

1 **Environmental and sediment conditions, infaunal benthic communities and**  
2 **biodiversity in the Celtic Sea**

3

4 **Paul J. Somerfield<sup>a,\*</sup>, Imogen L. McClelland<sup>a,b</sup>, C. Louise McNeill<sup>a</sup>, Stefan G. Bolam<sup>c</sup>,**  
5 **Stephen Widdicombe<sup>a,b</sup>**

6 <sup>a</sup> *Plymouth Marine Laboratory, Prospect Place, Plymouth, PL1 3DH UK*

7 <sup>b</sup> *Plymouth University, Drake Circus, Plymouth, England, PL4 8AA UK*

8 <sup>c</sup> *Centre for Environment, Fisheries and Aquaculture Science, Pakefield Road, Lowestoft,*  
9 *Suffolk NR33 0HT UK*

10

11 \* Corresponding author: email [piso@pml.ac.uk](mailto:piso@pml.ac.uk), telephone +44 1752 633100

12

13 Continental Shelf Research, accepted manuscript available online 6<sup>th</sup> September 2018

14 <https://doi.org/10.1016/j.csr.2018.09.002>

15

16 © 2018. This manuscript version is made available under the CC-BY-NC-ND 4.0 license

17 <http://creativecommons.org/licenses/by-nc-nd/4.0/>

18

19 **ABSTRACT**

20 While it is recognised that the nature of the sediment is a major driver of benthic macro-infaunal  
21 community structure, it is also true that diverse environmental factors determine the distribution  
22 and composition of sediments. Among those factors are depth, tidal stress and seasonal  
23 stratification of the water column. In the Celtic Sea an area of seabed approximately 20 km wide and  
24 125 km long was selected in which variation in water depth, stratification, primary production and  
25 current velocity were minimised, but which contained sediments ranging from fine muds to coarse  
26 gravelly sands. 55 stations were sampled across the area using a box-corer. At each station a  
27 comprehensive suite of sediment and biogeochemical measurements were made. Macrofauna were  
28 identified and weighed. Of the stations sampled, four had been chosen as focal sites for a study of  
29 relationships between benthic biogeochemistry and sediment type. Relationships between variation  
30 in environmental and sediment variables and macrofaunal community structure were analysed using  
31 a range of non-parametric multivariate techniques. Environmental variables were discriminated into  
32 situational variables that broadly encapsulate potential drivers of spatial heterogeneity in the  
33 benthos such as depth and fishing effort, and *in-situ* variables that were measured at each site  
34 concurrently with the sampling of the macrobenthos, such as sediment properties and  
35 biogeochemical measurements. Among the former, analyses tended to identify the importance of  
36 average shear stress and depth in explaining observed variation in benthic community structure,  
37 even though the area had been chosen to minimise variation in those factors. Analyses using *in-situ*  
38 measurements of sedimentary conditions at each site identified very fine sand content (correlated  
39 with average shear stress) as the most important explanatory variable. Most of the measured  
40 biogeochemical variables varied with sediment structure, particularly reflecting differences between  
41 finer-grained sediments with higher organic content (generally from deeper areas) and coarser  
42 sediments with lower organic content. While clear spatial heterogeneity in sediments and  
43 associated biogeochemical variables could be demonstrated, spatial variation in benthic abundance  
44 and biomass was less clear. Benthic community structure varied significantly with sediment type, but  
45 did not vary closely with the *in-situ* environmental variables measured at the same sites. This may  
46 indicate that the samples collected were too small to accurately characterise the benthic assemblage  
47 at each site, or that most species inhabiting the area inhabit a range of sediment types, or that  
48 processes which are not reflected in sediment or biogeochemical measurements are also important  
49 determinants of benthic community structure.

50  
51  
52

53 **Key Words:** Benthos . Celtic Sea . Biogeochemistry . Biodiversity . Infauna . Mapping

54  
55  
56

HIGHLIGHTS

- 57 • 55 stations were sampled in the Celtic Sea using a box corer
- 58 • Stations represented a range of environmental and sediment conditions
- 59 • Infaunal microbenthic abundance and biomass were determined
- 60 • Infaunal communities varied weakly with sediment type
- 61 • Variation in communities did not closely match measured environmental variables

62  
63  
64

65 1. INTRODUCTION

66 It is a long-held belief that one of the key determinants of marine benthic infaunal community  
67 structure is the composition of the sediment (Holme, 1966; Thorson, 1957). In continental shelf  
68 systems, sediments generally range from coarse gravels to fine silts, with most of the seabed  
69 consisting mainly of various grades of sand. The composition of sediments found at any particular  
70 location represents the end result of a complex interplay between long-term geological processes  
71 and shorter term physical (hydrodynamic) processes (Gray & Elliott, 2009). Along the shelf, a  
72 gradient in sediment composition is often found from coarse gravels, where tidal and wave energies  
73 are high, to fine silty sediments where levels of physical energy are low. These spatial differences in  
74 sediment compositions are thought to have concomitant effects on the infaunal species composition  
75 (Bolam et al., 2008; Rees et al., 1999; Thorson, 1957). This may reflect adaptation of species to  
76 different physicochemical environments in diverse sediment types, but may also reflect the effects  
77 of physical processes which not only move sediment particles but also distribute pelagic larvae  
78 (Gray, 1974) and the plankton which are the primary source of energy for the benthos (Snelgrove &  
79 Butman 1994). The presence and activities of the organisms themselves can modify the structure  
80 and biogeochemistry of sediments (Gray, 1974; Pearson & Rosenberg 1978; Queiros et al., 2013;  
81 Rhoads, 1974) and the selection of sediment for the settlement of larvae is governed by a  
82 combination of (often interrelated) physical, chemical and biological factors, such as the structure  
83 and contours of the surface, sediment particle size, the presence of organic and inorganic  
84 compounds, biofilms and populations of the same species (Gray, 1974).

85 In addition to sediment measures such as mean grain size, large-scale (> 1000 km<sup>2</sup>) investigations of  
86 the benthos in the NE Atlantic (Ellingsen et al., 2005; Renaud et al., 2009) and around the UK (Rees  
87 et al. 1999, Bolam et al. 2010, Barrio Froján et al. 2012) have identified latitude, depth, tidal stress  
88 and stratification as correlates with large-scale community structure. These large-scale factors are  
89 obviously related to those physical factors that determine the sedimentary environment and its food  
90 supply. Wave energy, and to some extent tidal stress, decline with depth. Stratified waters tend to  
91 be deeper, especially in the North Sea, while shallower waters may remain tidally mixed. Primary  
92 production tends to be less in strongly stratified waters where nutrients can become depleted, but is  
93 also affected by complex combinations of multiple physical drivers (Holt et al., 2016). Jones (1950)  
94 concluded that the distribution of benthic communities was determined by a range of environmental  
95 conditions, and that physical factors were more influential in determining community type than  
96 biological ones, proposing a benthic community classification system based mainly on temperature,  
97 salinity, sediment type and depth. Gray (2002) concluded that environmental factors such as  
98 productivity, temperature, and sediment grain-size diversity play dominant roles in determining  
99 patterns of regional-scale benthic species richness and patterns in species turnover. The overall  
100 picture, therefore, is that the processes identified as correlates of benthic community structure at  
101 large scales are also those that play a role in determining the nature of the sediment in which the  
102 animals live.

103 Given the complex inter-relationships between all of the various factors which may influence the  
104 structure and diversity of infaunal assemblages, disentangling these factors using *in-situ*  
105 observational data and quantifying the relative importance of individual, or subsets, of specific  
106 environmental variables is inherently difficult. In this study, we use marine benthic invertebrate  
107 community structure data together with environmental and biogeochemical data from 55 sampling

108 stations situated in the southern Celtic Sea to identify which physical and chemical aspects of the  
109 sediment environment could best explain the variability seen in benthic fauna across a range of  
110 sediment types. The specific survey area was selected to minimise many of the compromising  
111 environmental factors that could mask the effects of sediment type, such as water depth,  
112 stratification, primary production and current velocity, all of which were comparatively consistent in  
113 the study area (Thompson et al., 2017). The sediment in the survey area covered a broad range of  
114 sediment types from coarse sand with varying gravel components to soft muds. Within this area,  
115 four focal sites were selected for a comprehensive research programme (see Thompson et al., 2017)  
116 investigating seasonality in interactions between sediment type, biology and chemistry, results from  
117 which are presented in this volume and a previous special issue in the journal Biogeochemistry  
118 (volume 135, 2017).

119 The aims of this paper are to examine relationships between macrofaunal abundance, biomass and  
120 diversity and environmental variables which reflect potential drivers of spatial heterogeneity in the  
121 benthos at larger scales (here termed situational environmental variables) and the suite of measured  
122 variables collected concurrently with the macrofaunal samples (here termed *in-situ* environmental  
123 variables) over a large area of Celtic Sea selected to minimise the influence of the situational  
124 variables.

## 125 2. MATERIALS AND METHODS

### 126 2.1 Study area

127 Stretching to the edge of the European continental shelf, and bordered to the north by Ireland, by  
128 England and Wales to the east, and by the western extremity of Brittany to the southeast, the Celtic  
129 Sea is a low-gradient shelf sea with an area of about 130,000 km<sup>2</sup> (Haas et al., 2002). Although large  
130 tidal ridges are found on the mid and outer shelf, the inner shelf is almost featureless with surface  
131 sediments consisting largely of reworked mobile sediments deposited in Pleistocene and early  
132 Holocene transgressions, along with biogenic carbonate. These are mostly fine and coarse sands  
133 with lesser amounts of gravel and mud. Within this large geographic area, a smaller survey area of  
134 approximately 87 x 95 km in the Celtic Sea was identified, with minimal variation in bathymetry  
135 (average water depth 95 m below chart datum), hydrodynamic or water column properties, away  
136 from shallower coastal regions where bed stresses are higher and variable, and from freshwater  
137 inputs which could affect salinity and temperature (Thompson et al., 2017). Although restricted in  
138 spatial extent this selected area still encompassed a wide range of sediment types. Within the study  
139 area a broad transect (approximately 20 km wide with an area of approximately 2500 km<sup>2</sup>), aligned  
140 with the major tidal flow and wave directions running from the south-west to the north-east, was  
141 defined, within which a spatial survey consisting of 55 individual stations was conducted (Fig. 1).

### 142 2.2 Situational variables

143 Six variables were recorded for each sampling station to encapsulate potential drivers of spatial  
144 heterogeneity in the benthos. These were latitude, longitude, water depth, average and maximum  
145 bed shear stress from the NEMO framework adapted for use on the 7 km Atlantic Margin Model  
146 (AMM7) domain (Plymouth Marine Laboratory, unpublished data), and fishing pressure as effort  
147 (h.engine power(kw).yr<sup>-1</sup>), based on data from 2009-2014 (Cefas, unpublished data).

148 2.3 Collection, processing and identification of macrofaunal samples

149 Macrofauna samples were collected from 55 sites (Fig. 1) in March 2015 during cruise DY021  
150 ([https://www.bodc.ac.uk/resources/inventories/cruise\\_inventory/reports/dy021.pdf](https://www.bodc.ac.uk/resources/inventories/cruise_inventory/reports/dy021.pdf)). Samples were  
151 taken with a 0.08 m<sup>2</sup> NIOZ box corer then sieved over a 1 mm mesh and preserved in 10 % buffered  
152 formaldehyde. In the laboratory, each sample was washed over a nested sieve stack of 4 mm, 2 mm  
153 and 1 mm meshes and elutriated to extract the fauna which was then transferred to 70 % industrial  
154 methanol solution (IMS). Organisms were identified to species level using a stereo microscope and  
155 abundances recorded. For each sample, the blotted wet weight of each species was measured on an  
156 electronic microbalance.

157 2.4 Collection of *in-situ* environmental data

158 Physical and chemical data were collected with the macrofauna samples. Although measurements  
159 were made in different depth horizons only measurements from the top 5 cm are used here.  
160 Oxygen penetration depth (OPD) was measured with a Clark-type oxygen microelectrode (Unisense),  
161 equipped with an internal reference and a guard cathode (Revsbech, 1989). Pore-water samples  
162 were taken from NIOZ cores at 1 cm sediment-depth intervals. From each sediment slice  
163 approximately 10 ml of water was extracted using a vacuum motor, syringe filtered (0.2 µm) and  
164 analysed for nutrients using a micro-molar segmented flow auto-analyser. Nutrient data were  
165 pooled, and the mean and maximum measurements for the top 5 cm of nitrite (NO<sub>2</sub>), nitrate (NO<sub>3</sub>),  
166 oxidised nitrogen (NO<sub>2</sub>+NO<sub>3</sub> = NO<sub>x</sub>), phosphate (PO<sub>4</sub>), silicate (SiO<sub>4</sub>) and ammonium (NH<sub>4</sub>) are used in  
167 this study. Sub cores were taken from the NIOZ cores using a 5 cm diameter perspex tube for  
168 particle size analysis (PSA), organic carbon and nitrogen, porosity, chlorophyll a and phaeopigment  
169 measurements. PSA was conducted following the NMBAQC method (Mason, 2011) and graphical  
170 methods were used to determine a range of sediment parameters including median, mean, standard  
171 deviation, skewness and kurtosis. For organic carbon and nitrogen (OC, ON) measurements the  
172 method of Kirsten (1979) was used. Sediment was freeze-dried, ground and analysed using a Carlo  
173 Erba EA1108 Elemental analyser.

174 2.5 Statistical analyses

175 Abundance, biomass and environmental data (Somerfield et al., 2018) were analysed using a range  
176 of univariate, graphical and multivariate approaches (Clarke et al., 2014), primarily using PRIMER v7  
177 (Clarke & Gorley, 2016). Where appropriate, environmental variables (generally those representing  
178 concentrations) were log transformed to reduce the effect of outliers, and normalised by subtracting  
179 the mean and dividing by the standard deviation. The correlation structure among variables was  
180 analysed using a combination of clustering and Type 3 similarity profiles (SIMPROF) analysis  
181 (Somerfield & Clarke, 2013), testing the hypothesis that groups of variables identified by the  
182 clustering algorithm are coherent (varying in the same way across samples). Resemblances among  
183 sites were calculated using Euclidean distance and tested for multivariate structure using clustering  
184 and Type 1 SIMPROF (Clarke et al., 2008). Relationships among samples were ordinated using metric  
185 multidimensional scaling (Clarke et al., 2014). Biotic (abundance and biomass) data were analysed in  
186 their raw form (to focus on patterns in numerical and biomass dominants) and following a fourth-  
187 root transformation. Resemblances among samples were calculated using the Bray-Curtis similarity  
188 measure, tested for structure using SIMPROF and ordinated using nonmetric MDS. Relationships  
189 between biotic similarities and environmental variables were analysed using a non-parametric

190 Mantel test (RELATE) which tests a hypothesis of 'no relationship' between resemblance matrices  
191 (Somerfield & Gage, 2000), the biota-environment matching (BIO-ENV) routine which searches for a  
192 subset of predictor variables that most closely matches a target resemblance matrix, where the  
193 closest match is defined as the highest rank correlation which is then tested using an appropriate  
194 permutation test (Clarke et al., 2008), and analysis of similarities (ANOSIM) tests (Clarke et al., 2014)  
195 which test a hypothesis of 'no difference among (predefined) groups of samples (Clarke, 1993). Taxa  
196 contributing to differences among groups of samples were examined with similarities percentages  
197 (SIMPER) analysis (Clarke, 1993).

198 Univariate measures of community structure were calculated for each sample. These were species  
199 richness (S), total abundance (N), Margalef's d, Pielou's J', Shannon (H'), Simpson (1-λ'), taxonomic  
200 distinctness (Δ\*), average taxonomic distinctness (Δ+), total biomass (B), biomass diversity (as  
201 Pielou's J', Shannon (H'), Simpson (1-λ)) and the Warwick statistic (W), a measure of the difference  
202 in abundance and biomass profiles within each sample (Clarke et al., 2014). N and B were log  
203 transformed prior to analysis. Univariate measures were included in correlation analyses with  
204 environmental variables. Normalised values were used to construct a Euclidean distance matrix  
205 representing variation in multivariate diversity, and this resemblance matrix was subjected to many  
206 of the analyses applied to the biotic resemblance matrices described above (e.g. nMDS, RELATE,  
207 ANOSIM, BIO-ENV).

### 208 3. RESULTS

#### 209 3.1 Environmental variables

210 Analysis of the correlations among variables (Fig. 2) with hierarchical agglomerative clustering of  
211 Pearson correlations and Type 3 SIMPROF analysis showed that many variables are intercorrelated,  
212 many of them significantly so. Group A, longitude and latitude, reflects the overall gradient from  
213 north-east to south-west. No other variables group significantly with these positional variables,  
214 although those that are closest include fishing pressure (Group E), nitrite (Group M) and kurtosis  
215 (Group I). Weakly, but positively, correlated with these variables are depth (Group B) and the C:N  
216 ratio of the sediment (Group J) which do not covary significantly with other variables, but are most  
217 closely grouped with a group of covarying variables (Group H) which includes organic carbon and  
218 nitrogen, porosity, mud content, median and graphical mean particle diameter, and the significantly  
219 intercorrelated variables chlorophyll a and phaeopigments (Group K). Average bed-stress and very  
220 fine sand content covary significantly (Group C), as do silicate and ammonium (Group O), and these  
221 form a group with phosphate which correlates ( $\rho > 0.5$ ) with depth and the other associated  
222 variables. The overall picture, therefore, is one of variation in depth and mud-content, with  
223 associated measures. Variables that are negatively correlated with those already mentioned include  
224 a significantly intercorrelated set of variables (Group G) that includes coarse sand, medium sand,  
225 total sand content and skewness. Oxygen penetration depth (Group L) is most closely associated  
226 with this group, followed by the covarying nitrate and total oxidised nitrogen (NO<sub>x</sub>, Group N), and  
227 then by the significantly covarying very coarse sand and gravel content (Group F). A third group of  
228 covarying variables (Group D) consists of fine sand, maximum stress, and the inclusive graphical  
229 standard deviation of the sediment.

230 Variables can be discriminated into situational variables that give information about the sites  
231 (latitude, longitude, depth, average stress, maximum stress, fishing pressure) and *in-situ* variables  
232 that are derived from sampling at the site (the rest). It is notable that between these groups only  
233 average shear stress and very fine sand content covary significantly.

234 Hierarchical agglomerative clustering with Type 1 SIMPROF analysis (9999 permutations,  $p < 0.001$ )  
235 based on those variables that are derived from sites shows that samples fall into 10 groups (Fig. 3),  
236 ranging from muddy sediments (Group i) to coarse gravelly sediments (Group j). In between, with  
237 significant internal heterogeneity, are sediments which are predominantly sand with variable mud  
238 content. For further analysis samples were grouped into fine (groups h and i), medium (groups c –  
239 g), coarse (groups a and b) and very coarse (group j) sediment types.

### 240 3.2 Relationships with infaunal community structure

241 3402 individuals belonging to 322 taxa were identified, with an average of 61.9 individuals and 26.3  
242 species in each 0.08 m<sup>2</sup> sample. The average biomass in each sample was 0.8 g.

243 To examine which variables or groups of variables may be important determinants of benthic  
244 community structure a series of analyses were conducted (Table 1) using Bray-Curtis similarity  
245 matrices derived from raw and fourth-root transformed abundance and biomass data. These were a  
246 non-parametric Mantel test (RELATE) with a Euclidean distance matrix representing differences in  
247 sample location (a test of spatial autocorrelation), a full-subset (BIO-ENV) search with a subsequent  
248 significance test for the most closely matching subset of situational variables, and another for the  
249 most closely matching subset of measured *in-situ* variables, and finally an analysis of similarities  
250 (ANOSIM) test for differences among sediment groups. For the BIO-ENV tests with *in-situ* variables  
251 one variable from each correlated group was chosen (Table 3), to act as a proxy for all group  
252 members. This is because adding correlated variables does not add information, and the all-subsets  
253 search rapidly becomes prohibitive if there are too many variables. All tests were significant (Table  
254 1), the majority highly so ( $p < 0.001$ ).

255 Among the situational (environmental) variables, a combination of average and maximum shear  
256 stress and depth most closely matched patterns in raw and transformed abundances and  
257 transformed biomass (Table 2). Variation in raw biomass was less closely linked to variation in the  
258 situational variables, with longitude replacing maximum shear stress in the best subset. For BIO-ENV  
259 analyses linking the *in-situ* variables to biotic resemblance matrices a subset of measured variables  
260 were selected (Table 3) to act as proxies for variable groups identified in the Type 3 SIMPROF  
261 analysis (Fig. 2). Among the measured variables (Table 3), the subsets of variables best explaining  
262 variation in benthic community structure all contained very fine sand content (correlated with  
263 average shear stress), median particle diameter (correlated with mud content, mean particle  
264 diameter, organic carbon and nitrogen and porosity), skewness (correlated with total, coarse and  
265 medium sand content) and kurtosis. For raw and transformed abundance, and transformed  
266 biomass, chlorophyll a (correlated with phaeopigments) and phosphate were included, while for raw  
267 and transformed abundance the inclusive graphical standard deviation was included. For both  
268 abundance and biomass NO<sub>x</sub> (correlated with NO<sub>3</sub>) was included in the subsets explaining raw  
269 values, but replaced by oxygen penetration depth in subsets explaining transformed values. The  
270 only other variable included was the C:N ratio, in the subset best explaining variation in raw

271 biomass. The single variable giving the highest rank correlation with inter-sample similarities in each  
272 case was median particle diameter.

273 Nonmetric MDS ordinations of the inter-sample Bray-Curtis similarities calculated from fourth-root  
274 transformed abundance (Fig. 3A) and biomass (Fig. 4A) show that both indicate a gradient in  
275 community structure related to sediment type. The stress of these ordinations is high (>0.2) so the  
276 fine detail should not be interpreted, but the Shepard diagrams (not shown) indicate broad  
277 agreement between the plots and the underlying data. While ANOSIM (Table 1) confirms these  
278 differences among sediment types, analyses using the same methods and criteria applied to the  
279 sedimentary variables (hierarchical agglomerative clustering with Type 1 SIMPROF with 9999  
280 permutations and a significance level of 0.001) shows that while there is significant heterogeneity  
281 among groups of samples, these groups do not map onto those identified in the sediment analyses.  
282 Overlaying these groupings on the same ordination plots indicate that, in terms of abundance and  
283 excepting some outliers, samples fall into one of three groups along a gradient broadly related to  
284 sediment type (Fig. 3B), while in terms of biomass most samples fall into one highly variable group,  
285 with the rest representing samples from the coarser sediments (Fig. 4B).

286 Similarities percentages (SIMPER) analysis based on raw abundances identify the numerical  
287 dominants (contributing up to 25 % of average within-group similarity) in fine sediments as the  
288 polychaetes *Abyssinoe hibernica* and *Mediomastus fragilis*, in medium sediments as the polychaete  
289 *Magelona minuta* and juveniles of the ophiuroid *Amphiura*, in coarse sediments as juvenile  
290 *Amphiura* alone and in very coarse sediments as juvenile *Amphiura* and the small echinoid  
291 *Echinocyamus pusillus*. Analysis of fourth-root transformed abundances adds nemerteans in  
292 medium sediments, the polychaete *Ampharete falcata* and the bivalve *Abra nitida* in coarse  
293 sediments and swaps the capitellid polychaete *Notomastus* sp. for *E. pusillus* in very coarse  
294 sediments. Biomass dominants are *Abyssinoe hibernica* and the capitellid polychaete *Dasybranchus*  
295 sp. in fine sediments, the burrowing shrimp *Callinassa subterranea*, *Notomastus* sp. and the  
296 polychaete *Nephtys hystricus* in medium sediments, the burrowing anemone *Edwardsia claparedii*,  
297 ophiuroid *Amphiura filiformis* and *Notomastus* sp. in coarse sediments and also in very coarse  
298 sediments, but contributions are highly variable and within-group similarities are low (< 10),  
299 reflecting major contributions from small numbers of large individuals. Analysis of transformed  
300 biomass also reflects highly variable contributions. On the transformed scale *Abra nitida* replaces  
301 *Dasybranchus* sp. in fine sediments, a completely different group of taxa (Nemertea, juvenile  
302 *Amphiura*, *Abra nitida* and the polychaete *Amphictene auricoma*) characterises medium sediments  
303 and another characterises coarse sediments (juvenile *Amphiura*, *Abra nitida*, the polychaete  
304 *Ampharete falcata*) while in very coarse sediments juvenile *Amphiura* are added as an important  
305 contributor. The overall pattern is of variable contributions from a number of taxa, with a great deal  
306 of overlap in taxonomic composition between sediment groups.

307 While the four focal sites (A, H, I, G) chosen to represent the range of sedimentary and biological  
308 conditions on the UK shelf fall into different groups defined on the basis of sediment type and  
309 chemical measurements (Figure 2), in terms of community structure based on abundance site A,  
310 representing fine sediments, and I, one of two sites representing medium sand, group together (Fig.  
311 3). Site H does represent intermediate conditions, and site G groups with the coarser sites. In terms  
312 of biomass (Figure 4), however, all sites fall within one group.



313 3.3 Relationships with diversity

314 A correlation analysis (using absolute Pearson correlation) showed that none of the univariate  
315 measures of community structure was closely correlated with any of the situational or measured  
316 variables. To encapsulate variation in diversity among samples a normalised Euclidean distance  
317 matrix was created using values of the full range of univariate measures calculated, which was then  
318 subject to the same range of tests applied to the Bray-Curtis similarities among samples (Tables 2  
319 and 3). Relationships with diversity were generally weaker and less significant. The subset of  
320 situational variables most closely matching variation in diversity included depth and fishing pressure,  
321 and the subset of measured variables included median grain size, inclusive graphical standard  
322 deviation, skewness, kurtosis, nitrite and NO<sub>x</sub>. Notably, the subsets did not include average shear  
323 stress or very fine sand content.

324 4. DISCUSSION

325 A range of different analyses were used to explore relationships between variation in infaunal  
326 community structure (abundance and biomass) and variables reflecting environmental  
327 heterogeneity. Each analysis tests a specific hypothesis, and those hypotheses need not be mutually  
328 exclusive. Significant RELATE tests with a matrix representing distances among samples indicate that  
329 benthic communities are spatially autocorrelated, with a tendency for the assemblage in one sample  
330 to be more similar to those in samples collected close by than to those in samples collected further  
331 away. This could simply reflect the fact that there is spatial autocorrelation among the driving  
332 variables, and as a consequence, among sedimentary measures. BIO-ENV analyses using potential  
333 driving variables tended to identify the importance of average stress and depth, while analyses using  
334 measurements of conditions at each site identified very fine sand content (correlated with average  
335 stress) and those related to sediment type, especially those separating finer sediments from coarser  
336 ones. ANOSIM analyses test whether the communities in groups of samples determined by their  
337 sediment differ, confirming that they do. These findings reflect the fact that environmental  
338 conditions in the sediment are ultimately driven by hydrodynamics of the area, for example high  
339 wave and current activity increase homogeneity of grain sizes in the sediment (Gray & Elliott, 2009).  
340 Other environmental parameters also reflect, directly or indirectly, the hydrodynamic regime.  
341 Particle size and the sorting of the sediment influence the porosity and permeability, which in turn  
342 affect oxygen content and sediment chemistry. Thus, the hydrodynamic nature of the region  
343 determines the sediment type which is linked to the chemical nature of the site. Given these facts, it  
344 is unsurprising that sites chosen as representative of different sediment conditions, but otherwise  
345 similar, turn out to have differing hydrodynamic conditions as a result of, sometimes subtle,  
346 differences in depth and bed stress.

347 Among the situational variables which reflect potential drivers of spatial differences in the  
348 distribution of infaunal communities are average bed shear stress and depth, despite the area being  
349 chosen specifically to minimise such variation. Groups of *in-situ* measured variables were  
350 intercorrelated, and few clear patterns were found. The clearest set of relationships show that the  
351 main differences among groups of samples result from differences between finer and coarser  
352 sediments, or from differences in the chemistry of coarse and fine sediments. The importance to  
353 biotic community structure of variation in the very fine sand content of the sediment, and the  
354 correlation of this with average bed stress, reflects that fact that particles of about 0.18 mm are the

355 easiest to move (Gray & Elliott, 2009). Coarser particles are more difficult for currents to mobilise,  
356 whereas particles finer than 0.18 mm tend to pack into a smooth surface making them difficult to  
357 resuspend. Sediments made up largely of fine sand should be the most stable, as they can only  
358 occur when current or wave disturbance are minimal (Gray & Elliott, 2009).

359 Four focal sites were chosen to represent the range of sediments found on the Celtic Sea continental  
360 shelf (Thompson et al., 2017). While the sites fall within expected groups of samples defined on the  
361 basis of sediment measures, their distinctiveness in terms of biological community structure is less  
362 clear. Partially reflecting the weak relationships between infaunal community structure, sediment  
363 and chemical measures and community structure (as determined by species' abundances) site A,  
364 which represents fine sediments, groups with site I (representing medium sand). In a comparable  
365 analysis based on biomass, all four focal sites fall within a single biotically-defined group. Thus, the  
366 biota inhabiting different sediments in the study area do not seem to discriminate amongst sites as  
367 much as anticipated. Although some species appear to be limited to certain sediment types, such as  
368 the small urchin *Echinocyamus pusillus* which only tends to occur in coarse sediments, there is a high  
369 degree of taxonomic overlap among samples from different grades of sediment with some taxa,  
370 such as *Abra nitida*, frequently occurring in all of them. Analyses suggest that those species which  
371 tend to dominate the biomass at each site are highly variable, while the numerical dominants differ,  
372 to some extent, between different sediments. Thus, the strength of the relationship between  
373 infaunal assemblage structure and sediment composition depends on whether assessments are  
374 biomass- or abundance-based.

375 Communities in sediments with significantly different environmental conditions are similar, and  
376 although there is a gradient in community structure from fine to very coarse sediments there is also  
377 a great deal of overlap. This could reflect the fact that the single samples taken from each site  
378 insufficiently characterise the community present or differences in communities between sites. This  
379 observation also supports the notion that other non-sediment related processes operating at a  
380 range of spatial and temporal scales determine which species may be found at any location. Of note  
381 are the processes that determine larval production, transport, settlement and survival, which were  
382 highlighted as being important in the classic reviews of relationships between sediment and biota  
383 (e.g. Gray, 1974; Rhoads, 1974; Thorson, 1957). Most of the species inhabiting the sediments in the  
384 study area produce pelagic larvae, and larvae produced within the area can potentially reach and  
385 settle elsewhere within the area. How individuals arrive, colonise and survive in different sediments  
386 is poorly understood. Despite the apparent importance of larval biology to understanding  
387 distributions it has not been the focus of research that perhaps it should be. Other factors operate at  
388 larger scales, such as those determining regional and biogeographic differences. Barrio-Froján et al.  
389 (2012) concluded that relationships between patterns in benthic macrofaunal assemblages and the  
390 physical conditions that shape them at broad spatial scales are complex and interconnected, but that  
391 similar faunal assemblages occurred wherever the same environmental conditions were present on  
392 the UK shelf. They concluded that no single variable can be used to predict richness or diversity, but  
393 three together (depth, stratification and kurtosis) could possibly be used to infer relative diversity,  
394 and the species contributing to that diversity in different regions. Operating at a relatively large  
395 spatial scale, their study grouped samples from large parts of the Celtic Sea together as being similar  
396 in terms of species composition. Within a comparatively small part of the Celtic Sea in which  
397 stratification is relatively constant, the results from the present study suggest that depth and  
398 kurtosis (along with several other measures) are important correlates of faunal community

399 structure. Predicting community structure from environmental information to extrapolate broad-  
400 scale patterns from limited sampling (a general problem in sublittoral studies), for “habitat” or  
401 “biotope” mapping, predicting functioning or studying changes in response to long-term variations  
402 such as climate change, is a common approach (e.g. Dutertre et al., 2013; McBreen et al., 2008;  
403 McArthur et al., 2010). However, in addition to potential flaws in the ecological thinking  
404 underpinning some such approaches (Warwick & Somerfield, 2015) the present study supports many  
405 of the conclusions in the review by McArthur et al. (2010), suggesting that the complexity of inter-  
406 relationships among variables, a reliance on samplers which only collect small samples of one  
407 component of the benthic community to ground-truth larger-scale remotely-sensed maps, and the  
408 lack of information about important processes, mean that the use of environmental information to  
409 meaningfully predict benthic community structure remains problematic.

410 In a review of species richness in marine benthic communities, Gray (2002) concluded that available  
411 food resources probably control population densities at a variety of scales and set the maximum  
412 range of species richness, but that variability in species richness for a given resource level is  
413 determined by spatial and temporal heterogeneity in sediment structure caused by both biological  
414 activities and by hydrodynamical and geomorphological factors. As with larval supply, recruitment  
415 and survival, genuine information about food supply is lacking among the variables used in this  
416 study. It may be inferred that chlorophyll, phaeopigments and organic carbon in the sediments  
417 provide some relevant information, but these measured variables probably reflect unconsumed food  
418 rather than estimating the amount available. No single univariate measure of diversity correlated  
419 closely with the situational or measured variables. That being said, regarding diversity as a  
420 multivariate concept there is evidence that it varies with environmental factors in a similar, if  
421 weaker, fashion to community structure defined by abundance, that variation being weakly  
422 influenced by sediment conditions and the processes that drive variation in them. Of course, the  
423 two approaches are not entirely independent, as total abundance and biomass (both log  
424 transformed) are included in the multivariate diversity matrix.

425 Finally, for all the interest in the relationships between macrofaunal community structure and  
426 sediment biogeochemistry, there is little evidence that, at the scales of this investigation, the  
427 relationships are particularly strong. Most of the measured biogeochemical variables varied with  
428 sediment structure, particularly reflecting differences between finer-grained sediments with higher  
429 organic content (generally from deeper areas) and coarser sediments with lower organic content.  
430 While clear spatial heterogeneity in sediments and associated biogeochemical variables could be  
431 demonstrated, spatial variation in benthic abundance and biomass was less clear.

432

#### 433 ACKNOWLEDGEMENTS

434 This work was supported by the Natural Environment Research Council and Department for  
435 Environment, Food and Rural Affairs [grant numbers NE/K00204X/1, Shelf Seas Biogeochemistry  
436 Research Programme and NE/L003279/1, Marine Ecosystems Research Programme]. We thank all  
437 those involved with collecting the samples on DY021, M. Butenschön for providing bed shear stress  
438 data from the AMM7 hindcast and B. Silburn for help with producing Figure 1.

439

440 REFERENCES

- 441 Barrio-Froján, C.R.S., Bolam, S.G., Eggleton, J.D., Mason, C. (2012). Large-scale characterisation of  
442 benthic macrofaunal assemblages in UK waters. *Journal of Sea Research* 69, 53–65.
- 443 Bolam, S.G., Barrio-Froján, C.R.S., Eggleton, J. (2010). Macrofaunal production along the UK  
444 continental shelf. *Journal of Sea Research* 64, 166–179.
- 445 Bolam, S.G., Eggleton, J., Smith, R., Mason, C., Vanstaen, K., Rees, H. (2008) Spatial distribution of  
446 macrofaunal assemblages along the English Channel. *Journal of the Marine Biological Association of*  
447 *the United Kingdom* 88, 675–687.
- 448 Clarke, K.R. (1993) Non-parametric multivariate analyses of changes in community structure.  
449 *Australian Journal of Ecology* 18, 117–143.
- 450 Clarke, K.R., Gorley, R.N. (2016) *Primer v7 User manual/tutorial*. PRIMER-E, Plymouth.
- 451 Clarke, K.R., Somerfield, P.J., Gorley, R.N. (2008). Exploratory null hypothesis testing for community  
452 data: similarity profiles and biota-environment linkage. *Journal of Experimental Marine Biology and*  
453 *Ecology* 366, 56–69.
- 454 Clarke, K.R., Gorley, R.N., Somerfield, P.J., Warwick, R.M. (2014) *Change in marine communities: an*  
455 *approach to statistical analysis and interpretation*, 3rd edn. PRIMER-E, Plymouth.
- 456 Dutertre, M., Hamon, H., Chevalier, C., Ehrhold, A. (2013) The use of the relationships between  
457 environmental factors and benthic macrofaunal distribution in the establishment of a baseline for  
458 coastal management. *ICES Journal of Marine Science* 70, 294–308.
- 459 Ellingsen, K.E., Clarke, K.R., Somerfield, P.J., Warwick, R.M. (2005) Taxonomic distinctness as a  
460 measure of diversity applied over a large scale: the benthos of the Norwegian continental shelf.  
461 *Journal of Animal Ecology* 74, 1069–1079.
- 462 Gray, J.S. (1974) Animal-sediment relationships. *Oceanography and Marine Biology: An Annual*  
463 *Review* 12, 223–261.
- 464 Gray, J.S. (2002) Species richness of marine soft sediments. *Marine Ecology–Progress Series* 244,  
465 285–297.
- 466 Gray, J.S., Elliott, M. (2009) *Ecology of marine sediments*. Oxford University Press, UK.
- 467 Haas, H., van Weering, T.C.E., de Stigter, H. (2002).. Organic carbon in shelf seas: sinks or sources,  
468 processes and products. *Continental Shelf Research* 22, 691–717.
- 469 Holme N.A. (1966) The bottom fauna of the English Channel. Part II. *Journal of the Marine Biological*  
470 *Association of the United Kingdom* 46, 401–493.
- 471 Holt, J., Schrumb, C., Cannaby, H., Daewel, U., Allen, I., Artioli, Y., Bopp, L., Butenshon, M., Fach, B.A.,  
472 Harle, J., Pushpadas, D., Salihoglu, B., Wakelin, S. (2016) Potential impacts of climate change on the  
473 primary production of regional seas: A comparative analysis of five European seas. *Progress in*  
474 *Oceanography* 140, 91–115.

- 475 Jones, N.S. (1950) Marine bottom communities. *Biological Reviews* 25, 283–313.
- 476 Kirsten, W.J. (1979) Automatic methods for the simultaneous determination of carbon, hydrogen,  
477 nitrogen and sulfur, and for sulfur alone in organic and inorganic materials. *Analytical chemistry* 51,  
478 1173–1179.
- 479 Mason, C. (2011) Particle Size Analysis (PSA) for Supporting Biological Analysis. National Marine  
480 Biological AQC Coordinating Committee, UK.
- 481 McArthur, M.A., Brooke, B.P., Przeslawski, R., Ryan, D.A., Lucieer, V.L., Nichol, S., McCallum, A.W.,  
482 Mellin, C., Creswell, I.D., Radke, L.C. (2010) On the use of abiotic surrogates to describe marine  
483 benthic biodiversity. *Estuarine Coastal and Shelf Science* 88, 21–32.
- 484 McBreen, F., Wilson, J.G., Mackie, A.S.Y., Nic Aonghusa, C. (2008) Seabed mapping in the southern  
485 Irish Sea: predicting benthic biological communities based on sediment characteristics.  
486 *Hydrobiologia* 606, 93–103.
- 487 Pearson, T.H., Rosenberg, R. (1978) Macrobenthic succession in relation to organic enrichment and  
488 pollution of the marine environment. *Oceanography and Marine Biology: An Annual Review* 16, 229–  
489 311.
- 490 Queirós, A.M., Birchenough, S.N., Bremner, J., Godbold, J.A., Parker, R.E., Romero-Ramirez, A., Reiss,  
491 H., Solan, M., Somerfield, P.J., Van Colen, C., Van Hoey, G., Widdicombe, S. (2013) A bioturbation  
492 classification of European marine infaunal invertebrates. *Ecology and Evolution* 3, 3958–3985.
- 493 Rees, H.L., Pendle, M.A., Waldock, R., Limpenny, D.S., Boyd S.E. (1999) A comparison of benthic  
494 biodiversity in the North Sea, English Channel, and Celtic Seas. *ICES Journal of Marine Science* 56,  
495 228–246.
- 496 Renaud, P.E., Bjørgesæter, A., Karakassis, I., Kędra, M., Kendall, M.A., Labrune, C., Lampadariou, N.,  
497 Somerfield, P.J., Webb, T.A., Włodarska-Kowalczyk, M., Vanden Berghe, E., Claus, S., Aleffi, F.,  
498 Amoroux, J.M., Bryne, K.H., Cochrane, S.J., Dahle, S., Degraer, S., Denisenko, S.G., Deprez, T.,  
499 Dounas, C., Faulwetter, S., Fleischer, D., Gil, J., Grémare, A., Janas, U., Mackie, A.S.Y., Palerud, R.,  
500 Rumohr, H., Sarda, R., Speybroeck, J., Taboada, S., Van Hoey, G., Węśławski, J.M., Whomersley, P.,  
501 Zettler, M. (2009) Continental-scale patterns in benthic invertebrate diversity: insights from the  
502 MacroBen database. *Marine Ecology–Progress Series* 382, 239–252.
- 503 Revsbech, N.P. (1989) An oxygen microsensor with a guard cathode. *Limnology and Oceanography*  
504 34, 474–478.
- 505 Rhoads, D. C. (1974) Organism-sediment relations on the muddy sea floor. *Oceanography and*  
506 *Marine Biology: An Annual Review* 12, 263–300.
- 507 Snelgrove, P.V.R., Butman, C.A. (1994) Animal-sediment relationships revisited: cause versus effect.  
508 *Oceanography and Marine Biology: An Annual Review* 32, 111–177.
- 509 Somerfield, P.J., Clarke, K.R. (2013) Inverse analysis in non-parametric multivariate analyses:  
510 distinguishing groups of associated species which covary coherently across samples. *Journal of*  
511 *Experimental Marine Biology and Ecology* 449, 261–273.

512 Somerfield, P.J., Gage, J.D. (2000) Community structure of the benthos in Scottish sea-lochs. IV.  
513 Multivariate spatial pattern. *Marine Biology* 136, 1133–1145.

514 Somerfield P, McClelland IL, McNeill CL, Bolam SG, Widdicombe S (2018). Environmental and  
515 sediment conditions, infaunal benthic communities and biodiversity in the Celtic Sea collected from  
516 cruise DY021 spatial survey on the RSS Discovery, March 2015. British Oceanographic Data Centre -  
517 Natural Environment Research Council, UK. doi: 10.5285/6cde14f7-57ab-2be2-e053-6c86abc07695.

518 Thompson, C.E.L., Silburn, B., Williams, M.E., Hull, T., Sivyer, D., Amoudry, L.O., Widdicombe, S.,  
519 Ingels, J., Carnovale, G., McNeill, C.L., Hale, R., Laguionie Marchais, C., Hicks, N., Smith, H., Klar, J.K.,  
520 Hiddink, J.G., Kowalik, J, Kitidis, V., Reynolds, S., Woodward, E.M.S., Tait, K., Homoky, W.B, Kröger, S.,  
521 Godbold, J.A., Aldridge, J., Bolam, S.G., Mayor, D.J, Benoist, N.M.A., Bett, B.J., Morris, K.J., Parker  
522 E.R., Ruhl, H.A., Statham, P.J., Solan, M. (2017) An approach for the identification of exemplar sites  
523 for scaling up targeted field observations of benthic biogeochemistry in heterogeneous  
524 environments. *Biogeochemistry* 135, 1–34.

525 Thorson, G. (1957) Bottom communities (sublittoral or shallow shelf). In: Hedgepeth, J. (ed) *Treatise*  
526 *on marine ecology and paleoecology*. Volume 1. Ecology. Geological Society of America Memoir 67,  
527 461–534.

528 Warwick, R.M., Somerfield, P.J. (2015) Assessing the conservation status of marine habitats:  
529 thoughts from a sandflat on the Isles of Scilly. *Journal of Sea Research* 98, 109–119.

530

531 Table 1. Summary results of tests for relationships between biotic and environmental variables.  
 532 Resemblance: the biotic Bray-Curtis similarity matrix used for the test derived from raw or fourth-  
 533 root transformed (\*) data, or a normalised Euclidean distance matrix calculated from a range of  
 534 diversity measures; Lat/Long: RELATE test with positional variables (Spearman's  $\rho$  and  $p$ ); Situational:  
 535 BIO-ENV with the 6 situational variables (maximum Spearman's  $\rho$  and  $p$ ); In-situ: BIO-ENV with the  
 536 measured sediment variables (maximum Spearman's  $\rho$  and  $p$ , see Table 2 for detail); ANOSIM:  
 537 Global test for differences between sediment groups (Global  $R$  statistic and  $p$ ).

Resemblance	Lat/Long	$p$	Situational	$p$	<i>In-situ</i>	$p$	ANOSIM	$p$
Abundance	0.248	<0.001	0.411	<0.001	0.498	<0.001	0.478	<0.001
Abundance*	0.180	<0.001	0.357	<0.001	0.496	<0.001	0.505	<0.001
Biomass	0.125	0.002	0.174	0.032	0.287	0.003	0.250	<0.001
Biomass*	0.173	<0.001	0.303	<0.001	0.453	<0.001	0.479	<0.001
Diversity	0.065	0.022	0.273	0.002	0.254	0.049	0.089	0.068

538

539

540

541 Table 2. Summary of BIO-ENV analyses with the situational variables. Those variables within the  
 542 subset best explaining variation in each of the resemblance matrices (Ab: Bray-Curtis, raw  
 543 abundances; Ab\*: Bray-Curtis, fourth-root transformed abundances; Bm: Bray-Curtis, raw  
 544 biomasses; Bm\*: Bray-Curtis, fourth-root transformed biomasses; Div: normalised Euclidean  
 545 distance, univariate measures of community structure) denoted by x.

Situational variables	Ab	Ab*	Bm	Bm*	Div
Latitude					
Longitude			x		
Depth	x	x	x	x	x
Average shear stress	x	x	x	x	
Maximum shear stress	x	x		x	
Fishing pressure					x

546

547



548 Table 3. Summary of BIO-ENV analyses with the *in-situ* environmental variables. Group: variables  
 549 grouped together in an analysis of correlations (see Fig. 2); Reduced: variables (R) selected for  
 550 inclusion in analyses, acting as proxies for other group members. Those variables within the subset  
 551 best explaining variation in each of the resemblance matrices (Ab: Bray-Curtis, raw abundances; Ab\*:  
 552 Bray-Curtis, fourth-root transformed abundances; Bm: Bray-Curtis, raw biomasses; Bm\*:  
 553 Bray-Curtis, fourth-root transformed biomasses; Div: normalised Euclidean distance, univariate measures of  
 554 community structure) denoted by x.

<i>In-situ</i> variables	Group	Reduced	Ab	Ab*	Bm	Bm*	Div
Gravel	F						
Very coarse sand	F	R					
Coarse sand	G						
Medium sand	G						
Fine sand	D						
Very fine sand	C	R	x	x	x	x	
Total sand	G						
Mud	H						
Median particle diameter	H	R	x	x	x	x	x
Graphic mean M	H						
Inclusive graphical sd D	D	R	x	x			x
Skewness S	G	R	x	x	x	x	x
Kurtosis K	I	R	x	x	x	x	x
OC	H						
ON	H						
C:N	J	R		x			
Chlorophyll a	K	R	x	x		x	
Phaeopigment	K						
OPD	L	R		x		x	
Porosity	H						
NO <sub>2</sub>	M	R					x
NO <sub>x</sub>	N	R	x		x		x
NO <sub>3</sub>	N						
NH <sub>4</sub>	O	R					
SiO <sub>4</sub>	O						
PO <sub>4</sub>	P	R	x	x		x	

555

556

557 FIGURE LEGENDS

558 1. Map of sampling locations and broad sediment characteristics, modified from Thompson et al.  
559 (2017).

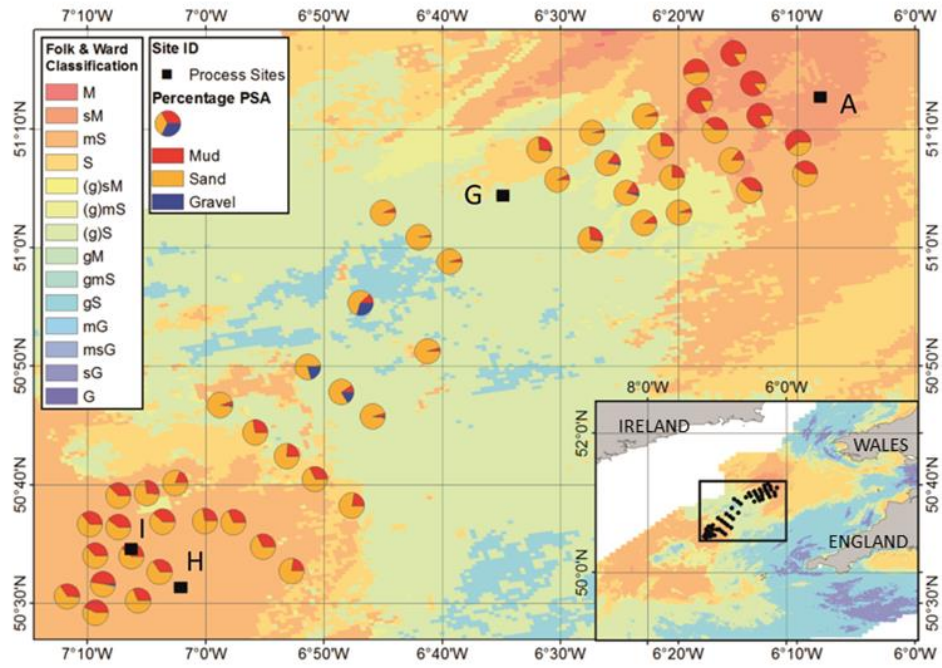
560 2. Dendrogram from hierarchical agglomerative clustering of variables based on Pearson  
561 correlations. Variables grouped using Type 3 SIMPROF ( $p < 0.001$ ).

562 3. Metric MDS (2D stress = 0.09) of sites based on normalised Euclidean distances calculated from  
563 sediment measures. Symbols indicate significant groupings from Type 1 SIMPROF analysis. Letters  
564 (A, G, H, I) indicate focal stations.

565 4. Nonmetric MDS of sites (2D stress = 0.24) based on Bray-Curtis similarities calculated from fourth-  
566 root transformed abundance data overlain with symbols denoting (A) sediment groups (VCoarse =  
567 very coarse) and (B) results from Type 1 SIMPROF analysis ( $p < 0.001$ ). Letters (A, G, H, I) indicate focal  
568 stations.

569 5. Nonmetric MDS of sites (2D stress = 0.25) based on Bray-Curtis similarities calculated from fourth-  
570 root transformed biomass data overlain with symbols denoting (A) sediment groups (VCoarse = very  
571 coarse) and (B) results from Type 1 SIMPROF analysis ( $p < 0.001$ ). Letters (A, G, H, I) indicate focal  
572 stations.

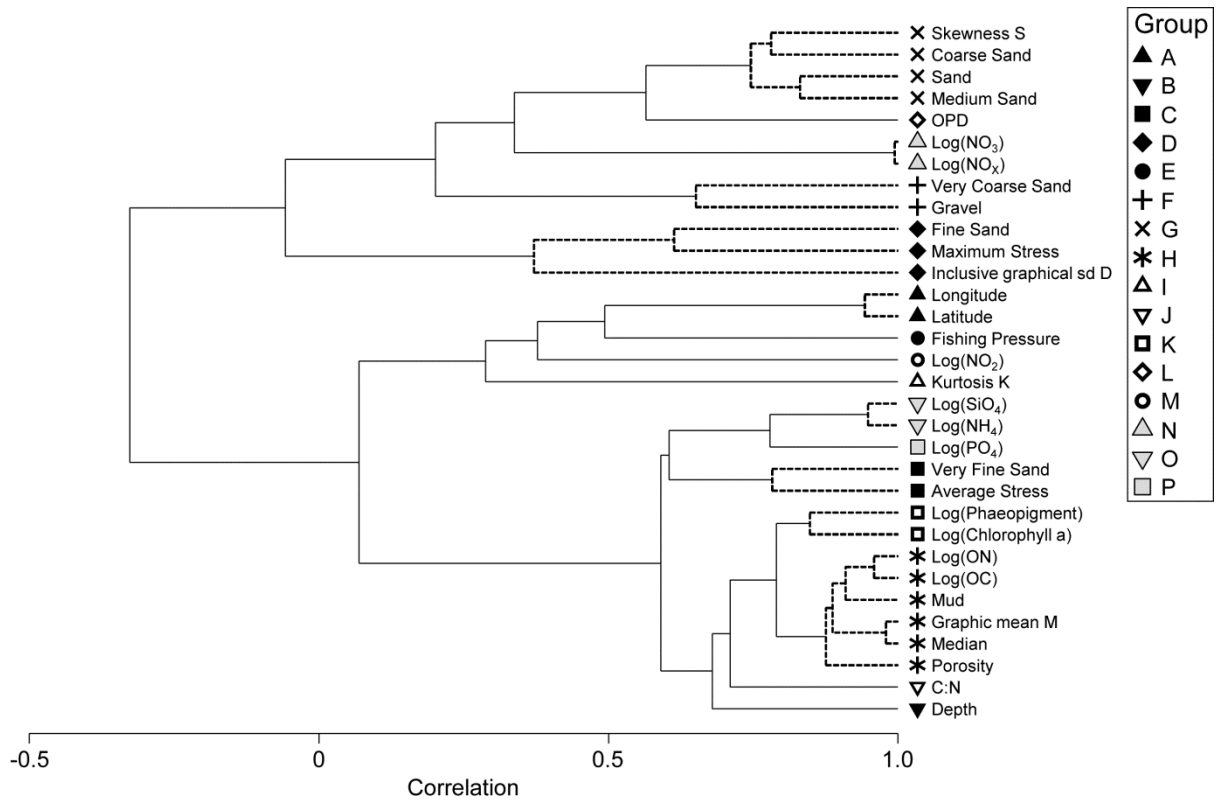
573



574

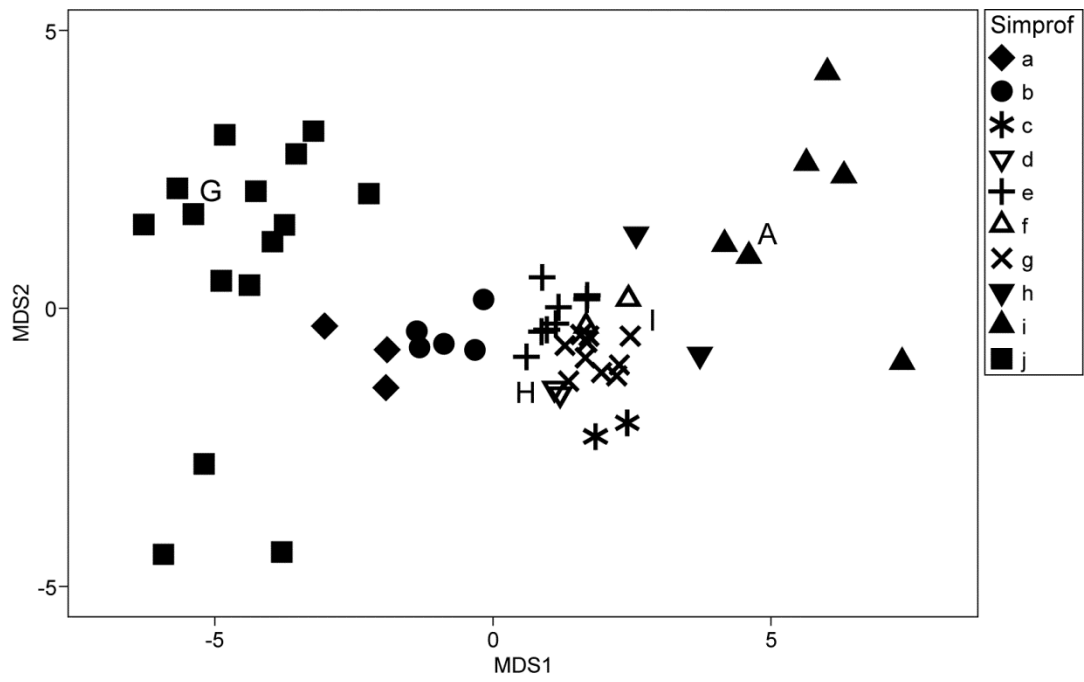
575

576



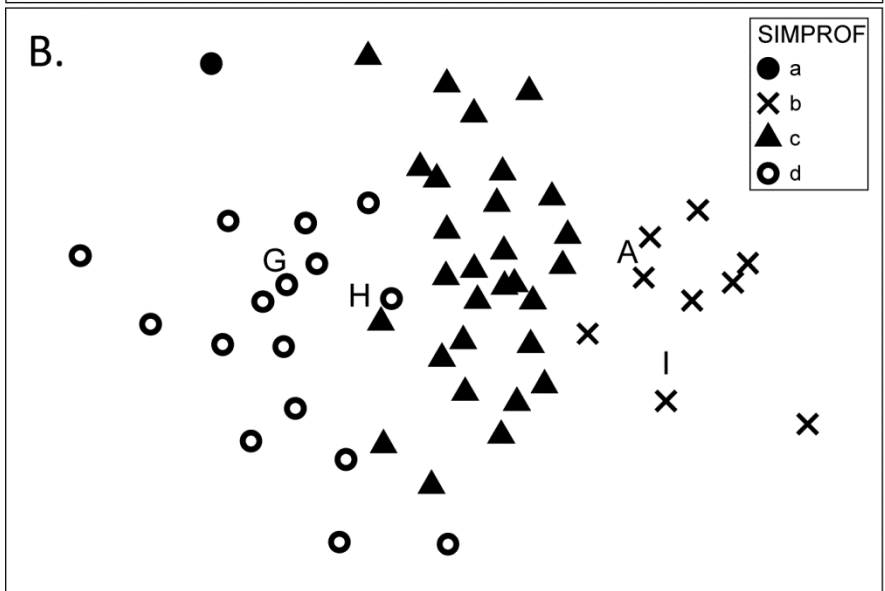
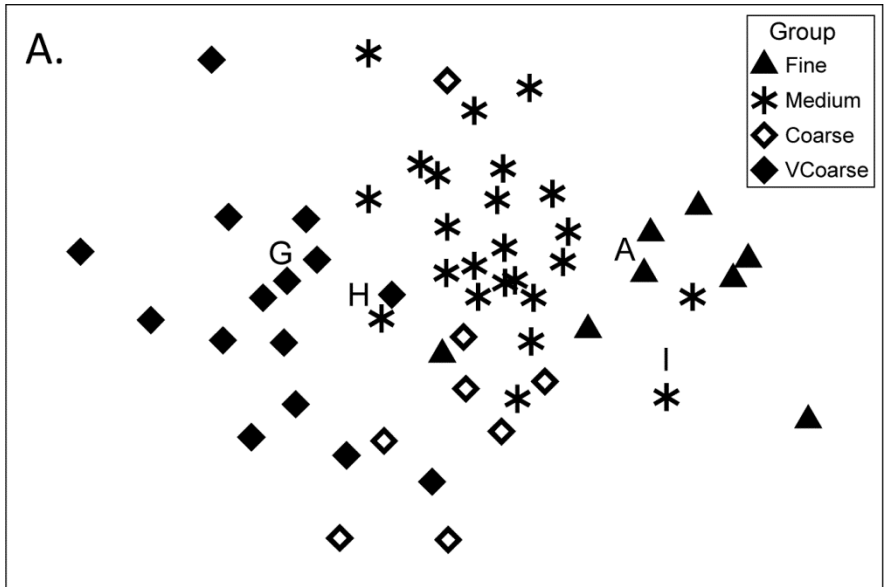
577

578



579

580



581

582

