transport by migrating organisms and the release of dissolved and suspended carbon at depth (Boyd et al. 2019). Krill play a key role in both the gravitational and migrant pumps.

The role of krill in the gravitational pump is particularly evident in the Southern Ocean, where the large individual body size and extraordinary abundances of *E. superba* result in the dominance of krill-derived particles (faecal pellets) at depth, especially during spring and summer (Gleiber et al. 2012, Cavan et al. 2015, Belcher

et al. 2019, Manno et al. 2020, 2024). Furthermore, regular moulting throughout the krill life cycle produces a continuous flux of carbon-rich exuviae, recently shown to contribute as much to carbon export in the Scotia Sea as faecal pellets (Manno et al. 2020). Another potentially important but poorly constrained pathway is the passive export of carbon associated with the descent and mortality of krill embryos (Thorpe et al. 2019), which sink to depth during development but for which direct estimates of mortality-driven carbon flux are lacking. Sinking carcasses add a further important pathway for carbon flux. In waters off southwest Greenland, the euphausiids *T. raschii* and *M. norvegica* were found to sink at rates of 1500–3000 m d<sup>-1</sup>, with minimal carbon loss (<10%) during descent (Franco-Cisterna et al. 2022). Such rapid export enhances the efficiency of sequestration and supports benthic productivity by supplying organic carbon and nutrients (Franco-Cisterna et al. 2022). As an important counterpoint, swimming behavior of *E. pacifica* off Southern California was shown to fragment marine snow aggregates into smaller particles, potentially altering sinking rates and attenuating levels of carbon export (Dilling and Alldredge 2000).

In addition to these gravitational pathways, diel vertical migration (DVM; Hays 2003) may provide an active transport route for carbon to depth in krill species that exhibit this behavior. By feeding near the surface and respiring, excreting, and egesting at depth, vertically migrating krill actively inject organic carbon into the mesopelagic zone (Steinberg et al. 2000, Darnis et al. 2017, Boyd et al. 2019). However, not all DVM is equal. In krill, DVM includes (i) the vertical migration of epipelagic species between the epipelagic and mesopelagic (e.g. Taki 2008, Haraldsson and Siegel 2014, Sogawa et al. 2016), (ii) the vertical migration of epipelagic species within the epipelagic (e.g. Taki 2008, Conroy et al. 2020), and (iii) the vertical migration of mesopelagic species between the mesopelagic and epipelagic (e.g. Steinberg et al. 2000). These migrations are typically upward at night, but the reverse has also been observed (Meyer et al. 2017). There is often a seasonal component to DVM, particularly in regions with strongly seasonal food supply (Werner and Buchholz 2013, Haraldsson and Siegel 2014, Sogawa et al. 2016, Darnis et al. 2017, Smith et al. 2025).

Through feeding, egestion, and excretion in surface waters, krill release essential elements—such as iron (Tovar-Sanchez et al. 2007, Schmidt et al. 2016), ammonium (Atkinson and Whitehouse 2000, Steinberg et al. 2002, Ikeda 2012, Lehette et al. 2012, Kiko et al. 2016, Darnis et al. 2017), and dissolved organic carbon (Steinberg et al. 2000, Ruiz-Halpern et al. 2011)—that are required for primary and microbial production. Furthermore, by migrating to the seafloor, ingesting lithogenic particles with high iron content, and returning to surface waters, krill redistribute this essential trace metal throughout the water column, locally enhancing primary productivity, particularly in regions with low iron concentrations such as the Southern Ocean where *E. superba* has been shown to transport iron to the surface waters (Schmidt et al. 2016, Cavan et al. 2019).

## Response of krill to a changing ocean world

The global oceans are changing rapidly with significant ramifications for marine ecosystem functioning and productivity (Richardson 2008, Brierley and Kingsford 2009, Poloczanska et al. 2013,

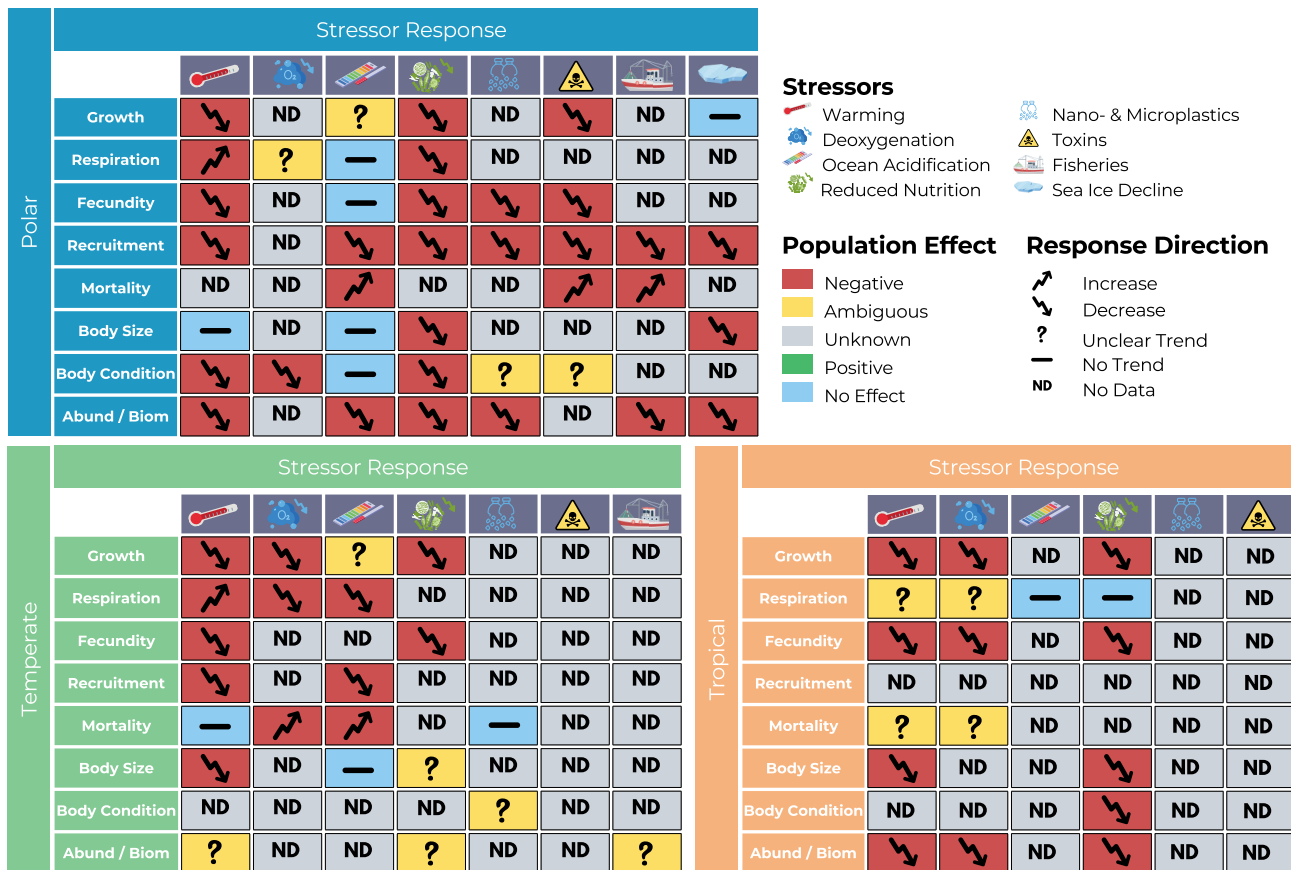
2016). Warming associated with long-term human-induced climate change is causing a general poleward shift in isotherms, resulting in the contraction of biogeographic habitat for some species and expansion for others. Changes in large-scale ocean circulation patterns, frontal positions, and upwelling are modifying transport, advection, and nutrient delivery (Bakun et al. 2015, Hunt et al. 2016, Wilson et al. 2016, Ashjian et al. 2021, Lévy et al. 2024). In addition to long-term ocean warming, many parts of the world ocean are also experiencing acute MHWs, the ecological effects of which are not yet fully understood. Ocean warming has also reduced ventilation (through increased stratification) and solubility of the global oceans (de Boer et al. 2007, Shepherd et al. 2017). These processes, in combination with altered ocean circulation, increased biological oxygen demand, and greater eutrophication near the coast, have resulted in the deoxygenation of several oceanic regions (Bakun 2017, Kwiatkowski et al. 2020) with profound negative consequences for marine life (Kwiatkowski et al. 2020, Morée et al. 2023). While this is problematic at a global scale, deoxygenation has a particularly strong negative impact on marine organisms living above OMZs, which are a predominant feature in the Eastern North and South Pacific, the Arabian Sea, and the Bay of Bengal (Dalpadado et al. 2024b, Fernández-Álamo and Färber-Lorda 2006, Paulmier and Ruiz-Pino 2009). In addition to warming and deoxygenation, rapidly increasing atmospheric CO<sub>2</sub> concentrations have led to an increase in dissolved CO<sub>2</sub> and inorganic carbon concentrations in seawater, resulting in a reduction in pH and changes to the acid-base chemistry of marine environments (Orr et al. 2005, Fabry et al. 2008, Doney et al. 2009).

Human-induced climate change is, justifiably, a dominant theme in marine research, but we would be remiss to neglect the review of other anthropogenic stressors, including pollution (nano- and microplastics, heavy metal contamination, anthropogenic noise, and eutrophication) and fishing. These additional stressors can act to compound those caused by climate change, further exacerbating the negative effects on marine ecosystems and organisms. Considering the ecological and socio-economic importance of krill, it is vital to establish an overview of how our changing oceans will affect their populations. In this section, we discuss the various implications of changing oceanic conditions for global krill populations, including changes in their biogeography, physiology, life history, the impacts of krill fishing, and interactions with pathogens and parasites.

## Changes in biomass and species distribution

The biogeography of krill has been comprehensively described by Brinton et al. (2015) in the online resource “Euphausiids of the World Ocean.” This work, that includes all known species at the time of publication (86 species), provides a valuable baseline from which to quantify climate driven shifts in biogeography. Understanding the changing biogeography of krill species globally is complex and requires time-series spanning multiple decades to disentangle (Ducklow et al. 2022). Such programs are rare for krill but, where they have been maintained, they shed light on the varied responses of different krill species to climate-driven stressors (e.g. those outlined in Ducklow et al. 2022). These stressors can be categorized as (i) press (long-term, directional change, e.g. long-term warming), (ii) pulse (intense, short-term deviations from the





**Figure 2.** Region-specific stressor–response matrices for euphausiids in tropical, temperate, and polar ecosystems. Each matrix summarizes observed or inferred responses of euphausiids to key environmental and anthropogenic stressors—warming, deoxygenation, ocean acidification, declining food quality and quantity, pollution, fisheries, and (for polar regions only) sea-ice loss. Responses are shown for eight biological or population-level metrics: growth, respiration, fecundity, recruitment, mortality, body size, body condition, and abundance/biomass. Icons indicate the direction of response, while box colors represent the overall consequence for population health. Fisheries targeting euphausiids are absent in tropical systems; thus, only the temperate and polar matrices include a fisheries column. References used to inform the matrices are provided in the Supplementary materials.

ifornia, *N. simplex* abundances increase during EP El Niños, but show the opposite during CP events (Parés-Escobar et al. 2018).

In the Southern Ocean, ENSO and the SAM are among the dominant large-scale climate modes affecting krill biogeography. In the SW Atlantic sector, positive SAM (warmer, windier conditions) resulted in a decrease in *E. superba* biomass and a southward contraction of their population centre (Atkinson et al. 2019). At the northern Antarctic Peninsula, La Niña (colder, more productive conditions) was associated with increased abundances of *E. superba* (Loeb and Santora 2015). By contrast, along the western Antarctic Peninsula, *E. superba* showed no directional long-term trend (Steinberg et al. 2015). Responses of *E. superba* to ENSO and SAM likely arise through their effects on sea ice, upper-ocean stratification or mixed-layer structure, and primary productivity, which together influence reproductive development and recruitment (Saba et al. 2014, Steinke et al. 2021, 2024, Atkinson et al. 2022) (Fig. 2). Responses of other Southern Ocean krill species are also variable, and appear to reflect differences in thermal tolerances, habitat affinity, and position along latitudinal gradients. At the northern Antarctic Peninsula, *Euphausia crystallorophias*, *Euphausia frigida*, and larval *Thysanoessa macrura* increased in abundance following a shift to more La Niña and Niño-neutral conditions, whereas the sub-Antarctic *Euphausia triacantha* tended

to increase during warmer El Niño conditions (Loeb and Santora 2015). Along the western Antarctic Peninsula, *E. crystallorophias* was more abundant under higher-ice conditions and was best explained by SAM and MEI variability rather than by a simple long-term warming response (Steinberg et al. 2015).

The responses of krill to climate modes may also reflect cumulative integration of environmental variability over organismal life history. Di Lorenzo and Ohman (2013) showed that variability in the abundance of *N. simplex* was better explained by an integrated PDO signal than by the PDO itself, and referred to this as the Double Integration Hypothesis—a concept under which atmospheric variability is first integrated by the ocean and then by population dynamics over the species' lifespan. While double-integration explained variability in abundances of *N. simplex*, it could not explain patterns in abundance of *E. pacifica*, which exhibited a more direct, single-integration response. The double-integration framework has important implications for interpreting lags, apparent regime shifts, and climate-change signals in krill time series.

## Effects on physiology

The additive effects of increased seawater temperatures with low  $pO_2$  and elevated  $pCO_2$  concentrations, in addition to pollution (in-



smaller body sizes (Gómez-Gutiérrez et al. 2012, Atkinson et al. 2019, Robertson and Bjorkstedt 2020, Lagos et al. 2021, Killeen et al. 2022, Shaw and Fisher 2026) (Fig. 2). Ocean warming, deoxygenation, acidification, and exposure to nanoplastics may also disproportionately affect early life stages of krill, reducing recruitment to the population (Kawaguchi et al. 2013, Meyer et al. 2017, Rowlands et al. 2021a, b) (Fig. 2). For example, in the Humboldt Current System, larval krill abundances were lower during El Niño compared to La Niña conditions (Aronés et al. 2019, Díaz-Astudillo et al. 2022b). The decrease in abundance of krill in the California Current System during anomalously warm years was likely the result of lower spring recruitment and adult survival throughout the year due, in part, to increased metabolic requirements and decreased food availability (Marinovic et al. 2002). Reduced food availability in response to warming has an adverse effect on early life stages in high latitude regions too, where changes in the timing of the spring phytoplankton bloom may result in a temporal mismatch between krill larvae and their phytoplankton food (Silva et al. 2014).

## Krill fisheries

Commercial exploitation of krill began in the 1970s, spurred by projections that global catches could reach 50–100 million tonnes annually for human consumption (Gulland 1972, Sprague and Arnold 1972). In practice, present-day catches remain far lower, marginally over 0.5 million tonnes per year and dominated by the Southern Ocean fishery for *E. superba* (~0.5 Mt), alongside smaller fisheries targeting *E. pacifica* in Japanese (~20 000 t) and Canadian (<250 t) waters (Fisheries and Oceans Canada 2022, Wakamatsu et al. 2022, CCAMLR 2024) (Fig. 2). These harvests represent only a small fraction of total stock biomass—less than 1% in the case of *E. superba* (Hill et al. 2020)—yet the growing demand for fishmeal and omega-3 supplements has driven substantial corporate investment, with companies collectively spending hundreds of millions of dollars on modern krill-fishing fleets (Cappell et al. 2022). Exploratory harvesting has also been considered in other regions, including the Icelandic Sea (Gíslason et al. 2021, Sigurðardóttir and Gíslason 2021). Krill populations are naturally variable in abundance and distribution, influencing both fishery yields and prey availability for predators (Wakamatsu et al. 2022). In the North Pacific, *E. pacifica* fisheries compete with major krill-feeding fish such as Pacific cod and Alaska pollock, whose annual krill consumption equals 15%–64% of the commercial catch (Yamamura et al. 1998).

In the Southern Ocean, *E. superba* supports the world's largest krill fishery, managed under the Commission for the Conservation of Antarctic Marine Living Resources (CCAMLR). Current catch limits theoretically allow a 22-fold increase in legal harvest (Nicol and Foster 2016), although quotas have remained unchanged for more than a decade. Recent studies highlight the tension between industrial expansion and ecosystem needs: catch concentrations around the Antarctic Peninsula are intensifying local depletion and predator competition (Freer et al. 2025, Trathan et al. 2025). Meyer et al. (2025) propose a “krill-stock hypothesis” framework that integrates climate variability, spatial connectivity, and recruitment dynamics into adaptive management. Together, these findings indicate that, as commercial demand for krill products continues to grow, governance reforms and more spatially respon-

sive regulation will be essential to sustain both the fishery and the broader Southern Ocean ecosystem.

## Interactions with pathogens and parasites

Impacts of pathogens and parasites on krill individuals can range from mild to severe including sluggish, erratic swimming, loss of body transparency, black spots on exoskeleton, slow or desynchronized heartbeat, opaque and/or nonfunctional chromatophores, abnormal digestive gland and intestinal function prolonged fasting, malnutrition, weight loss, irregular growth and moulting, impairment in reproduction, castration, or death (Gómez-Gutiérrez 2003, Gómez-Gutiérrez and Morales-Ávila 2016, Gómez-Gutiérrez et al. 2017, Cleary et al. 2024, 2025). Krill may also serve as intermediate hosts, transmitting parasites on to vertebrate predators (Hays et al. 1998, Cleary et al. 2019, Flores-Cascante et al. 2019). Little is currently known on how infections impact the susceptibility of krill to external stressors, nor on the impact of external stressors on susceptibility of krill to pathogens and parasites. Some linkages would appear mechanistically likely. For example, the common krill protistan parasites Gregarines (Apicomplexa) infest gut tissue and reduce digestion efficiency; related gregarine parasites in insects increase susceptibility to negative impacts of food limitation, which may also occur in krill, though this phenomenon is not yet documented (Takahashi et al. 2011, Randall et al. 2013). Some have speculated that the euphausiid trait of continuous moulting throughout the life-cycle, which is unusual in crustaceans, may be an adaptation to reduce parasitic infections (Tarling and Cuzin-Roudy 2008). It has been suggested that pathogenic and parasitic organisms globally may have larger impacts on host individuals and populations in a warmer world because smaller organisms typically respond metabolically more rapidly to increases in temperature (Byers 2021). But shifting distributions of both krill and parasites, as well as their intertwined coevolutionary histories and thermal niches makes this difficult to predict (Studer et al. 2010, Danovaro et al. 2011).

## Knowledge gaps

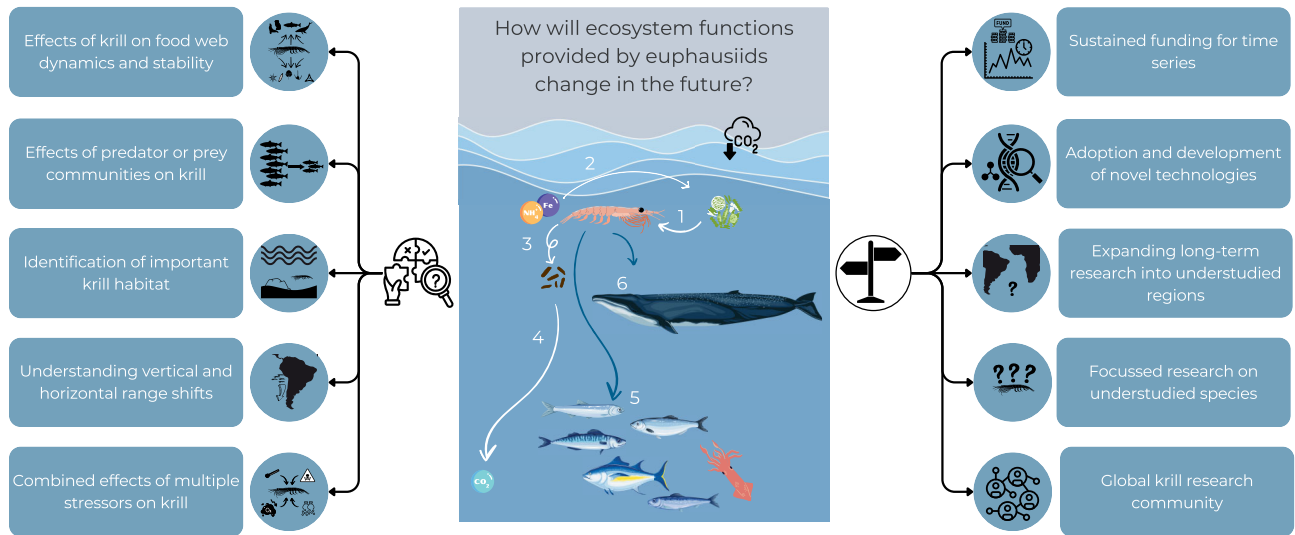
Due to the challenges with studying krill, there are numerous gaps in our knowledge about this group, globally. We conducted a Delphi-style assessment among the coauthors to gain consensus on the top five gaps in our knowledge on krill (Fig. 3; see the Supplementary material for details on this assessment approach). In addition, we have identified understudied species, regions, and habitats.

### Top five knowledge gaps

#### *The role of krill in controlling trophic dynamics and affecting the stability of marine food webs*

The term *wasp-waist* has been used to define ecosystems where a single mid-trophic level species exerts top-down control on lower trophic levels and bottom-up control on higher trophic levels (Curry et al. 2000, Atkinson et al. 2014). This attribute has been ascribed to *E. superba* in the Southern Ocean, and it implies that the removal of the species would have severe consequences for all levels of the food web, potentially disrupting its stability. Given the

## Knowledge Gaps and Future Directions



**Figure 3.** Knowledge gaps and future directions for understanding euphausiid ecology and ecosystem roles. Central panel illustrates key ecosystem functions mediated by krill, including nutrient recycling (1), grazing and carbon export (2–4), and their role as prey for fish and megafauna (5–6). Surrounding panels summarize priority research needs. On the left, major knowledge gaps, on the right, strategic actions needed to address these gaps.

role of other krill species as critical consumers and prey in other regions of the global ocean, there is potential for krill in other regions to play similarly crucial roles in their respective ecosystems, influencing both primary productivity and predator populations. Consequently, shifts in krill populations due to environmental changes or exploitation could destabilize these ecosystems, underscoring the need for region-specific studies on their ecological functions and vulnerabilities.

### Impacts of changing food webs on global krill

Surface ocean stratification has increased globally due to rising temperature, preventing vertical mixing and the resupply of nutrients from deeper water (Li et al. 2020). Such conditions favor smaller phytoplankton taxa that lack essential polyunsaturated fatty acids (Marañón 2009, Schmidt et al. 2020, 2024) and thereby reduce the efficiency of trophic transfer from primary producers to zooplankton consumers, including krill (Atkinson et al. 2024). Simultaneously, the recovery of marine megafauna, including fur seals, humpback and fin whales (Baines et al. 2021, Forcada et al. 2023), from human exploitation has led to an increase in predation on krill. Additionally, anthropogenic activities such as overfishing and eutrophication have resulted in localized increases in gelatinous zooplankton (Richardson et al. 2009, Brodeur et al. 2018), with increased predation on krill larvae (Suchman et al. 2008) and the potential to outcompete krill for food resources (Bernard et al. 2012, O’Loughlin et al. 2020). Understanding the ramifications of food web shifts on krill is essential for predicting future krill population changes.

### Identification of key habitats for global krill

Krill exist in a highly advective environment and their capacity to control their spatial distribution continuously develops over their life cycle. Consequently, krill occupy a range of marine habitats that may vary by life stage or behavior. This means that hotspots

for spawning or aggregations of early developmental stages can often be considerably displaced from those of the adults (Perry et al. 2019). For most krill species, we do not know which habitats are vital at which stages. For instance, while we know that sea ice is important for the winter survival of larval and juvenile *E. superba* in Antarctica (Meyer 2012, Meyer et al. 2017, Bernard et al. 2018), we do not know if this is true for other polar krill. Spawning and recruitment locations are unknown for almost all krill species, yet this information is critical to understand population dynamics and predict population fluctuations, which will in turn inform marine food web and biogeochemical models, fisheries management, and conservation efforts. Different life-stages may be exposed to contrasting environments and rates of change. Identifying which krill life cycle stages are most vulnerable is a fundamental requirement in krill ecology.

### Habitat compression and distribution shifts in response to climate change

Ocean warming and expanding OMZs impose major stresses on krill, and their response in terms of distributional shifts was identified as a major knowledge gap. Studies of krill and other plankton species have shown that ranges cannot simply be predicted from water temperature alone, with highly nonlinear and sometimes counter-intuitive distribution shifts observed (Chivers et al. 2017, Edwards et al. 2021, Atkinson et al. 2022). Likewise, expansion of the horizontal and vertical extent of OMZs may compress the vertical habitat (Fernández-Álamo and Färber-Lorda 2006, Färber-Lorda and Färber-Data 2023) with species showing variable responses in terms of acclimation and adaptation. Understanding the mechanisms behind these redistributions is crucial for distinguishing long-term distribution shifts from short-term population responses to climatic drivers like ENSO or SAM or extreme events like MHWs.

## Response of krill to multiple human-induced stressors

Exposure of krill to multiple human-induced stressors may adversely impact their robustness to environmental change, as combined stressors can lower biological thresholds (Hunter et al. 2024). For example, simultaneous exposure to OA and nanoplastics reduces krill larval development more than the exposure to each stressor individually (Rowlands et al. 2021a). Studying single drivers can therefore produce misleading conclusions about individual or population responses to multiple stressors (Bopp et al. 2013), which can vary by region, habitat, species, and life stage (Hobday and Lough 2011). Recent studies increasingly use multifactorial designs to address this complexity (Saba et al. 2021, Rowlands et al. 2021b). However, interactions between stressors—whether additive, synergistic, or antagonistic—make it difficult to extrapolate from single-stressor responses, often producing conflicting results (Riebesell and Gattuso 2015). Addressing this complexity demands interdisciplinary work involving modeling, empirical, and experimental efforts.

## Understudied species, regions, and habitats

A review of Web of Science searches (conducted on 10 November 2025) for each of the Euphausiacea species revealed that the number of publications per year on krill has increased over time since 1965, dominated almost entirely by research on polar krill species (Fig. 4A). Since 1965, there have been nearly 4000 publications on polar krill species, ~1000 on temperate species, and <500 on tropical species. The number of publications on polar krill have increased at a rate of ~2.4 per year, compared to 0.5 per year for temperate krill and 0.05 per year for tropical species. Research on polar krill thus far exceeds both the total number of publications and the rate of increase in publication numbers for either temperate or tropical species (Fig. 4A).

Of the 87 known species, research and the scientific literature focus predominantly on the Antarctic krill, *E. superba* (2708 publication counts since 1965, Fig. 4A and B). Species for which at least 100 articles have been published include *M. norvegica* (592 counts), *E. pacifica* (379 counts), *T. inermis* (240 counts), *T. raschii* (137 counts), *E. crystallorophias* (134 counts), and *T. macrura* (127 counts). *Euphausia superba* and *E. pacifica* are both the focus of fisheries, and *E. pacifica*, *M. norvegica*, *T. inermis*, and *T. raschii* are major prey items of commercially fished species in their respective regions. *Euphausia crystallorophias* and *T. macrura* are relatively abundant in the Southern Ocean, where substantial research effort on krill has focused. These seven species occur primarily in northern hemisphere temperate-polar waters, or in the Southern Ocean (Fig. 4C). Notably, none of these species occur in the tropical ocean or in the temperate southern hemisphere.

Fish consumption in the global south is an important source of nutrition (Allegretti and Hicks 2023, Maulu et al. 2024). Commercially important and subsistence fish in those regions depend on krill species that we currently lack even the basic biological information on. For instance, *E. eximia* (23 publication counts) and *E. mucronata* (37 counts) in the Humboldt Current off the coasts of Chile and Peru, *Nyctiphanes capensis* (25 counts) and *Euphausia lucens* (71 counts) in the Benguela Current off the west coast of South Africa and Namibia, and *N. australis* (78 counts) in the Tas-

man Sea and waters off New Zealand all are vital prey for commercially fished species.

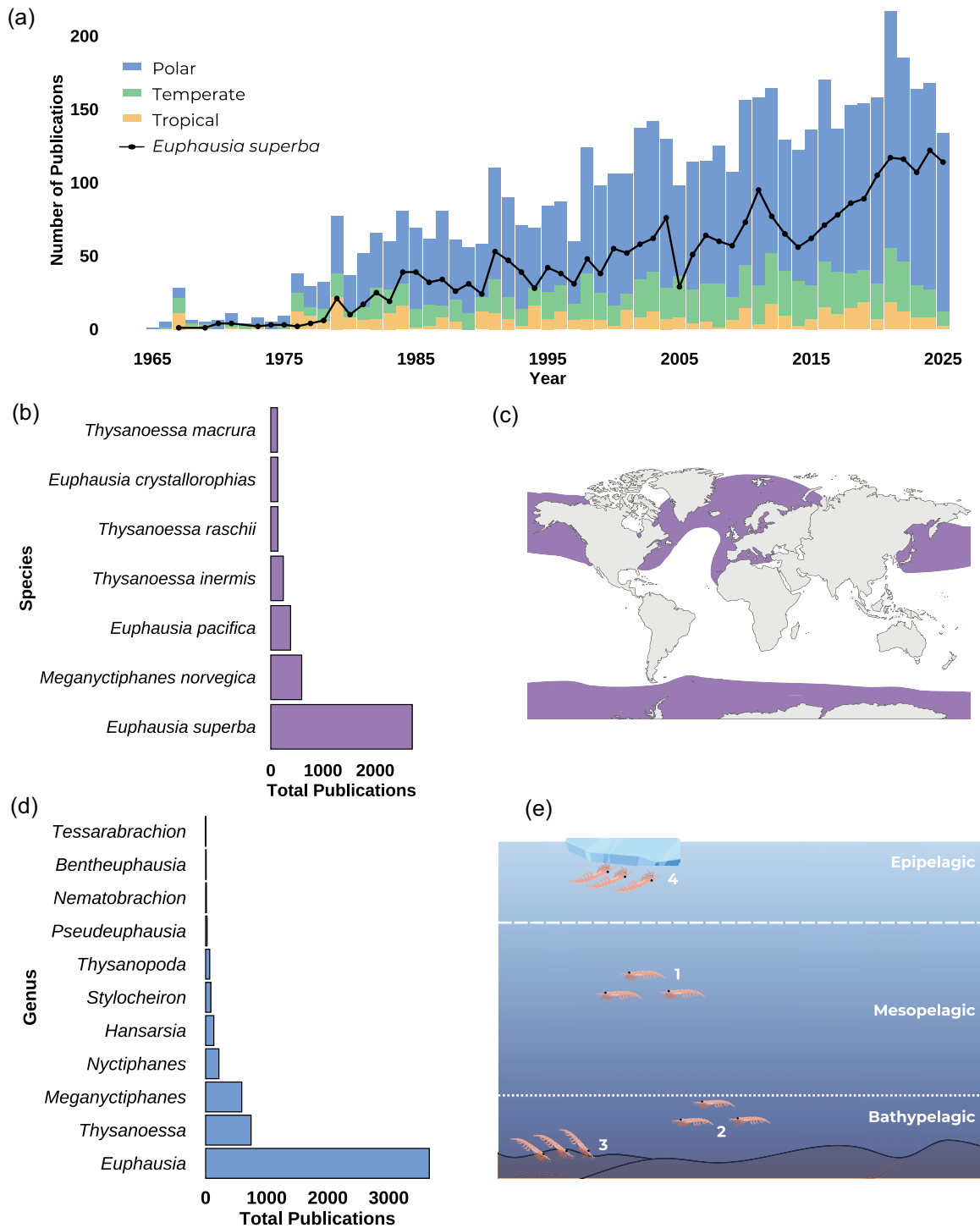
Of the 11 genera of Euphausiacea, the *Euphausia* genus has received by far the most attention, roughly five times the publication counts than the next most mentioned genus, *Thysanoessa*, and two orders of magnitude more than the least mentioned genera: *Nematobrachion*, *Bentheuphausia*, and *Tessarabrachion* (Fig. 4D). Species in these latter genera occur predominantly in the meso- and bathypelagic realms, which are severely understudied krill habitat (Fig. 4E). The role of meso- and bathypelagic krill in carbon sequestration and open-ocean fisheries is likely immense (Irigoien et al. 2014, Siegel et al. 2023, Burd 2024), but inadequately quantified. Another krill habitat that remains poorly studied is the sea floor (Fig. 4E). Anthropogenic activities on the sea floor (dredging, mining, benthic trawling, and drilling) may negatively impact krill populations that rely on this habitat, but at present we do not know how important sea floor interactions are for krill. In the polar regions, sea ice is also an important habitat for some krill species (Fig. 4E), but global warming is reducing this habitat and the consequences of this to polar krill species have not yet been fully resolved.

## Future directions

To better anticipate future changes in ecosystem functions provided by krill that influence the health and resilience of our oceans (e.g. food to higher trophic levels, carbon sequestration, and biogeochemical cycling), research aimed at addressing the top five knowledge gaps and increasing the representation of understudied species, regions, and habitats is critical (Fig. 3). This includes improved long-term monitoring, implementing the use of new technologies, enhanced global networking, and expanding research on understudied regions and species.

### Improved long-term monitoring of global krill abundances, species composition, and distribution patterns

Long-term monitoring is critical to understand the response of marine ecosystems to a changing climate but faces a wide range of challenges (Benway et al. 2019). These include the need to maintain sufficiently long time series for statistical power to reveal climate change responses, the fact that most research funding is shorter term (1–5 years), that traditional ship-based sampling is suffering funding cuts worldwide (Mackas and Beaugrand 2010, Mackas et al. 2012, Vucetich et al. 2020, Ratnarajah et al. 2023), and that taxonomic expertise requires training and is a skill that develops over years. Notwithstanding these challenges, a few ocean-basin scale surveys are on-going, a notable example being the CPR survey. However, most long-term surveys are at regional scales, run by national monitoring programs that recognize the importance of krill in commercially exploited food webs. Good examples include work in the California Current (Lilly and Ohman 2021, Phillips et al. 2022, Dorman et al. 2023), off Iceland (Silva et al. 2014), and the Peru–Chile upwelling (Díaz-Astudillo et al. 2022a). Long-term monitoring of krill could be expanded (both temporally and regionally) by employing nontraditional survey platforms such as ships of opportunity (as is often used by the CPR program) and krill fishing vessels (Meyer et al. 2020).



**Figure 4.** Global research effort and distribution patterns for euphausiids (krill) based on a Web of Science survey conducted on 10 November 2025. (A) Annual number of krill-related publications from 1965 to 2025, categorized by habitat domain (Polar, Temperate, and Tropical), with the black line showing annual number of publications for *Euphausia superba*. (B) Total publications for the seven most frequently studied (at least 100 publications) euphausiid species. (C) Geographic ranges of those seven species (*E. superba*, *E. crystallorophias*, *T. macrura* in Antarctica, *M. norvegica* in the North Atlantic, and *E. pacifica*, *T. inermis*, and *T. raschii* in the North Pacific) illustrating the uneven global coverage of euphausiid research, with most effort concentrated in high-latitude and North Atlantic–North Pacific systems. (D) Total publications by genus, highlighting the taxonomic bias toward *Euphausia* and *Thysanoessa*. (E) Conceptual schematic showing major euphausiid habitats where research remains limited: (1) mesopelagic, (2) bathypelagic, (3) seafloor, and (4) under-ice environments.





- Antezana T. *Euphausia mucronata*: a keystone herbivore and prey of the Humboldt Current System. *Deep Sea Res Part II* 2010;**57**:652–62. <https://doi.org/10.1016/j.dsr2.2009.10.014>
- Aronés K, Grados D, Ayón P *et al.* Spatio-temporal trends in zooplankton biomass in the northern Humboldt current system off Peru from 1961–2012. *Deep Sea Res Part II* 2019;**169**–170:104656.
- Ashjian CJ, Okkonen SR, Campbell RG *et al.* Lingering Chukchi Sea sea ice and Chukchi Sea mean winds influence population age structure of euphausiids (krill) found in the bowhead whale feeding hotspot near Pt. Barrow, Alaska. *PLoS One* 2021;**16**:e0254418. <https://doi.org/10.1371/journal.pone.0254418>
- Asthorsson OS, Gislason A. On the food of capelin in the subarctic waters north of Iceland. *Sarsia* 1997;**82**:81–6. <https://doi.org/10.1080/00364827.1997.10413641>
- Atkinson A, Hill S, Pakhomov E *et al.* KRILLBASE: a database of Antarctic krill and salp densities in the Southern Ocean, 1926 to 2016. *Earth Sys Sci Data* 2017;**9**:193–210.
- Atkinson A, Hill SL, Barange M *et al.* Sardine cycles, krill declines, and locust plagues: revisiting “wasp-waist” food webs. *Trends Ecol Evol* 2014;**29**:309–16. <https://doi.org/10.1016/j.tree.2014.03.011>
- Atkinson A, Hill SL, Pakhomov EA *et al.* Krill (*Euphausia superba*) distribution contracts southward during rapid regional warming. *Nat Clim Change* 2019;**9**:142–7. <https://doi.org/10.1038/s41558-018-0370-z>
- Atkinson A, Hill SL, Reiss CS *et al.* Stepping stones towards Antarctica: switch to southern spawning grounds explains an abrupt range shift in krill. *Global Change Biol* 2022;**28**:1359–75. <https://doi.org/10.1111/gcb.16009>
- Atkinson A, Rossberg AG, Gaedke U *et al.* Steeper size spectra with decreasing phytoplankton biomass indicate strong trophic amplification and future fish declines. *Nat Commun* 2024;**15**:381. <https://doi.org/10.1038/s41467-023-44406-5>
- Atkinson A, Shreeve RS, Hirst AG *et al.* Natural growth rates in Antarctic krill (*Euphausia superba*): II. Predictive models based on food, temperature, body length, sex, and maturity stage. *Limnol Oceanogr* 2006;**51**:973–87. <https://doi.org/10.4319/lo.2006.51.2.0973>
- Atkinson A, Whitehouse MJ. Ammonium excretion by Antarctic krill *Euphausia superba* at South Georgia. *Limnol Oceanogr* 2000;**45**:55–63. <https://doi.org/10.4319/lo.2000.45.1.0055>
- Baines M, Kelly N, Reichelt M *et al.* Population abundance of recovering humpback whales *Megaptera novaeangliae* and other baleen whales in the Scotia Arc, South Atlantic. *Mar Ecol Progr Ser* 2021;**676**:77–94. <https://doi.org/10.3354/meps13849>
- Baker A de C, Boden BP, Brinton E. *Practical Guide to the Euphausiids of the World*. New York: Natural History Museum Publications, 1990.
- Bakun A. Climate change and ocean deoxygenation within intensified surface-driven upwelling circulations. *Philos Trans R Soc A* 2017;**375**:20160327. <https://doi.org/10.1098/rsta.2016.0327>
- Bakun A, Black BA, Bograd SJ *et al.* Anticipated effects of climate change on coastal upwelling ecosystems. *Curr Clim Change Rep* 2015;**1**:85–93. <https://doi.org/10.1007/s40641-015-0008-4>
- Barlow DR, Bernard KS, Escobar-Flores P *et al.* Links in the trophic chain: modeling functional relationships between in situ oceanography, krill, and blue whale distribution under different oceanographic regimes. *Mar Ecol Progr Ser* 2020;**642**:207–25. <https://doi.org/10.3354/meps13339>
- Barlow KE, Boyd IL, Croxall JP *et al.* Are penguins and seals in competition for Antarctic krill at South Georgia? *Mar Biol* 2002;**140**:205–13.
- Basedow SL, McKee D, Lefering I *et al.* Remote sensing of zooplankton swarms. *Sci Rep* 2019;**9**:686. <https://doi.org/10.1038/s41598-018-37129-x>
- Bax NJ, Miloslavich P, Muller-Karger FE *et al.* A response to scientific and societal needs for marine biological observations. *Front Mar Sci* 2019;**6**:395. <https://doi.org/10.3389/fmars.2019.00395>
- Belcher A, Biermann L, Fielding S *et al.* Experimental determination of reflectance spectra of Antarctic krill (*Euphausia superba*) in the Scotia Sea. *Antarct Sci* 2021;**33**:402–14. <https://doi.org/10.1017/S0954102021000262>
- Belcher A, Henson SA, Manno C *et al.* Krill faecal pellets drive hidden pulses of particulate organic carbon in the marginal ice zone. *Nat Commun* 2019;**10**:889. <https://doi.org/10.1038/s41467-019-08847-1>
- Benedetti F, Vogt M, Elizondo UH *et al.* Major restructuring of marine plankton assemblages under global warming. *Nat Commun* 2021;**12**:5226. <https://doi.org/10.1038/s41467-021-25385-x>
- Benoit-Bird KJ, Patrick Welch T, Waluk CM *et al.* Equipping an underwater glider with a new echosounder to explore ocean ecosystems. *Limnol Oceanogr Methods* 2018;**16**:734–49.
- Benson A, Murray T, Canonico G *et al.* Data management and interactive visualizations for the evolving. *Mar Biodivers Obser Netw Oceanogr* 2021;**34**:131–41.
- Benway HM, Lorenzoni L, White AE *et al.* Ocean time series observations of changing marine ecosystems: an era of integration, synthesis, and societal applications. *Front Mar Sci* 2019;**6**:393. <https://doi.org/10.3389/fmars.2019.00393>
- Bergami E, Manno C, Cappello S *et al.* Nanoplastics affect moulting and faecal pellet sinking in Antarctic krill (*Euphausia superba*) juveniles. *Environ Int* 2020;**143**:105999. <https://doi.org/10.1016/j.envint.2020.105999>
- Bernard KS, Bernardino AF, Buck KN *et al.* Hydrothermal vent habitat use by gravid Antarctic krill reveals a hidden life-history strategy. *Commun Biol*
- Bernard KS, Gunther LA, Mahaffey SH *et al.* The contribution of ice algae to the winter energy budget of juvenile Antarctic krill in years with contrasting sea ice conditions. *ICES J Mar Sci* 2018;**76**:206–16. <https://doi.org/10.1093/icesjms/fsy145>
- Bernard KS, Steinberg DK, Schofield OME. Summertime grazing impact of the dominant macrozooplankton off the Western Antarctic Peninsula. *Deep Sea Res Part I* 2012;**62**:111–22. <https://doi.org/10.1016/j.dsr.2011.12.015>
- Bernard KS, Steinke KB, Fontana JM. Winter condition, physiology, and growth potential of juvenile Antarctic krill. *Front Mar Sci* 2022;**9**:990853. <https://doi.org/10.3389/fmars.2022.990853>
- Biuw M, Lindstrøm U, Jackson JA *et al.* Estimated summer abundance and krill consumption of fin whales throughout the Scotia Sea during the 2018/2019 summer season. *Sci Rep* 2024;**14**:7493. <https://doi.org/10.1038/s41598-024-57378-3>
- Bjelland O, Monstad T. Blue whiting in the Norwegian Sea, spring and summer 1995 and 1996. *ICES*. 1997

- Bocher P, Caurant F, Miramand P *et al.* Influence of the diet on the bioaccumulation of heavy metals in zooplankton-eating petrels at Kerguelen archipelago, Southern Indian Ocean. *Polar Biol* 2003;**26**:759–67. <https://doi.org/10.1007/s00300-003-0552-6>
- Bograd SJ, Castro CG, Di Lorenzo E *et al.* Oxygen declines and the shoaling of the hypoxic boundary in the California Current. *Geophys Res Lett* 2008;**35**:L12607. <https://doi.org/10.1029/2008GL034185>
- Bopp L, Resplandy L, Orr JC *et al.* Multiple stressors of ocean ecosystems in the 21st century: projections with CMIP5 models. *Biogeosciences* 2013;**10**:6225–45. <https://doi.org/10.5194/bg-10-6225-2013>
- Boyd PW, Claustre H, Levy M *et al.* Multi-faceted particle pumps drive carbon sequestration in the ocean. *Nature* 2019;**568**:327–35. <https://doi.org/10.1038/s41586-019-1098-2>
- Brierley AS, Kingsford MJ. Impacts of climate change on marine organisms and ecosystems. *Curr Biol* 2009;**19**:R602–14. <https://doi.org/10.1016/j.cub.2009.05.046>
- Brierley AS, Ward P, Watkins JL *et al.* Acoustic discrimination of Southern Ocean zooplankton. *Deep Sea Res II* 1998;**45**:1155–73.
- Brinton E, Ohman MD, Townsend AW *et al.* 2015 Euphausiids of the World Ocean. <https://euphausiids.linnaeus.naturalis.nl/> (1 April 2026, date last accessed).
- Brinton E, Townsend A. Decadal variability in abundances of the dominant euphausiid species in southern sectors of the California Current. *Deep Sea Res Part II* 2003;**50**:2449–72. [https://doi.org/10.1016/S0967-0645\(03\)00126-7](https://doi.org/10.1016/S0967-0645(03)00126-7)
- Brodeur R, Perry I, Boldt J *et al.* An unusual gelatinous plankton event in the NE Pacific: the Great Pyrosome Bloom of 2017. *PICES Press* 2018;**26**:22–7.
- Brodeur RD, Francis RC, Percy WG. Food consumption of juvenile Coho (*Oncorhynchus kisutch*) and Chinook Salmon (*O. tshawytscha*) on the continental shelf off Washington and Oregon. *Can J Fish Aquat Sci* 1992;**49**:1670–85. <https://doi.org/10.1139/f92-186>
- Brodeur RD, Hunsicker ME, Hann A *et al.* Effects of warming ocean conditions on feeding ecology of small pelagic fishes in a coastal upwelling ecosystem: a shift to gelatinous food sources. *Mar Ecol Progr Ser* 2019;**617-618**:149–63. <https://doi.org/10.3354/meps12497>
- Brodie P, Sameoto D, Sheldon R. Population densities of euphausiids off Nova Scotia as indicated by net samples, whale stomach contents, and sonar. *Limnol Oceanogr* 1978;**23**:1264–7. <https://doi.org/10.4319/lo.1978.23.6.1264>
- Brown M, Kawaguchi S, Candy S *et al.* Temperature effects on the growth and maturation of Antarctic krill (*Euphausia superba*). *Deep Sea Res Part II* 2010;**57**:672–82. <https://doi.org/10.1016/j.dsr2.2009.10.016>
- Buchholz F. 1985 Molt and growth in euphausiids. In: Siegfried WR, Condy PR, Laws RM (eds), *Antarctic Nutrient Cycles and Food Webs*. Berlin, Heidelberg: Springer, 339–45. <https://doi.org/10.1007/978-3-642-82275-9>
- Bucklin A, Questel JM, Batta-Lona PG *et al.* Population genetic diversity and structure of the euphausiids *Thysanoessa inermis* and *T. raschii* in the Arctic Ocean: inferences from COI barcodes. *Mar Biodivers* 2023;**53**:70. <https://doi.org/10.1007/s12526-023-01371-y>
- Burd AB. Modeling the vertical flux of organic carbon in the global ocean. *Ann Rev Mar Sci* 2024;**16**:135–61. <https://doi.org/10.1146/annurev-marine-022123-102516>
- Butterley A, Kawaguchi S, Bach LT *et al.* The production of ‘food boluses’ by Antarctic krill and implications for organic matter transport. *Biol Lett* 2025;**21**:20250312. <https://doi.org/10.1098/rsbl.2025.0312>
- Byers JE. Marine parasites and disease in the era of global climate change. *Ann Rev Mar Sci* 2021;**13**:397–420. <https://doi.org/10.1146/annurev-marine-031920-100429>
- Cade DE, Kahane-Rapport SR, Wallis B *et al.* Evidence for size-selective predation by Antarctic humpback whales. *Front Mar Sci* 2022;**9**:747788. <https://doi.org/10.3389/fmars.2022.747788>
- Campbell MD, Schoeman DS, Venables W *et al.* Testing Bergmann’s rule in marine copepods. *Ecography* 2021;**44**:1283–95. <https://doi.org/10.1111/ecog.05545>
- Cappell R, MacFadyen G, Constable A. Research funding and economic aspects of the Antarctic krill fishery. *Mar Policy* 2022;**143**:105200. <https://doi.org/10.1016/j.marpol.2022.105200>
- Carothers C, Black J, Langdon SJ *et al.* Indigenous peoples and salmon stewardship: a critical relationship. *Ecol Soc* 2021;**26**:art16. <https://doi.org/10.5751/ES-11972-260116>
- Carreiro AR, Ramos JA, Mata VA *et al.* DNA metabarcoding to assess prey overlap between tuna and seabirds in the Eastern tropical Atlantic: implications for an ecosystem-based management. *Mar Environ Res* 2023;**187**:105955. <https://doi.org/10.1016/j.marenvres.2023.105955>
- Cavan EL, Laurenceau-Cornec EC, Bressac M *et al.* Exploring the ecology of the mesopelagic biological pump. *Prog Oceanogr* 2019;**176**:102125. <https://doi.org/10.1016/j.pocean.2019.102125>
- Cavan EL, Le Moigne FAC, Poulton AJ *et al.* Attenuation of particulate organic carbon flux in the Scotia Sea, Southern Ocean, is controlled by zooplankton fecal pellets. *Geophys Res Lett* 2015;**42**:821–30. <https://doi.org/10.1002/2014GL062744>
- Cavan EL, Mackay N, Hill SL *et al.* Antarctic krill sequester similar amounts of carbon to key coastal blue carbon habitats. *Nat Commun* 2024;**15**:7842. <https://doi.org/10.1038/s41467-024-52135-6>
- Cavole L, Demko A, Diner R *et al.* Biological impacts of the 2013–2015 warm-water anomaly in the Northeast Pacific: winners, losers, and the future. *Oceanography* 2016;**29**. <https://doi.org/10.5670/oceanog.2016.32>
- CCAMLR. Fishery report 2023: *e euphausia superba* in Area 48. Tasmania, 2024.
- Checkley DM, Barth JA. Patterns and processes in the California Current System. *Prog Oceanogr* 2009;**83**:49–64. <https://doi.org/10.1016/j.pocean.2009.07.028>
- Chelton DB, Bernal PA, McGowan JA. Large-scale interannual physical and biological interaction in the California Current. *J Mar Res* 1982;**40**:1095–125.
- Chen I-C, Hill JK, Ohlemüller R *et al.* Rapid range shifts of species associated with high levels of climate warming. *Science* 2011;**333**:1024–6. <https://doi.org/10.1126/science.1206432>
- Chess JR, Smith SE, Fischer PC. Trophic relationships of the shortbelly rockfish, *Sebastes jordani*, off Central California. 1988;**XXIX**.

- Chivers WJ, Walne AW, Hays GC. Mismatch between marine plankton range movements and the velocity of climate change. *Nat Commun* 2017;**8**:14434. <https://doi.org/10.1038/ncomms14434>
- Choquet M, Lenner F, Cocco A *et al.* Comparative population transcriptomics provide new insight into the evolutionary history and adaptive potential of world ocean krill. *Mol Biol Evol* 2023;**40**:msad225. <https://doi.org/10.1093/molbev/msad225>
- Chu D, Wiebe PH. Measurements of sound-speed and density contrasts of zooplankton in Antarctic waters. *ICES J Mar Sci* 2005;**62**:818–31. <https://doi.org/10.1016/j.icesjms.2004.12.020>
- Clarke LJ, Suter L, King R *et al.* Bacterial epibiont communities of panmictic Antarctic krill are spatially structured. *Mol Ecol* 2021;**30**:1042–52. <https://doi.org/10.1111/mec.15771>
- Cleary AC, Casas MC, Durbin EG *et al.* Parasites in Antarctic krill guts inferred from DNA sequences. *Antarct Sci* 2019;**31**:16–22. <https://doi.org/10.1017/S0954102018000469>
- Cleary AC, Durbin EG, Casas MC. Feeding by Antarctic krill *Euphausia superba* in the West Antarctic Peninsula: differences between fjords and open waters. *Mar Ecol Progr Ser* 2018;**595**:39–54. <https://doi.org/10.3354/meps12568>
- Cleary AC, Kawaguchi S, King R *et al.* Antarctic Krill with parasites grow slower than uninfected peers. *Mar Biol* 2025;**172**:113. <https://doi.org/10.1007/s00227-025-04673-w>
- Cleary AC, Perry F, Tarling GA. Influence of parasites on spawning success and egg quality in Antarctic krill *Euphausia superba*. *Mar Ecol Progr Ser* 2024;**739**:1–14. <https://doi.org/10.3354/meps14617>
- Conroy JA, Steinberg DK, Thibodeau PS *et al.* Zooplankton diel vertical migration during Antarctic summer. *Deep Sea Res Part I* 2020;**162**:103324. <https://doi.org/10.1016/j.dsr.2020.103324>
- Cooper HL, Potts DC, Paytan A. Metabolic responses of the North Pacific krill, *Euphausia pacifica*, to short- and long-term pCO<sub>2</sub> exposure. *Mar Biol* 2016;**163**:207. <https://doi.org/10.1007/s00227-016-2982-z>
- Curry P, Bakun A, Crawford RJM *et al.* Small pelagics in upwelling systems: patterns of interaction and structural changes in “wasp-waist” ecosystems. *ICES J Mar Sci* 2000;**57**:603–18. <https://doi.org/10.1006/jmsc.2000.0712>
- Dalpadado P, Mowbray F. Comparative analysis of feeding ecology of capelin from two shelf ecosystems, off Newfoundland and in the Barents Sea. *Prog Oceanogr* 2013;**114**:97–105. <https://doi.org/10.1016/j.pocean.2013.05.007>
- Dalpadado P, Prokopchuk IP, Bogstad B *et al.* Zooplankton link climate to capelin and polar cod in the Barents Sea. *Prog Oceanogr* 2024a;**226**:103302. <https://doi.org/10.1016/j.pocean.2024.103302>
- Dalpadado P, Roxy MK, Arrigo KR *et al.* Rapid climate change alters the environment and biological production of the Indian Ocean. *Sci Total Environ* 2024b;**906**:167342. <https://doi.org/10.1016/j.scitotenv.2023.167342>
- Dana JD. Synopsis generum crustaceorum ordinis “Schizopoda. *Am J Sci Arts* 1850;**9**:129–33.
- Danovaro R, Corinaldesi C, Dell’anno A *et al.* Marine viruses and global climate change. *FEMS Microbiol Rev* 2011;**35**:993–1034. <https://doi.org/10.1111/j.1574-6976.2010.00258.x>
- Darnis G, Hobbs L, Geoffroy M *et al.* From polar night to midnight sun: diel vertical migration, metabolism and biogeochemical role of zooplankton in a high Arctic fjord (Kongsfjorden, Svalbard). *Limnol Oceanogr* 2017;**62**:1586–605. <https://doi.org/10.1002/lno.10519>
- Dawson AL, Kawaguchi S, King CK *et al.* Turning microplastics into nanoplastics through digestive fragmentation by Antarctic krill. *Nat Commun* 2018;**9**:1001. <https://doi.org/10.1038/s41467-018-03465-9>
- de Boer AM, Sigman DM, Toggweiler JR *et al.* Effect of global ocean temperature change on deep ocean ventilation. *Paleoceanography* 2007;**22**:PA2210. <https://doi.org/10.1029/2005PA001242>
- Desforges J-PW, Galbraith M, Dangerfield N *et al.* Widespread distribution of microplastics in subsurface seawater in the NE Pacific Ocean. *Mar Pollut Bull* 2014;**79**:94–9. <https://doi.org/10.1016/j.marpolbul.2013.12.035>
- Deutsch C, Brix H, Ito T *et al.* Climate-forced variability of ocean hypoxia. *Science* 2011;**333**:336–9. <https://doi.org/10.1126/science.1202422>
- de Vos A, Faux CE, Marthick J *et al.* New determination of prey and parasite species for northern Indian Ocean blue whales. *Front Mar Sci* 2018;**5**:104. <https://doi.org/10.3389/fmars.2018.00104>
- Díaz-Astudillo M, Riquelme-Bugueño R, Bernard KS *et al.* Disentangling species-specific krill responses to local oceanography and predator’s biomass: the case of the Humboldt krill and the Peruvian anchovy. *Front Mar Sci* 2022a;**9**:3389.
- Díaz-Astudillo M, Riquelme-Bugueño R, Saldías GS *et al.* Mesoscale and climate environmental variability drive krill community changes in the Humboldt Current System. *J Mar Syst* 2024;**245**:103998. <https://doi.org/10.1016/j.jmarsys.2024.103998>
- Díaz-Astudillo M, Saldías GS, Letelier J *et al.* Spatial and interannual variability in the distribution of euphausiid life stages in the permanent upwelling system off northern Chile. *ICES J Mar Sci* 2022b;**79**:61–75. <https://doi.org/10.1093/icesjms/fsab241>
- Dilling L, Alldredge AL. Fragmentation of marine snow by swimming macrozooplankton: a new process impacting carbon cycling in the sea. *Deep Sea Res Part I* 2000;**47**:1227–45. [https://doi.org/10.1016/S0967-0637\(99\)00105-3](https://doi.org/10.1016/S0967-0637(99)00105-3)
- Di Lorenzo E, Ohman MD. A double-integration hypothesis to explain ocean ecosystem response to climate forcing. *Proc Natl Acad Sci USA* 2013;**110**:2496–9. <https://doi.org/10.1073/pnas.1218022110>
- Doney SC, Busch DS, Cooley SR *et al.* The impacts of ocean acidification on marine ecosystems and reliant human communities. *Annu Rev Environ Resour* 2020;**45**:83–112. <https://doi.org/10.1146/annurev-environ-012320-083019>
- Doney SC, Fabry VJ, Feely RA *et al.* Ocean acidification: the other CO<sub>2</sub> problem. *Ann Rev Mar Sci* 2009;**1**:169–92. <https://doi.org/10.1146/annurev.marine.010908.163834>
- Dong Y, Qi G, Feng C *et al.* Genetic diversity and structure of *Euphausia superba* in the South Shetland Islands using the mitochondrial ND6 gene. *Crustaceana* 2019;**92**:1295–309. <https://doi.org/10.1163/15685403-00003950>
- Dorman JG, Sydeman WJ, Thompson SA *et al.* Environmental variability and krill abundance in the central California current: implications for ecosystem monitoring. *Front Mar Sci* 2023;**10**:1099482. <https://doi.org/10.3389/fmars.2023.1099482>

- Ducklow H, Cimino M, Dunton KH *et al.* Marine pelagic ecosystem responses to climate variability and change. *Bioscience* 2022;**72**:827–50. <https://doi.org/10.1093/biosci/biac050>
- Edwards M, Helaouet P, Goberville E *et al.* North Atlantic warming over six decades drives decreases in krill abundance with no associated range shift. *Commun Biol* 2021;**4**:644. <https://doi.org/10.1038/s42003-021-02159-1>
- Elmahdy YM, Orams M, Lück M *et al.* Indigenous communities and marine mammal tourism management: incorporating the perspectives of the indigenous Māori people of Aotearoa/New Zealand. *Front Sustain Tour* 2025;**4**:1510025. <https://doi.org/10.3389/frsut.2025.1510025>
- Eriksen E, Skjoldal HR, Dolgov AV *et al.* The Barents Sea euphausiids: methodological aspects of monitoring and estimation of abundance and biomass. *ICES J Mar Sci* 2016;**73**:1533–44. <http://doi.org/10.1093/icesjms/fsw022>
- Espinoza P, Bertrand A. Revisiting Peruvian anchovy (*Engraulis ringens*) trophodynamics provides a new vision of the Humboldt Current system. *Prog Oceanogr* 2008;**79**:215–27. <https://doi.org/10.1016/j.pocean.2008.10.022>
- Evans R, English PA, Anderson SC *et al.* Factors affecting the seasonal distribution and biomass of *E. pacifica* and *T. spinifera* along the Pacific coast of Canada: a spatiotemporal modelling approach. *PLoS One* 2021;**16**:e0249818. <https://doi.org/10.1371/journal.pone.0249818>
- Evans R, English PA, Gauthier S *et al.* Quantifying the effects of extreme events and oceanographic variability on the spatiotemporal biomass and distribution of two key euphausiid prey species. *Front Mar Sci* 2023;**10**:1031485. <https://doi.org/10.3389/fmars.2023.1031485>
- Evans R, Lea MA, Hindell MA *et al.* Significant shifts in coastal zooplankton populations through the 2015/16 Tasman Sea marine heatwave. *Estuar Coast Shelf Sci* 2020;**235**:106538. <https://doi.org/10.1016/j.ecss.2019.106538>
- Everson I. *Krill Biology, Ecology and Fisheries*. Oxford: Blackwell Science Ltd, 2000. <https://doi.org/10.1002/9780470999493>
- Fabry VJ, Seibel BA, Feely RA *et al.* Impacts of ocean acidification on marine fauna and ecosystem processes. *ICES J Mar Sci* 2008;**65**:414–32. <https://doi.org/10.1093/icesjms/fsn048>
- Falk K, Jensen J-K, Kampp K. Winter Diet of Atlantic Puffins (*Fratercula arctica*) in the Northeast Atlantic. *Colonial Waterbirds* 1992;**15**:230–5. <https://doi.org/10.2307/1521457>
- Färber-Lorda J, Färber-Data B. Autumn vertical distribution of zooplankton in the oxygen minimum zone of the Eastern Tropical North Pacific. *Mar Environ Res* 2023;**190**:106116. <https://doi.org/10.1016/j.marenvres.2023.106116>
- Färber-Lorda J, Gaudy R, Mayzaud P. Elemental composition, biochemical composition and caloric value of Antarctic krill: implications in energetics and carbon balances. *J Mar Syst* 2009;**78**:518–24. <https://doi.org/10.1016/j.jmarsys.2008.12.021>
- Färber-Lorda J, Mayzaud P. Morphology and total lipids in *Thysanoessa macrura* from the southern part of the Indian Ocean during summer: spatial and sex differences. *Deep Sea Res Part II* 2010;**57**:565–71. <https://doi.org/10.1016/j.dsr2.2009.11.001>
- Färber-Lorda J, Murcia-Riáño M. Trophic position and partitioning differences between euphausiid species in the Southern California Current and eastern tropical North Pacific using stable isotope ratios ( $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ ), elemental composition, and lipid contents. *Mar Biol* 2021;**168**:121. <https://doi.org/10.1007/s00227-021-03928-6>
- Farley EV, Heintz RA, Andrews AG *et al.* Size, diet, and condition of age-0 Pacific cod (*Gadus macrocephalus*) during warm and cool climate states in the eastern Bering Sea. *Deep Sea Res Part II* 2016;**134**:247–54. <https://doi.org/10.1016/j.dsr2.2014.12.011>
- Fernández-Álamo MA, Färber-Lorda J. Zooplankton and the oceanography of the eastern tropical Pacific: a review. *Prog Oceanogr* 2006;**69**:318–59. <https://doi.org/10.1016/j.pocean.2006.03.003>
- Fernández-Barba M, Belyaev O, Huertas IE *et al.* Marine heatwaves in a shifting Southern Ocean induce dynamical changes in primary production. *Commun Earth Environ* 2024;**5**:404. <https://doi.org/10.1038/s43247-024-01553-x>
- Fiechter J, Santora JA, Chavez F *et al.* Krill hotspot formation and phenology in the California Current Ecosystem. *Geophys Res Lett* 2020;**47**:e2020GL088039. <https://doi.org/10.1029/2020GL088039>
- Fiedler PC, Reilly SB, Hewitt RP *et al.* Blue whale habitat and prey in the California Channel Islands. *Deep Sea Res Part II* 1998;**45**:1781–801. [https://doi.org/10.1016/S0967-0645\(98\)80017-9](https://doi.org/10.1016/S0967-0645(98)80017-9)
- Fielding S, Watkins JL, Trathan PN *et al.* Interannual variability in Antarctic krill (*Euphausia superba*) density at South Georgia, Southern Ocean: 1997–2013. *ICES J Mar Sci* 2014;**71**:2578–88. <https://doi.org/10.1093/icesjms/fsu104>
- Fisher JL, Menkel J, Copeman L *et al.* Comparison of condition metrics and lipid content between *Euphausia pacifica* and *Thysanoessa spinifera* in the northern California Current, USA. *Prog Oceanogr* 2020;**188**:102417. <https://doi.org/10.1016/j.pocean.2020.102417>
- Fisheries and Oceans Canada. 2023–2027. Euphausiid Integrated Fisheries Management Plan January 1, 2023 to December 31, 2027. Ottawa, 2022.
- Flores-Cascante L, Gómez-Gutiérrez J, Gómez Del Prado-Rosas MDC *et al.* Helminth load in feces of free-ranging blue and fin whales from the Gulf of California. *Acta Parasitol* 2019;**64**:625–37. <https://doi.org/10.2478/s11686-019-00069-1>
- Forcada J, Hoffman JI, Gimenez O *et al.* Ninety years of change, from commercial extinction to recovery, range expansion and decline for Antarctic fur seals at South Georgia. *Global Change Biol* 2023;**29**:6867–87. <https://doi.org/10.1111/gcb.16947>
- Fortier L, Le Fèvre J, Legendre L. Export of biogenic carbon to fish and to the deep ocean: the role of large planktonic microphages. *J Plankton Res* 1994;**16**:809–39. <https://doi.org/10.1093/plankt/16.7.809>
- Franco-Cisterna B, Glud A, Bristow LA *et al.* Sinking krill carcasses as hotspots of microbial carbon and nitrogen cycling in the Arctic. *Front Mar Sci* 2022;**9**:1019727. <https://doi.org/10.3389/fmars.2022.1019727>
- Freer JJ, Warwick-Evans V, Skaret G *et al.* A new dynamic distribution model for Antarctic krill reveals interactions with their environment, predators, and the commercial fishery in the south Scotia Sea region. *Limnol Oceanogr* 2025;**70**:833–49. <https://doi.org/10.1002/lno.12809>
- Frolicher TL, Fischer EM, Gruber N. Marine heatwaves under global warming. *Nature* 2018;**560**:360–4. <https://doi.org/10.1038/s41586-018-0383-9>

- Frolicher TL, Laufkötter C. Emerging risks from marine heat waves. *Nat Commun* 2018;**9**:650. <https://doi.org/10.1038/s41467-018-03163-6>
- Gammon E, Moloney CL, Lipinski MR. Diet and condition of juvenile deep-water hake *Merluccius paradoxus* on the west coast of southern Africa. *Afr J Mar Sci* 2022;**44**:311–20. <https://doi.org/10.2989/1814232X.2022.2130427>
- Gíslason Á, Reynisson P, Karlsson H et al. *Euphausiids in Ísafjarðardjúp—A Harvestable Resource?*. Hafnarfjörður: Marine and Freshwater Research Institute, 2021.
- Gleiber MR, Steinberg DK, Ducklow HW. Time series of vertical flux of zooplankton fecal pellets on the continental shelf of the western Antarctic Peninsula. *Mar Ecol Progr Ser* 2012;**471**:23–36. <https://doi.org/10.3354/meps10021>
- Gómez-Gutiérrez J. Comparative study of the population dynamics, secondary production, and reproductive ecology of the euphausiids *Euphausia pacifica* and *Thysanoessa spinifera* in the Oregon upwelling region. Corvallis: Oregon State University, 2003.
- Gómez-Gutiérrez J, Kawaguchi S, Morales-Ávila JR. *Global Diversity and Ecological Function of Parasites of Euphausiids*. New York: Springer International Publishing, 2017.
- Gómez-Gutiérrez J, Martínez-Gómez S, Robinson CJ. Seasonal growth, molt, and egg production rates of *Nyctiphanes simplex* (Crustacea: euphausiacea) juveniles and adults in the Gulf of California. *Mar Ecol Progr Ser* 2012;**455**:173–94.
- Gómez-Gutiérrez J, Morales-Ávila JR. Parasites and diseases. In: Siegel V (ed.), *Biology and Ecology of Antarctic Krill*. Advances in Polar Ecology. New York: Springer, 2016.
- González HE, Sobarzo M, Figueroa D et al. Composition, biomass and potential grazing impact of the crustacean and pelagic tunicates in the northern Humboldt Current area off Chile: differences between El Niño and non-El Niño years. *Mar Ecol Progr Ser* 2000;**195**:201–20. <https://doi.org/10.3354/meps195201>
- Gulland JA. *Population Dynamics of World Fisheries*. Washington: University of Washington, Division of Marine Resources, 1972.
- Gutt J, Siegel V. Benthopelagic aggregations of krill (*Euphausia superba*) on the deeper shelf of the Weddell Sea (Antarctica). *Deep Sea Res Part I* 1994;**41**:169–78. [https://doi.org/10.1016/0967-0637\(94\)90031-0](https://doi.org/10.1016/0967-0637(94)90031-0)
- Hagen W, Yoshida T, Virtue P et al. Effects of a carnivorous diet on the lipids, fatty acids and condition of Antarctic krill, *Euphausia superba*. *Antarct Sci* 2007;**19**:183–8. <https://doi.org/10.1017/S0954102007000259>
- Haraldsson M, Siegel V. Seasonal distribution and life history of *Thysanoessa macrura* (Euphausiacea, Crustacea) in high latitude waters of the Lazarev Sea, Antarctica. *Mar Ecol Progr Ser* 2014;**495**:105–18. <https://doi.org/10.3354/meps10553>
- Harvey HR, Pleuthner RL, Lessard EJ et al. Physical and biochemical properties of the euphausiids *Thysanoessa inermis*, *Thysanoessa raschii*, and *Thysanoessa longipes* in the eastern Bering Sea. *Deep Sea Res II* 2012;**65–70**:173–83.
- Hays GC. A review of the adaptive significance and ecosystem consequences of zooplankton diel vertical migrations. *Hydrobiologia* 2003;**503**:163–70. <https://doi.org/10.1023/B:HYDR.000008476.23617.b0>
- Hays R, Measures LN, Huot J. Euphausiids as intermediate hosts of *Anisakis simplex* in the St. Lawrence estuary. *Can J Zool* 1998;**76**:1226–35. <https://doi.org/10.1139/z98-052>
- Hellessey N, Ericson JA, Nichols PD et al. Seasonal and inter-annual variation in the lipid content and composition of *Euphausia superba* Dana, 1850 (Euphausiacea) samples derived from the Scotia Sea fishery. *J Crustac Biol* 2018;**38**:673–81.
- Hellessey N, Johnson R, Ericson JA et al. Antarctic krill lipid and fatty acid content variability is associated to satellite derived chlorophyll a and sea surface temperatures. *Sci Rep* 2020;**10**:6060. <https://doi.org/10.1038/s41598-020-62800-7>
- Heneghan RF, Everett JD, Blanchard JL et al. Climate-driven zooplankton shifts cause large-scale declines in food quality for fish. *Nat Clim Change* 2023;**13**:470–7. <https://doi.org/10.1038/s41558-023-01630-7>
- Heneghan RF, Everett JD, Sykes P et al. A functional size-spectrum model of the global marine ecosystem that resolves zooplankton composition. *Ecol Modell* 2020;**435**:109265. <https://doi.org/10.1016/j.ecolmodel.2020.109265>
- Hill SL, Atkinson A, Arata JA et al. Observing change in pelagic animals as sampling methods shift: the case of Antarctic krill. *Front Mar Sci* 2024;**11**:1307402. <https://doi.org/10.3389/fmars.2024.1307402>
- Hill SL, Hinke J, Bertrand S et al. Reference points for predators will progress ecosystem-based management of fisheries. *Fish Fish* 2020;**21**:368–78. <https://doi.org/10.1111/faf.12434>
- Hobday AJ, Lough JM. Projected climate change in Australian marine and freshwater environments. *Mar Freshwater Res* 2011;**62**:1000–14. <https://doi.org/10.1071/MF10302>
- Höring F, Biscontin A, Harms L et al. Seasonal gene expression profiling of Antarctic krill in three different latitudinal regions. *Mar Geonomics* 2021;**56**:100806.
- Hünerlage K, Buchholz F. Thermal limits of krill species from the high-Arctic Kongsfjord (Spitsbergen). *Mar Ecol Progr Ser* 2015;**535**:89–98. <https://doi.org/10.3354/meps11408>
- Hunt GL, Ressler PH, Gibson GA et al. Euphausiids in the eastern Bering Sea: a synthesis of recent studies of euphausiid production, consumption and population control. *Deep Sea Res Part II* 2016;**134**:204–22. <https://doi.org/10.1016/j.dsr2.2015.10.007>
- Hunter A, Thorpe SE, McCarthy AE et al. Microplastic hotspots mapped across the Southern Ocean reveal areas of potential ecological impact. *Sci Rep* 2024;**14**:31599. <https://doi.org/10.1038/s41598-024-79816-y>
- Huyer A. Coastal upwelling in the California Current System. *Prog Oceanogr* 1983;**12**:259–84. [https://doi.org/10.1016/0079-6611\(83\)90010-1](https://doi.org/10.1016/0079-6611(83)90010-1)
- ICES. *Working Group on the Integrated Assessments of the Barents Sea (WGIBAR)*. Denmark. 2021.
- ICES. Working Group on Zooplankton Ecology (WGZE). Denmark. 2023.
- Iguchi N, Ikeda T. Growth, metabolism and growth efficiency of a euphausiid crustacean *Euphausia pacifica* in the southern Japan Sea, as influenced by temperature. *J Plankton Res* 1995;**17**:1757–69. <https://doi.org/10.1093/plankt/17.9.1757>
- Iguchi N, Ikeda T. Effects of temperature on metabolism, growth and growth efficiency of *Thysanoessa longipes* (Crustacea: euphausiacea) in the Japan Sea. *J Plankton Res* 2005;**27**:1–10. <https://doi.org/10.1093/plankt/fbh146>
- Ikeda T. Respiration and ammonia excretion of euphausiid crustaceans: synthesis toward a global-bathymetric model. *Mar Biol* 2012;**160**:251–62. <https://doi.org/10.1007/s00227-012-2150-z>

- Ingvaldsen RB, Eriksen E, Gjørseter H *et al.* Under-ice observations by trawls and multi-frequency acoustics in the Central Arctic Ocean reveals abundance and composition of pelagic fauna. *Sci Rep* 2023;**13**:1000. <https://doi.org/10.1038/s41598-023-27957-x>
- Irigoien X, Klevjer TA, Røstad A *et al.* Large mesopelagic fishes biomass and trophic efficiency in the open ocean. *Nat Commun* 2014;**5**:3271. <https://doi.org/10.1038/ncomms4271>
- IUCN. *The IUCN Red List of Threatened Species*. Gland, 2023.
- Jaworski A, Ragnarsson SÁ. Feeding habits of demersal fish in Icelandic waters: a multivariate approach. *ICES J Mar Sci* 2006;**63**:1682–94. <https://doi.org/10.1016/j.icesjms.2006.07.003>
- Johnson ML, Tarling GA. Influence of individual state on swimming capacity and behaviour of Antarctic krill *Euphausia superba*. *Mar Ecol Progr Ser* 2008;**366**:99–110. <https://doi.org/10.3354/meps07533>
- Ju S-J, Kang H-K, Kim WS *et al.* Comparative lipid dynamics of euphausiids from the Antarctic and Northeast Pacific Oceans. *Mar Biol* 2009;**156**:1459–73. <https://doi.org/10.1007/s00227-009-1186-1>
- Kane MK, Atkinson A, Menden-Deuer S. Lowered cameras reveal hidden behaviors of Antarctic krill. *Curr Biol* 2021;**31**:R237–8. <https://doi.org/10.1016/j.cub.2021.01.091>
- Kane MK, Yopak R, Roman C *et al.* Krill motion in the Southern Ocean: quantifying in situ krill movement behaviors and distributions during the late austral autumn and spring. *Limnol Oceanogr* 2018;**63**:2839–57. <https://doi.org/10.1002/lno.11024>
- Kaplan RL, Bernard KB, Derville S *et al.* Krill swarm biomass, energetic density, and species composition drive humpback whale distribution in the Northern California Current. *ICES J Mar Sci* 2025;**82**:fsaf005. <https://doi.org/10.1093/icesjms/fsaf005>
- Kawaguchi S, Atkinson A, Bahlburg D *et al.* Climate change impacts on Antarctic krill behaviour and population dynamics. *Nat Rev Earth Environ* 2024;**5**:43–58. <https://doi.org/10.1038/s43017-023-00504-y>
- Kawaguchi S, Ishida A, King R *et al.* Risk maps for Antarctic krill under projected Southern Ocean acidification. *Nat Clim Change* 2013;**3**:843–7. <https://doi.org/10.1038/nclimate1937>
- Kawaguchi S, Kilpatrick R, Roberts L *et al.* Ocean-bottom krill sex. *J Plankton Res* 2011;**33**:1134–8. <https://doi.org/10.1093/plankt/fbr006>
- Kiko R, Hauss H, Buchholz F *et al.* Ammonium excretion and oxygen respiration of tropical copepods and euphausiids exposed to oxygen minimum zone conditions. *Biogeosciences* 2016;**13**:2241–55. <https://doi.org/10.5194/bg-13-2241-2016>
- Killeen H, Dorman J, Sydeman W *et al.* Effects of a marine heat-wave on adult body length of three numerically dominant krill species in the California Current Ecosystem. *ICES J Mar Sci* 2022;**79**:761–74. <https://doi.org/10.1093/icesjms/fsab215>
- Korejwo E, Panasiuk A, Wawrzynek-Borejko J *et al.* Mercury concentrations in Antarctic zooplankton with a focus on the krill species, *Euphausia superba*. *Sci Total Environ* 2023;**905**:167239. <https://doi.org/10.1016/j.scitotenv.2023.167239>
- Kubilius R, Ona E, Calise L. Measuring in situ krill tilt orientation by stereo photogrammetry: examples for *Euphausia superba* and *Meganyctiphanes norvegica*. *ICES J Mar Sci* 2015;**72**:2494–505. <https://doi.org/10.1093/icesjms/fsv077>
- Kulagin DN, Simakova UV, Lunina AA *et al.* An integrative taxonomic approach to the Atlantic *Hansarsia* (formerly *Nematoscelis*) yields new krill taxa (Crustacea: euphausiidae). *Invertebr Syst* 2024;**38**:IS23034. <https://doi.org/10.1071/IS23034>
- Kwiatkowski L, Torres O, Bopp L *et al.* Twenty-first century ocean warming, acidification, deoxygenation, and upper-ocean nutrient and primary production decline from CMIP6 model projections. *Biogeosciences* 2020;**17**:3439–70. <https://doi.org/10.5194/bg-17-3439-2020>
- Lagos PF, Curtsdotter A, Agüera A *et al.* Fast changes in the bioenergetic balance of krill in response to environmental stress. *Front Mar Sci* 2022a;**8**:782524. <https://doi.org/10.3389/fmars.2021.782524>
- Lagos PF, Curtsdotter A, Agüera A *et al.* Modelling the effects of food limitation and temperature on the growth and reproduction of the krill *Nyctiphanes australis*. *Estuar Coast Shelf Sci* 2022b;**268**:107785. <https://doi.org/10.1016/j.ecss.2022.107785>
- Lagos PF, Gómez-Gutiérrez J, Sabadel AJM *et al.* Review of the biology of the krill genus *Nyctiphanes* G.O. Sars, 1883 (Euphausiacea: euphausiidae): challenges for future research on environmental change. *J Crustac Biol* 2021;**41**:1–17. <https://doi.org/10.1093/jcobiol/ruaa088>
- Laws RM. Seals and whales of the Southern Ocean. *Philos Trans R Soc B* 1977;**279**:81–96.
- Lehette P, Tovar-Sánchez A, Duarte CM *et al.* Krill excretion and its effect on primary production. *Mar Ecol Progr Ser* 2012;**459**:29–38. <https://doi.org/10.3354/meps09746>
- Lévy M, Couespel D, Haëck C *et al.* The impact of fine-scale currents on biogeochemical cycles in a changing ocean. *Ann Rev Mar Sci* 2024;**16**:191–215.
- Li G, Cheng L, Zhu J *et al.* Increasing ocean stratification over the past half-century. *Nat Clim Change* 2020;**10**:1116–23. <https://doi.org/10.1038/s41558-020-00918-2>
- Lilliendahl K, Solmundsson J. An estimate of summer food consumption of six seabird species in Iceland. *ICES J Mar Sci* 1997;**54**:624–30. <https://doi.org/10.1006/jmsc.1997.0240>
- Lilly LE, Ohman MD. Euphausiid spatial displacements and habitat shifts in the southern California Current System in response to El Niño variability. *Prog Oceanogr* 2021;**193**:102544. <https://doi.org/10.1016/j.pocean.2021.102544>
- Linstone HA, Turoff M. *The Delphi Method: Techniques and Applications*. Reading: Addison-Wesley Publishing Company, 1975.
- Loeb VJ, Santora JA. Climate variability and spatiotemporal dynamics of five Southern Ocean krill species. *Prog Oceanogr* 2015;**134**:93–122. <https://doi.org/10.1016/j.pocean.2015.01.002>
- Mackas DL, Beaugrand G. Comparisons of zooplankton time series. *J Mar Syst* 2010;**79**:286–304. <https://doi.org/10.1016/j.jmarsys.2008.11.030>
- Mackas DL, Greve W, Edwards M *et al.* Changing zooplankton seasonality in a changing ocean: comparing time series of zooplankton phenology. *Prog Oceanogr* 2012;**97–100**:31–62. <https://doi.org/10.1016/j.pocean.2011.11.005>
- Mackas DL, Kieser R, Saunders M *et al.* Aggregation of euphausiids and Pacific hake (*Merluccius productus*) along the outer continental shelf off Vancouver Island. *Can J Fish Aquat Sci* 1997;**54**:2080–96. <https://doi.org/10.1139/f97-113>

- Manno C, Corsi I, Rowlands E *et al.* Plastics counteract the ability of Antarctic krill to promote the blue carbon pathway in the deep ocean. *Mar Pollut Bull* 2024;**209**:117238. <https://doi.org/10.1016/j.marpolbul.2024.117238>
- Manno C, Fielding S, Stowasser G *et al.* Continuous moulting by Antarctic krill drives major pulses of carbon export in the north Scotia Sea, Southern Ocean. *Nat Commun* 2020;**11**:6051. <https://doi.org/10.1038/s41467-020-19956-7>
- Marañón E. Phytoplankton size structure. In: Steele JH, Thorpe SA, Turekian KK (eds), *Marine Biology: A derivative of Encyclopedia of Ocean Sciences*. 2nd edn. London: Academic Press, 2009, 4249–56.
- Marinovic BB, Croll DA, Gong N *et al.* Effects of the 1997-1999 El Niño and La Niña events on zooplankton abundance and euphausiid community composition within the Monterey Bay coastal upwelling system. *Prog Oceanogr* 2002;**54**:265–77. [https://doi.org/10.1016/S0079-6611\(02\)00053-8](https://doi.org/10.1016/S0079-6611(02)00053-8)
- Mauchline J. Functional morphology and feeding of euphausiids. In: Felgenhauer BE, Thistle AB, Watling L (eds), *Functional Morphology of Feeding and Grooming in Crustace*. London: CRC Press, 1989, 173–84.
- Maulu S, Musuka CG, Molefe M *et al.* Contribution of fish to food and nutrition security in Southern Africa: challenges and opportunities in fish production. *Front Nutr* 2024;**11**:1424740. <https://doi.org/10.3389/fnut.2024.1424740>
- McCauley RD, Day RD, Swadling KM *et al.* Widely used marine seismic survey air gun operations negatively impact zooplankton. *Nat Ecol Evol* 2017;**1**:0195. <https://doi.org/10.1038/s41559-017-0195>
- McElhany P, Shallin Busch D. Appropriate pCO<sub>2</sub> treatments in ocean acidification experiments. *Mar Biol* 2013;**160**:1807–12. <https://doi.org/10.1007/s00227-012-2052-0>
- McLaskey AK, Keister JE. An integrated field-laboratory investigation of the effects of low oxygen and pH on North Pacific krill (*Euphausia pacifica*). *Mar Biol* 2021;**168**:43. <https://doi.org/10.1007/s00227-021-03845-8>
- McLaskey AK, Keister JE, McElhany P *et al.* Development of *Euphausia pacifica* (krill) larvae is impaired under pCO<sub>2</sub> levels currently observed in the Northeast Pacific. *Mar Ecol Progr Ser* 2016;**555**:65–78. <https://doi.org/10.3354/meps11839>
- Meredith MP, King JC. Rapid climate change in the ocean west of the Antarctic Peninsula during the second half of the 20th century. *Geophys Res Lett* 2005;**32**:L19604. <https://doi.org/10.1029/2005GL024042>
- Meyer B. The overwintering of Antarctic krill, *Euphausia superba*, from an ecophysiological perspective. *Polar Biol* 2012;**35**:15–37. <https://doi.org/10.1007/s00300-011-1120-0>
- Meyer B, Arata JA, Atkinson A *et al.* Adjusting the management of the Antarctic krill fishery to meet the challenges of the 21st century. *Proc Natl Acad Sci USA* 2025;**122**:e2412624122. <https://doi.org/10.1073/pnas.2412624122>
- Meyer B, Atkinson A, Bernard KS *et al.* Successful ecosystem-based management of Antarctic krill should address uncertainties in krill recruitment, behaviour and ecological adaptation. *Commun Earth Environ* 2020;**1**:28. <https://doi.org/10.1038/s43247-020-00026-1>
- Meyer B, Freier U, Grimm V *et al.* The winter pack-ice zone provides a sheltered but food-poor habitat for larval Antarctic krill. *Nat Ecol Evol* 2017;**1**:1853–61. <https://doi.org/10.1038/s41559-017-0368-3>
- Meyer B, Martini P, Biscontin A *et al.* Pyrosequencing and de novo assembly of Antarctic krill (*Euphausia superba*) transcriptome to study the adaptability of krill to climate-induced environmental changes. *Mol Ecol Resour* 2015;**15**:1460–71. <https://doi.org/10.1111/1755-0998.12408>
- Miloslavich P, Seeyave S, Muller-Karger F *et al.* Challenges for global ocean observation: the need for increased human capacity. *J Oper Oceanogr* 2018;**12**:S137–56. <https://doi.org/10.1080/1755876X.2018.1526463>
- Morée AL, Clarke TM, Cheung WWL *et al.* Impact of deoxygenation and warming on global marine species in the 21st century. *Biogeosciences* 2023;**20**:2425–54.
- Nickels CF, Sala LM, Ohman MD. The euphausiid prey field for blue whales around a steep bathymetric feature in the southern California current system. *Limnol Oceanogr* 2018;**64**:390–405. <https://doi.org/10.1002/lno.11047>
- Nicol S, Foster J. The fishery for Antarctic krill: its current status and management regime. In: Siegel V (ed.), *Biology and Ecology of Antarctic krill. Advances in Polar Ecology*. Cham: Springer, 2016.
- Nicol S, O'dor RK. Predatory behaviour of squid (*Illex illecebrosus*) feeding on surface swarms of euphausiids. *Can J Zool* 1985;**63**:15–7. <https://doi.org/10.1139/z85-003>
- Obrodovich SG, Carruthers EH, Rose GA. Bottom-up limits to Newfoundland capelin (*Mallotus villosus*) rebuilding: the euphausiid hypothesis. *ICES J Mar Sci* 2013;**71**:775–83. <https://doi.org/10.1093/icesjms/fst184>
- O'Brien TD. What are marine ecological time series telling us about the ocean? A status report. Paris: IOC-UNESCO, 2017.
- Ohman MD, Davis RE, Sherman JT *et al.* Zooglider: an autonomous vehicle for optical and acoustic sensing of zooplankton. *Limnol Oceanogr Methods* 2019;**17**:69–86.
- O'Loughlin JH, Bernard KS, Daly EA *et al.* Implications of *Pyrosoma atlanticum* range expansion on phytoplankton standing stocks in the Northern California Current. *Prog Oceanogr* 2020;**188**:102424.
- Opstad I, Mangor-Jensen A, Sperfeld E *et al.* Effects of high pCO<sub>2</sub> on the northern krill *Thysanoessa inermis* in relation to carbonate chemistry of its collection area. *Mar Biol* 2018;**165**:116. <https://doi.org/10.1007/s00227-018-3370-7>
- Orlova EL, Dolgov AV, Renaud PE *et al.* Climatic and ecological drivers of euphausiid community structure vary spatially in the Barents Sea: relationships from a long time series (1952–2009). *Front Mar Sci* 2015;**1**:74. <https://doi.org/10.3389/fmars.2014.00074>
- Orr JC, Fabry VJ, Aumont O *et al.* Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature* 2005;**437**:681–6. <https://doi.org/10.1038/nature04095>
- Óskarsson GJ, Gudmundsdóttir A, Sveinbjörnsson S *et al.* Feeding ecology of mackerel and dietary overlap with herring in Icelandic waters. *Mar Biol Res* 2016;**12**:16–29. <https://doi.org/10.1080/17451000.2015.1073327>
- Parés-Escobar F, Lavaniegos BE, Ambriz-Arreola I. Interannual summer variability in oceanic euphausiid communities off the Baja California western coast during 1998–2008. *Prog Oceanogr* 2018;**160**:53–67. <https://doi.org/10.1016/j.pocean.2017.11.009>
- Park J-I, Kang C-K, Suh H-L. Ontogenetic diet shift in the euphausiid *Euphausia pacifica* quantified using stable isotope

- analysis. *Mar Ecol Progr Ser* 2011;**429**:103–9. <https://doi.org/10.3354/meps09091>
- Parouffe A, Garçon V, Dewitte B *et al.* Evaluating future climate change exposure of marine habitat in the South East Pacific based on metabolic constraints. *Front Mar Sci* 2023;**9**:1055875. <https://doi.org/10.3389/fmars.2022.1055875>
- Paulmier A, Ruiz-Pino D. Oxygen minimum zones (OMZs) in the modern ocean. *Prog Oceanogr* 2009;**80**:113–28. <https://doi.org/10.1016/j.pocean.2008.08.001>
- Perry FA, Atkinson A, Sailley SF *et al.* Habitat partitioning in Antarctic krill: spawning hotspots and nursery areas. *PLoS One* 2019;**14**:e0219325. <https://doi.org/10.1371/journal.pone.0219325>
- Peterson WT, Fisher JL, Strub PT *et al.* The pelagic ecosystem in the Northern California Current off Oregon during the 2014–2016 warm anomalies within the context of the past 20 years. *J Geophys Res Oceans* 2017;**122**:7267–90. <https://doi.org/10.1002/2017JC012952>
- Pétursdóttir H, Silva T, Jónsson SP *et al.* Feeding ecology of capelin at new feeding grounds. In: Singh W, Ólafsdóttir AH, Jónsson SP, Óskarsson GJ (eds), *Capelin in a Changing Environment*. Hafnarfirði: Haf- og vatnarsóknir, 2023, 38–44.
- Phillips EM, Chu D, Gauthier S *et al.* Spatiotemporal variability of euphausiids in the California Current Ecosystem: insights from a recently developed time series. *ICES J Mar Sci* 2022;**79**:1312–26. <https://doi.org/10.1093/icesjms/fsac055>
- Pillar SC, Barange M. Feeding habits, daily ration and vertical migration of the cape horse mackerel off South Africa. *South Afr J Mar Sci* 1998;**19**:263–74. <https://doi.org/10.2989/025776198784126683>
- Poloczanska ES, Brown CJ, Sydeman WJ *et al.* Global imprint of climate change on marine life. *Nat Clim Change* 2013;**3**:919–25. <https://doi.org/10.1038/nclimate1958>
- Poloczanska ES, Burrows MT, Brown CJ *et al.* Responses of marine organisms to climate change across oceans. *Front Mar Sci* 2016;**3**:62. <https://doi.org/10.3389/fmars.2016.0062>
- Portner HO. Oxygen- and capacity-limitation of thermal tolerance: a matrix for integrating climate-related stressor effects in marine ecosystems. *J Exp Biol* 2010;**213**:881–93. <https://doi.org/10.1242/jeb.037523>
- Primpke S, Meyer B, Falcou-Préfol M *et al.* At second glance: the importance of strict quality control—a case study on microplastic in the Southern Ocean key species Antarctic krill, *Euphausia superba*. *Sci Total Environ* 2024;**918**:170618. <https://doi.org/10.1016/j.scitotenv.2024.170618>
- Quetin LB, Ross RM, Frazer TK *et al.* Growth of larval krill, *Euphausia superba*, in fall and winter west of the Antarctic Peninsula. *Mar Biol* 2003;**143**:833–43. <https://doi.org/10.1007/s00227-003-1130-8>
- Rainbow PS. Copper, cadmium and zinc concentrations in oceanic amphipod and euphausiid crustaceans, as a source of heavy metals to pelagic seabirds. *Mar Biol* 1989;**103**:513–8. <https://doi.org/10.1007/BF00399583>
- Randall J, Cable J, Guschina IA *et al.* Endemic infection reduces transmission potential of an epidemic parasite during coinfection. *Proc R Soc B Biol Sci* 2013;**280**:20131500. <https://doi.org/10.1098/rspb.2013.1500>
- Ratnarajah L, Abu-Alhaila R, Atkinson A *et al.* Monitoring and modelling marine zooplankton in a changing climate. *Nat Commun* 2023;**14**:564. <https://doi.org/10.1038/s41467-023-36241-5>
- Reiss CS, Cossio AM, Loeb V *et al.* Variations in the biomass of Antarctic krill (*Euphausia superba*) around the South Shetland Islands, 1996–2006. *ICES J Mar Sci* 2008;**65**:497–508. <https://doi.org/10.1093/icesjms/fsn033>
- Reiss CS, Cossio AM, Walsh J *et al.* Glider-based estimates of mesozooplankton biomass density: a fisheries case study on Antarctic krill (*Euphausia superba*) around the Northern Antarctic Peninsula. *Front Mar Sci* 2021;**8**. <https://doi.org/10.3389/fmars.2021.604043>
- Richardson AJ. In hot water: zooplankton and climate change. *ICES J Mar Sci* 2008;**65**:279–95. <https://doi.org/10.1093/icesjms/fsn028>
- Richardson AJ, Bakun A, Hays GC *et al.* The jellyfish joyride: causes, consequences and management responses to a more gelatinous future. *Trends Ecol Evol* 2009;**24**:312–22. <https://doi.org/10.1016/j.tree.2009.01.010>
- Riebesell U, Gattuso J-P. Lessons learned from ocean acidification research. *Nat Clim Change* 2015;**5**:12–4. <https://doi.org/10.1038/nclimate2456>
- Riquelme-Bugueño R, Pantoja-Gutiérrez S, Jorquera E *et al.* Fatty acid composition in the endemic Humboldt Current krill, *Euphausia mucronata* (Crustacea, Euphausiacea) in relation to the phytoplankton community and oceanographic variability off Dichato coast in central Chile. *Prog Oceanogr* 2020a;**188**:102425. <https://doi.org/10.1016/j.pocean.2020.102425>
- Riquelme-Bugueño R, Pérez-Santos I, Alegría N *et al.* Diel vertical migration into anoxic and high-pCO<sub>2</sub> waters: acoustic and net-based krill observations in the Humboldt Current. *Sci Rep* 2020b;**10**:17181. <https://doi.org/10.1038/s41598-020-73702-z>
- Roberts PE. Selective feeding by albacore and skipjack tuna in the New Zealand region in spring. Wellington: Fisheries Research Division, 1974.
- Robertson RR, Bjorkstedt EP. Climate-driven variability in *Euphausia pacifica* size distributions off northern California. *Prog Oceanogr* 2020;**188**:102412. <https://doi.org/10.1016/j.pocean.2020.102412>
- Robinson C, Steinberg DK, Anderson TR *et al.* Mesopelagic zone ecology and biogeochemistry—a synthesis. *Deep Sea Res Part II* 2010;**57**:1504–18. <https://doi.org/10.1016/j.dsr2.2010.02.018>
- Robinson CLK. The consumption of euphausiids by the pelagic fish community off southwestern Vancouver Island, British Columbia. *J Plankton Res* 2000;**22**:1649–62. <https://doi.org/10.1093/plankt/22.9.1649>
- Rockwood RC, Elliott ML, Saenz B *et al.* Modeling predator and prey hotspots: management implications of baleen whale co-occurrence with krill in Central California. *PLoS One* 2020;**15**:e0235603. <https://doi.org/10.1371/journal.pone.0235603>
- Rohner CA, Burgess KB, Rambahinirison JM *et al.* Mobulid rays feed on euphausiids in the Bohol Sea. *R Soc Open Sci* 2017;**4**:161060. <https://doi.org/10.1098/rsos.161060>
- Rowlands E, Galloway T, Cole M *et al.* The effects of combined ocean acidification and nanoplastic exposures on the embryonic development of Antarctic krill. *Front Mar Sci* 2021a;**8**:709763. <https://doi.org/10.3389/fmars.2021.709763>
- Rowlands E, Galloway T, Cole M *et al.* Scoping intergenerational effects of nanoplastic on the lipid reserves of Antarctic krill em-



- phausiid species (*Euphausia pacifica*, *Thysanoessa inspinata*, *T. longipes*, and *Tessarabrachion oculatum*) in the northwestern Pacific. *Deep Sea Res Part I* 2016;**109**:1–9. <https://doi.org/10.1016/j.dsr.2015.12.010>
- Sperfeld E, Mangor-Jensen A, Dalpadado P. Effect of increasing sea water pCO<sub>2</sub> on the northern Atlantic krill species *Nyctiphanes couchii*. *Mar Biol* 2014;**161**:2359–70. <https://doi.org/10.1007/s00227-014-2511-x>
- Sprague LM, Arnold JH. Trends in use and prospects for the future harvest of world fisheries resources. *J Am Oil Chem Soc* 1972;**49**:345–50.
- Steinberg DK, Carlson CA, Bates NR *et al.* Zooplankton vertical migration and the active transport of dissolved organic and inorganic carbon in the Sargasso Sea. *Deep Sea Res Part I* 2000;**47**:137–58. [https://doi.org/10.1016/S0967-0637\(99\)00052-7](https://doi.org/10.1016/S0967-0637(99)00052-7)
- Steinberg DK, Goldthwait SA, Hansell DA. Zooplankton vertical migration and the active transport of dissolved organic and inorganic nitrogen in the Sargasso Sea. *Deep Sea Res Part I* 2002;**49**:1445–61. [https://doi.org/10.1016/S0967-0637\(02\)00037-7](https://doi.org/10.1016/S0967-0637(02)00037-7)
- Steinberg DK, Ruck KE, Gleiber MR *et al.* Long-term (1993–2013) changes in macrozooplankton off the Western Antarctic Peninsula. *Deep Sea Res Part I* 2015;**101**:54–70. <https://doi.org/10.1016/j.dsr.2015.02.009>
- Steinke K, Bernard K, Fontana J *et al.* The energetic cost of early reproductive development in juvenile Antarctic krill at the Western Antarctic Peninsula. *Front Mar Sci* 2022;**9**:1009385. <https://doi.org/10.3389/fmars.2022.1009385>
- Steinke K, Bernard K, Reiss C *et al.* Factors impacting the timing of reproductive development in female Antarctic krill at the northwestern Antarctic Peninsula. *Front Mar Sci* 2024;**11**:1383175. <https://doi.org/10.3389/fmars.2024.1383175>
- Steinke K, Bernard K, Ross R *et al.* Environmental drivers of the physiological condition of mature female Antarctic krill during the spawning season: implications for krill recruitment. *Mar Ecol Progr Ser* 2021;**669**:65–82. <https://doi.org/10.3354/meps13720>
- Stowasser G, Atkinson A, McGill RAR *et al.* Food web dynamics in the Scotia Sea in summer: a stable isotope study. *Deep Sea Res* // 2012;**59–60**:208–21.
- Stramma L, Johnson GC, Sprintall J *et al.* Expanding oxygen-minimum zones in the tropical oceans. *Science* 2008;**320**:655–8. <https://doi.org/10.1126/science.1153847>
- Stramma L, Prince ED, Schmidtko S *et al.* Expansion of oxygen minimum zones may reduce available habitat for tropical pelagic fishes. *Nat Clim Change* 2012;**2**:33–7. <https://doi.org/10.1038/nclimate1304>
- Studer A, Thielges DW, Poulin R. Parasites and global warming: net effects of temperature on an intertidal host–parasite system. *Mar Ecol Progr Ser* 2010;**415**:11–22. <https://doi.org/10.3354/meps08742>
- Suchman CL, Daly EA, Keister JE *et al.* Feeding patterns and predation potential of scyphomedusae in a highly productive upwelling region. *Mar Ecol Progr Ser* 2008;**358**:161–72. <https://doi.org/10.3354/meps07313>
- Sutton AL, Beckley LE. Krill along the 110°E meridian: oceanographic influences on assemblages in the eastern Indian Ocean. *Deep Sea Res Part II* 2022;**202**:105133. <https://doi.org/10.1016/j.dsr2.2022.105133>
- Szabo A. Immature euphausiids do not appear to be prey for humpback whales (*Megaptera novaeangliae*) during spring and summer in Southeast Alaska. *Mar Mammal Sci* 2015;**31**:677–87. <https://doi.org/10.1111/mms.12183>
- Takahashi KT, Kawaguchi S, Kobayashi M *et al.* Eugregarine infection within the digestive tract of larval Antarctic krill, *Euphausia superba*. *Polar Biol* 2011;**34**:1167–74. <https://doi.org/10.1007/s00300-011-0979-0>
- Taki K. Vertical distribution and diel migration of euphausiids from Oyashio Current to Kuroshio area off northeastern Japan. *Plankton Benthos Res* 2008;**3**:27–35. <https://doi.org/10.3800/pbr.3.27>
- Tarling GA, Cuzin-Roudy J. External parasite infestation depends on moult-frequency and age in Antarctic krill (*Euphausia superba*). *Polar Biol* 2008;**31**:121–30. <https://doi.org/10.1007/s00300-007-0339-2>
- Thorpe SE, Tarling GA, Murphy EJ. Circumpolar patterns in Antarctic krill larval recruitment: an environmentally driven model. *Mar Ecol Progr Ser* 2019;**613**:77–96. <https://doi.org/10.3354/meps12887>
- Tovar-Sanchez A, Duarte CM, Hernández-León S *et al.* Krill as a central node for iron cycling in the Southern Ocean. *Geophys Res Lett* 2007;**34**:L11601. <https://doi.org/10.1029/2006GL029096>
- Trathan P, Friedlaender A, Johnson C *et al.* The fishery for Antarctic krill—conflicts between industrial production, protection of biodiversity, and legal governance. *Mar Policy* 2025;**180**:106787. <https://doi.org/10.1016/j.marpol.2025.106787>
- Trathan PN, Hill SL. The importance of krill predation in the Southern Ocean. In: Siegel V (ed.), *Biology and Ecology of Antarctic Krill*. Advances in Polar Ecology. 2016, 321–50.
- Tremblay N, Abele D. Response of three krill species to hypoxia and warming: an experimental approach to oxygen minimum zones expansion in coastal ecosystems. *Mar Ecol* 2016;**37**:179–99. <https://doi.org/10.1111/maec.12258>
- Tremblay N, Cascella K, Toullec J-Y *et al.* Evaluating the hypoxic tolerance of two maturity stages of Antarctic krill (*Euphausia superba*) at its range edge. *Polar Biol* 2024;**47**:1241–54. <https://doi.org/10.1007/s00300-024-03295-0>
- Tremblay N, Gómez-Gutiérrez J, Zenteno-Savín T *et al.* Role of oxidative stress in seasonal and daily vertical migration of three krill species in the Gulf of California. *Limnol Oceanogr* 2010;**55**:2570–84. <https://doi.org/10.4319/lo.2010.55.6.2570>
- Tremblay N, Hunerlage K, Werner T. Hypoxia tolerance of 10 euphausiid species in relation to vertical temperature and oxygen gradients. *Front Physiol* 2020;**11**:248. <https://doi.org/10.3389/fphys.2020.00248>
- Valdez-Cibrián A, Kozak ER, Franco-Gordo C. Microplastic ingestion in marine mesozooplankton species associated with functional feeding traits. *Mar Environ Res* 2024;**200**:106650. <https://doi.org/10.1016/j.marenvres.2024.106650>
- Van de Putte AP, Griffiths HJ, Brooks C *et al.* From data to marine ecosystem assessments of the Southern Ocean: achievements, challenges, and lessons for the future. *Front Mar Sci* 2021;**8**:637063. <https://doi.org/10.3389/fmars.2021.637063>
- Venello TA, Calosi P, Turner LM *et al.* Overwintering individuals of the Arctic krill *Thysanoessa inermis* appear tolerant to short-

