

RESEARCH ARTICLE

# Pre-ingestive selection capacity and endoscopic analysis in the sympatric bivalves *Mulinia edulis* and *Mytilus chilensis* exposed to diets containing toxic and non-toxic dinoflagellates

Jorge M. Navarro<sup>1,2\*</sup>, John Widdows<sup>3</sup>, Oscar R. Chaparro<sup>1</sup>, Alejandro Ortíz<sup>1,2</sup>, Carla Mellado<sup>1</sup>, Paola A. Villanueva<sup>1</sup>

**1** Instituto de Ciencias Marinas y Limnológicas, Universidad Austral de Chile, Valdivia, Chile, **2** Centro FONDAF de Investigación en Dinámica de Ecosistemas Marinos de Altas Latitudes (IDEAL), Valdivia, Chile, **3** Plymouth Marine Laboratory, Prospect Place, West Hoe, Plymouth, England

\* [jnavarro@uach.cl](mailto:jnavarro@uach.cl)



**OPEN ACCESS**

**Citation:** Navarro JM, Widdows J, Chaparro OR, Ortíz A, Mellado C, Villanueva PA (2018) Pre-ingestive selection capacity and endoscopic analysis in the sympatric bivalves *Mulinia edulis* and *Mytilus chilensis* exposed to diets containing toxic and non-toxic dinoflagellates. PLoS ONE 13 (2): e0193370. <https://doi.org/10.1371/journal.pone.0193370>

**Editor:** Sébastien Duperron, Museum National d'Histoire Naturelle, FRANCE

**Received:** July 11, 2017

**Accepted:** February 8, 2018

**Published:** February 23, 2018

**Copyright:** © 2018 Navarro et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its Supporting Information file.

**Funding:** Study funded by the Comisión Nacional de Investigación Científica y Tecnológica de Chile (CONICYT), through the research grants to JMN: FONDECYT 1161420 and FONDAF IDEAL, grant N° 15150003.

## Abstract

This study investigates the effects of toxic and non-toxic dinoflagellates on two sympatric bivalves, the clam *Mulinia edulis* and the mussel *Mytilus chilensis*. Groups of bivalves were fed one of three diets: (i) the toxic paralytic shellfish producing (PSP) *Alexandrium catenella* + *Isochrysis galbana*; (ii) the non-toxic *Alexandrium affine* + *Isochrysis galbana* and (iii) the control diet of *Isochrysis galbana*. Several physiological traits were measured, such as, clearance rate, pre-ingestive selection efficiency and particle transport velocity in the gill. The clearance rates of both *M. chilensis* and *M. edulis* showed a significant reduction when fed a mixed toxic diet of 50% *Alexandrium catenella*: 50% *Isochrysis galbana*. Similarly, when both species of bivalves were fed with the non-toxic diet (50% *A. affine*: 50% *I. galbana*), clearance rate was significantly lower compared with a diet of 100% *I. galbana*. Under all the experimental diets, *M. chilensis* showed higher clearance rate values, slightly more than double that of *M. edulis*. *M. edulis* and *M. chilensis* have the ability to select particles at the pre-ingestive level, thus eliminating a larger proportion of the toxic dinoflagellate *A. catenella* as well as the non-toxic *A. affine* in the form of pseudofaeces. Higher values of selection efficiency were registered in *M. edulis* than in *M. chilensis* when exposed to the toxic diet. Similar results were observed when these two species were exposed to the diet containing the non-toxic dinoflagellate, explained by the fact that the infaunal *Mulinia edulis* is adapted to dealing with larger particle sizes and higher particle densities (Navarro et al., 1993). The lower transport particle velocity observed in the present work for both species, is related to the reduced clearance rate, the higher particle concentration, and the presence of larger, toxic dinoflagellates. In addition, the species differ in their feeding responses to diets, with and without *A. catenella* or *A. affine*, largely reflecting their adaptations to different environmental conditions. The results suggest that the presence of a dinoflagellate bloom, whether toxic or non-toxic spp in Yaldad Bay, is likely to have a greater impact on the *Mytilus*

**Competing interests:** The authors have declared that no competing interests exist.

*chilensis* than the infaunal *Mulinia edulis*, based on the combined effects on clearance rate, selection efficiency and particle transport velocity.

## Introduction

Suspension feeding bivalves live in environments characterized by large fluctuations in the quantity and quality of the suspended particulate matter. They are exposed to different particle sizes, with variable nutritional content that is sometimes mixed with toxic algal cells. The functional response of filter-feeding organisms to fluctuations in the quantity and quality of suspended particulate matter have been studied by many authors [1–5]. Filter-feeding bivalves can compensate for food quality reductions through physiological mechanisms that improve the energy gain from environments characterised by large fluctuations of the suspended particulate matter. Filter-feeding bivalves can compensate for the “dilution” of desirable particles (i.e. nutritive and/or non-toxic) by undesirable particles (i.e. inorganic and/or toxic) contained in the food supply, through preferential ingestion of desirable particles and selective rejection of undesirable particles via the pseudofaeces [1,3,6–9]. The exact mechanisms used by suspension-feeding bivalves for determining which particles are ingested or rejected as pseudofaeces remain unknown. Selection may involve a physico-chemosensory response by the feeding organs. Particle selection in bivalves could be related to the specific chemical interaction between lectins in the mucus of pallial organs and carbohydrates present on the surfaces of the suspended particles [10,11]. This selective feeding behaviour has been observed in bivalves as a response to the presence of toxic dinoflagellates [1,9].

Harmful algal blooms (HABs) leading to paralytic shellfish poisoning (PSP) events have increased worldwide [12,13] and dinoflagellates of the genus *Alexandrium* are the primary source of the paralytic toxin. The impacts of this genus result in serious economic losses for the shellfish industry and have a negative impact on public health. Numerous authors have studied the impact of HABs on marine filter feeders [14–18] and different physiological and behavioural effects have been described in bivalve species exposed to diets containing PSP [1,19–22]. The results suggest that the different species of filter feeding bivalves have different responses. Some mussels (*Modiolus modiolus*, *Mytilus edulis*, *Perna canaliculus*, *Choromytilus chorus*) appear to be highly unresponsive when feeding on toxic dinoflagellates [14,23–25], but *Mytilus chilensis* shows an initial reduction in filtration activity during the first hours of exposure to *A. catenella*, but returning to normal within 48 h [26]. In contrast, the feeding activity and the growth rate of juvenile and adult oysters and clams were reduced when exposed to algal cells containing PSP [22,27–29]. Similarly, the oyster *Crassostrea gigas* [30] and the clam *Ruditapes philippinarum* [31], significantly reduced filtration activity when exposed to the toxic dinoflagellate *Protogonyaulax tamarensis* strain GT429 (= *Alexandrium tamarensis*). The reduction in the feeding rate affects the energy intake of these organisms, thus reducing the energy available for growth and reproduction, which can negatively impact species fitness [22]. Shumway and Cucci [14] found that most specimens of the clam *Mya arenaria* showed withdrawn or partially withdrawn siphons, which remained retracted for extended periods and this was associated with a highly reduced clearance rate in the presence of *Protogonyaulax tamarensis* (strain GT429). These authors also found that *Mya arenaria* has the capacity, at the pre-ingestive level, to select non-toxic cells for ingestion and toxic dinoflagellate cells for rejection via the pseudofaeces. These authors concluded that the responses observed are species-specific and depend on the geographical origin of the organisms.

Blooms of the dinoflagellate *Alexandrium catenella* occur frequently in the most southerly administrative regions of Chile, and in the last decade the phenomenon has extended northward, reaching Chiloé Island, which is Chile's principal bivalve cultivation area (42° 38' S; 73° 65' W). In early 2016, *A. catenella* blooms extended hundreds of kilometres northward to the Valdivian coast (39° 87' S; 73° 40' W). The occurrence of toxic dinoflagellate blooms in Southern Chile has a negative impact on local bivalve populations as well as to aquaculture activities, leading to a temporary shutdown of harvesting activities. Understanding how to reduce HABs impacts on bivalves is therefore important in the management of these areas.

The main aim of this study was to evaluate the effects of toxic and non-toxic dinoflagellates on the feeding behaviour of the mussel *Mytilus chilensis* and the clam *Mulinia edulis*, two common and important sympatric bivalves inhabiting the fjords of Southern Chile, where HABs have increased in frequency and intensity. For this purpose the following measurements were carried out:

i) feeding rate, ii) capacity to sort toxic and non-toxic dinoflagellate cells (two species of similar size and shape) at the pre-ingestive level, and iii) endoscopic analyses of particle transport on the gills.

## Materials and methods

### Experimental animals

Adult specimens (ca. 5.5 cm shell length) of the mussel *Mytilus chilensis* were collected at the same time from culture rafts and the clam *Mulinia edulis* from tidal flats of the Yaldad Bay, Chiloé (43° 08' S, 73° 44' W). Both species with similar length (ca. 5.5 cm) were collected at the same time during autumn season and transported to the laboratory, acclimatized for two weeks at 14 ° C, 30 psu, and fed continuously with the microalgae *Isochrysis galbana* (ca. 1.0 mg L<sup>-1</sup>). No specific permissions were required for this location. In Chile there is free access to the coast with the public coastal footpath around nearly the whole country. Only for the case of protected areas and National Parks it is necessary to ask special permission. Our field studies do not involve endangered or protected species.

### Experimental design and diet preparation

Monoclonal cultures of the toxic dinoflagellate *Alexandrium catenella* (strain ACC02) and the non-toxic dinoflagellate *Alexandrium affine* and haptophyte *Isochrysis galbana* were used as food in a series of feeding experiments. In all cases, we used the exponential growth phase of the microalgae. In order to generate different food concentrations and levels of toxicity, three experimental diets were prepared (10, 20 and 50 mg L<sup>-1</sup>), all containing 50% of *A. catenella* and 50% of *I. galbana* (based on mg L<sup>-1</sup> but monitored in terms of cell concentration). The experiments were carried out using ten experimental aquaria (one bivalve per aquarium); five containing bivalves exposed to toxic diets (*A. catenella* 50% + *I. galbana* 50%) and five fed the control diet (*I. galbana* 100%). All experiments to determine clearance rate (CR) and pre-ingestive selection efficiency were carried out on three occasions.

The toxin concentration of *A. catenella* cells (strain ACC02) was  $10.3 \pm 0.9$  fmoles STX eq. cell<sup>-1</sup>, according to measurements made in a parallel study [32]. Experimental and control diets had the same weight (mg L<sup>-1</sup>) at each different concentration. One separate experiment (repeated three times) was carried out using a diet free of PSP toxins at a single concentration of 10 mg L<sup>-1</sup>, containing 50% non-toxic *A. affine* and 50% *I. galbana*. Ten bivalves were exposed to toxic free dinoflagellates (*A. affine* 50% + *I. galbana* 50%) and ten fed the control diet (*I. galbana* 100%). Unfortunately, all subsequent attempts at cultivating *A. affine* from various different source cultures failed. To verify the complete absence of PSP toxins in *A. affine*,

samples were sent to the Laboratorio de Toxinas Marinas at Universidad de Chile for quantification of their toxic content, electrophysiologically [33], which showed them to be non-toxic [26].

### Physiological measurements and endoscopy

**Clearance rate.** Clearance rate measurements were performed on individual specimens exposed to different diets prepared with *A. catenella*, *A. affine*, and with the control diet containing *I. galbana*. Food particle concentrations were monitored with an Elzone 180XY particle counter equipped with a 120  $\mu\text{m}$  aperture counting tube. An aquarium without animals was included as a control, to allow for possible cellular sedimentation. Homogenization of experimental medium was maintained by air diffusers, and the experimental specimens were left undisturbed for 1 hour, to allow them to open their valves and begin filtering, before initiating CR measurements. The experiments were performed for a period of 6 hours, with measurements every 30 minutes, and clearance rate ( $\text{L h}^{-1}$ ) was determined on single individuals using the Coughlan method [34]. The consumed cells were replaced every 30 minutes, following each measurement, returning the algal concentrations to their initial food concentrations.

### Pre-ingestive selection efficiency

Experimental mussels and clams were exposed to three food concentrations of toxic diets (equal mixtures of *A. catenella* + *I. galbana*; 10, 20 and 50  $\text{mg L}^{-1}$ ), and to one non-toxic concentration (10  $\text{mg L}^{-1}$ ) of equal mixtures of *A. affine* + *I. galbana*. The different experimental diets were over the threshold for pseudofaeces production [35]. Pseudofaeces were identified as particles wrapped in mucus that are rejected from the inhalant siphon and deposited as a separate pile from the faeces. At the end of each experiment, pseudofaeces were collected and disintegrated by one minute using a slow speed vortex for one minute. Pseudofaeces from each treatment were resuspended in filtered seawater and counted with an inverted microscope (Utermöhl method) for determining *Alexandrium/Isochrysis* cell proportions. Selection efficiency was calculated according to the formula given by Bayne and Hawkins [35]:

$$SE = 1 - \left(\frac{p}{f}\right)$$

Where:  $f$  and  $p$  represent the proportion of *Alexandrium/Isochrysis* in the food and pseudofaeces, respectively.  $SE = 0$  means that there is no selection ( $f = p$ ) and  $SE = 1$  means complete selection and ingestion of only *Isochrysis* cells.

### Particle transport on the gills

Endoscopic examinations were carried out using an Olympus OTV-S4 system, a rigid optic of 1.7 mm diameter, and a xenon light source. The system was connected to a video camera, a monitor, and a video recorder to account for movement and estimate the transport velocity of food particles. A series of six experiments were carried out, each using three different individuals (replicates) of each species (approximately 5.5 cm shell length). These were exposed to a food concentration of ca. 2.0  $\text{mg L}^{-1}$ , consisting of a toxic diet of *A. catenella* + *I. galbana* (50 + 50% based on mass,  $\text{mg L}^{-1}$ ) and a control diet of *I. galbana* (100%). During the endoscopic observations, the experimental individuals were fixed to plastic panels to keep them in a suitable position for the introduction of the endoscope tip at an angle of approximately 45°. In order to introduce the endoscope into the pallial cavity, the valves were perforated on one side and the animals acclimated for 2 days to allow recovery. Since the particle transport of these

two species occur only in the ventral groove of the gills [32], velocity was calculated by following a food particle along the ventral groove and counting the filaments crossed by the particle in a set amount of time. Gill filaments and the space between them were measured on 5 gill pieces using an ocular graduated microscope. Transport velocity was then calculated using the relationship between the time and distance travelled by the particles.

### Statistical analyses

A two-way ANOVA was used to estimate the effects of diet and food concentration on the clearance rates of individuals fed with the toxic diet and also to estimate the effects of diet and species on the clearance rate of individuals fed with *A. affine*. Tukey HSD was employed as a posteriori test on each factor that showed significant differences [36]. A two-way blocked ANOVA was conducted to analyse effects of diet (control and toxic) and species (*M. edulis* and *M. chilensis*) on particle transport velocity (experiments as random factor). Normality and homoscedasticity of the data were tested using Kolmogorov—Smirnov and Bartlett tests, respectively. All analyses were performed with RStudio, version 3.3.1, using the GAD library.

## Results

### Clearance rate studies

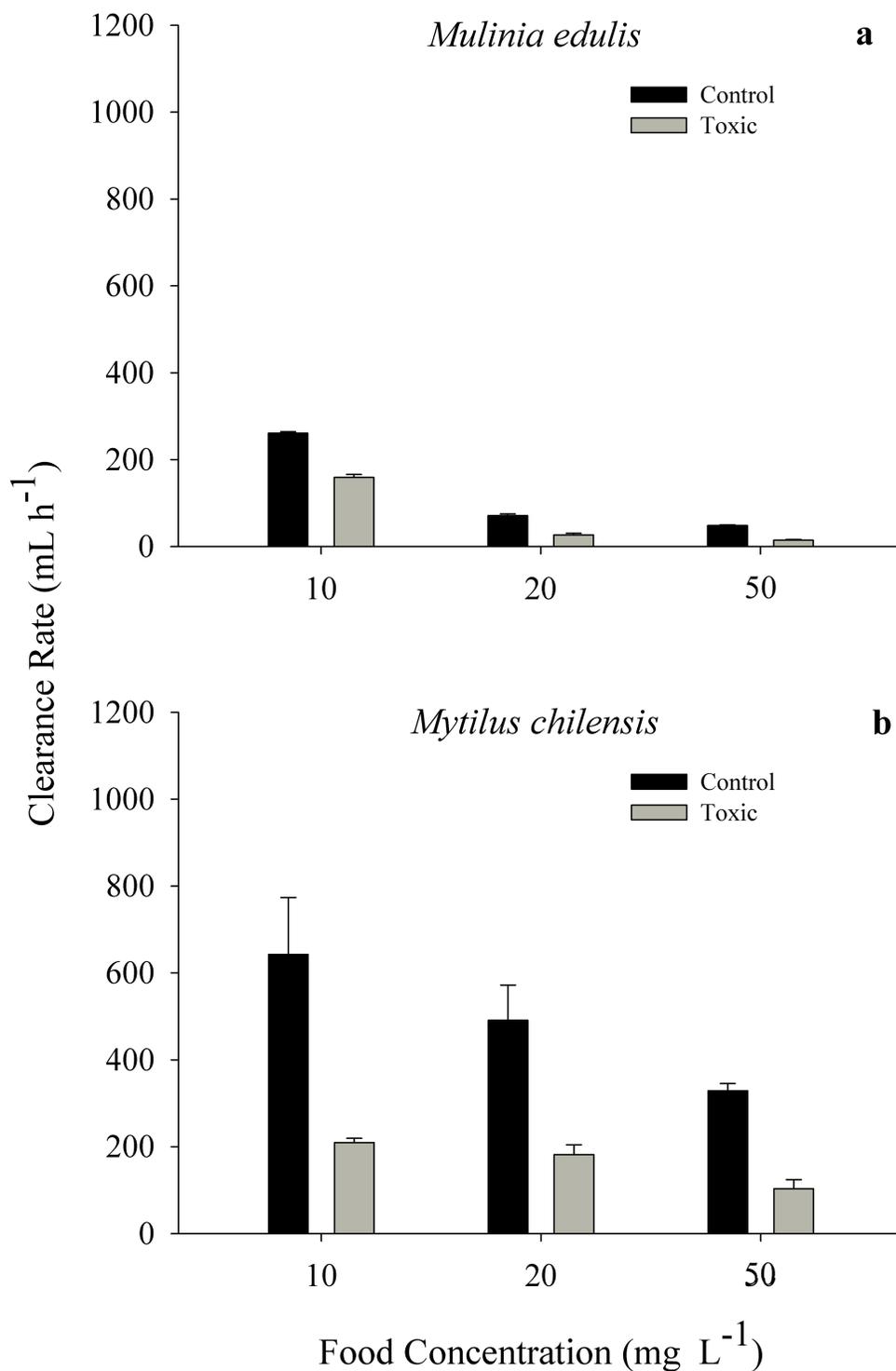
**Exposure to toxic dinoflagellate *Alexandrium catenella*.** Significantly lower clearance rates (CR) were recorded when the clam *Mulinia edulis* and the mussel *Mytilus chilensis* were exposed to the toxic diet and when food concentration increased from 10 to 50 mg L<sup>-1</sup>. The two-way ANOVA showed that both diet ( $F_{1,24} = 71.12, p = 0.001$ ) and food concentration ( $F_{2,24} = 221.10, p = 0.001$ ) had significant effects on the CR of *M. edulis*; the interaction between both factors ( $F_{2,24} = 1.44, p = 0.256$ ) showed no significant effect (Fig 1a; see S1 File). Similar results were obtained for *M. chilensis*, where diet ( $F_{1,24} = 56.08, p = 0.001$ ) and food concentration ( $F_{2,24} = 7.72, p = 0.003$ ) had significant effects on CR ( $F_{2,24} = 0.36, p = 0.700$ ), the interaction between the factors was not significant (Fig 1b; see S1 File). When both bivalves were exposed to different food concentrations of the *A. catenella* diet, higher values were observed in *M. chilensis*. The two-way ANOVA showed a significant effect of species ( $F_{1,24} = 94.05, p = 0.001$ ) and food concentration ( $F_{2,24} = 47.14, p = 0.001$ ), as well as the interaction between both factors ( $F_{2,24} = 11.48, p = 0.001$ ).

### Exposure to non-toxic dinoflagellate *Alexandrium affine*

The CR of *M. edulis* and *M. chilensis* fed with the non-toxic *A. affine* (10 mg L<sup>-1</sup>) was significantly affected by the factors diet and species (two-way ANOVA; diet:  $F_{1,116} = 180.87, p = 0.001$ ; species:  $F_{1,116} = 56.67, p = 0.001$ ). The interaction between these two factors was not significant ( $F_{1,116} = 0.001, p = 0.978$ ). The CR of *M. edulis* fed with the non-toxic *A. affine* diet (mean =  $60.22 \pm 7.06$  mL h<sup>-1</sup>) was markedly lower in comparison with the control diet CR (mean =  $343.29 \pm 49.75$  mL h<sup>-1</sup>) (Fig 2a; see S1 File). Similarly, lower values of CR were observed in *M. chilensis* when exposed to the non-toxic *A. affine* diet (mean =  $137.08 \pm 15.71$  mL h<sup>-1</sup>) in contrast to individuals fed with the control diet (mean =  $733.96 \pm 62.55$  mL h<sup>-1</sup>) (Fig 2b; see S1 File). Note that under both experimental diets (non-toxic *A. affine* and control), *M. chilensis* showed higher CR values slightly more than double that of *M. edulis*.

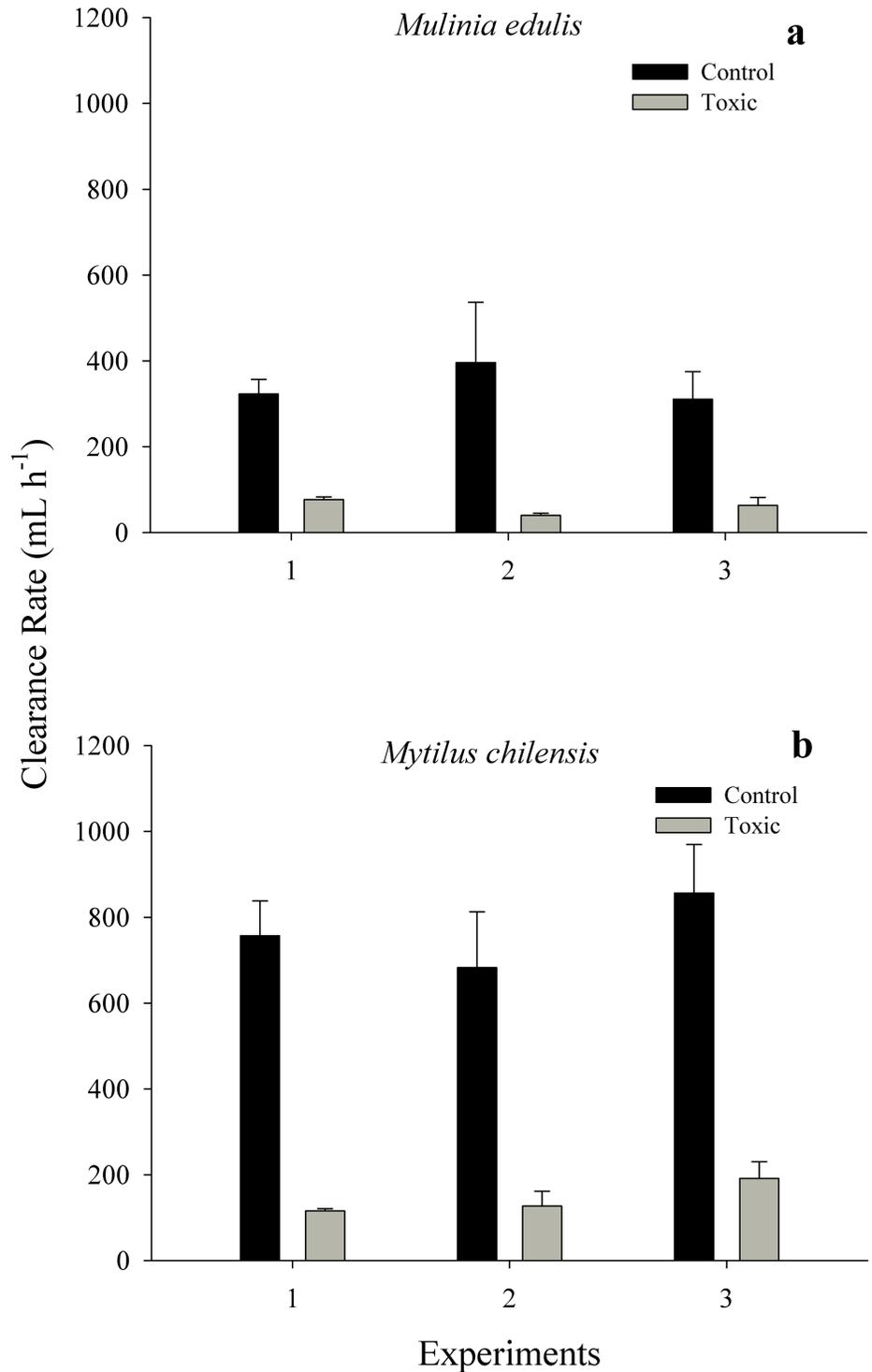
### Pre-ingestive selection efficiency under a toxic and non-toxic diet

*M. edulis* and *M. chilensis* exposed at a food concentration of 10 mg L<sup>-1</sup> have the ability to select particles at the pre-ingestive level, eliminating a larger proportion of the toxic dinoflagellate *A.*



**Fig 1.** Clearance rate of *Mulinia edulis* (a) and *Mytilus chilensis* (b) exposed to different food concentrations (10, 20, 50 mg L<sup>-1</sup>) of a toxic diet (50% *Alexandrium catenella*: 50% *Isochrysis galbana*) and a control diet (100% *Isochrysis galbana*). Values represent means ± SE.

<https://doi.org/10.1371/journal.pone.0193370.g001>

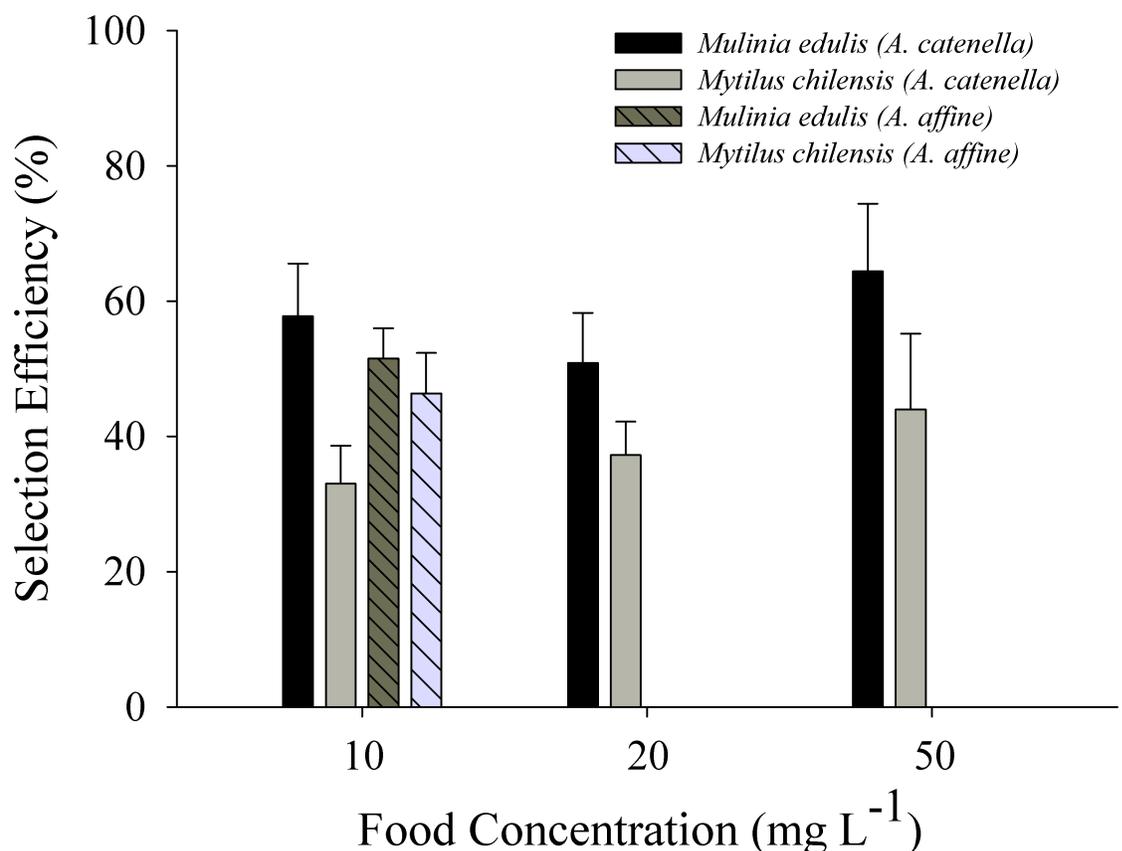


**Fig 2.** Clearance rate of *Mulinia edulis* (a) and *Mytilus chilensis* (b) exposed to a non-toxic diet (50% *Alexandrium affine*: 50% *Isochrysis galbana*) and a control diet (100% *I. galbana*) at 10 mg L<sup>-1</sup>. One experiment was repeated three times. Values represent means ± SE.

<https://doi.org/10.1371/journal.pone.0193370.g002>

*catenella* as well as the non-toxic *A. affine* in the form of pseudofaeces. Both bivalves showed differences between the proportions of *I. galbana*/*A. catenella* and *I. galbana*/*A. affine* cells in food and pseudofaeces. A two-way ANOVA indicated that pre-ingestive selection efficiency was significant for species ( $F_{1,36} = 8.92, p = 0.005$ ); but not for diet ( $F_{1,36} = 0.45, p = 0.506$ ) nor the interaction of species and diet ( $F_{1,36} = 3.77, p = 0.060$ ). Higher values of selection efficiency were registered in *M. edulis* than in *M. chilensis* when exposed to the lower food concentration ( $10 \text{ mg L}^{-1}$ ) of the toxic diet ( $57.8 \pm 7.8\%$  and  $33.0 \pm 5.6\%$ , respectively). Similar results were observed when these two species were exposed to the diet containing the non-toxic dinoflagellate (*M. edulis* mean =  $51.5 \pm 4.5\%$ ; *M. chilensis* mean =  $46.3 \pm 6.0\%$ ).

When both species of bivalves were exposed to different food concentrations (10, 20, and  $50 \text{ mg L}^{-1}$ ) of the toxic diet (50% *A. catenella* + 50% *I. galbana*), pre-ingestive selection efficiency was significant for species ( $F_{1,24} = 94.06, p = 0.001$ ), food concentration ( $F_{2,24} = 47.14, p = 0.001$ ), and the interaction of both factors ( $F_{2,24} = 11.48, p = 0.001$ ). Pre-ingestive selection efficiency measured in *M. edulis* ranged between  $51.5 \pm 7.1$  and  $65.5 \pm 10.1\%$  at concentrations of 20 and  $50 \text{ mg L}^{-1}$ , respectively. In *M. chilensis*, it ranged between  $33.0 \pm 6.0$  and  $44.0 \pm 11.0\%$ , at the lower ( $10 \text{ mg L}^{-1}$ ) and the higher ( $50 \text{ mg L}^{-1}$ ) food concentration, respectively (Fig 3; see S1 File).



**Fig 3. Pre-ingestive selection efficiency of *Mulinia edulis* and *Mytilus chilensis* exposed at three food concentrations (10, 20 and  $50 \text{ mg L}^{-1}$ ) containing mixture of 50% *Alexandrium catenella*: 50% *Isochrysis galbana*.** The selection efficiency of *Mulinia edulis* and *Mytilus chilensis* to a non-toxic diet (50% *Alexandrium affine*: 50% *I. galbana*) was only measured at  $10 \text{ mg L}^{-1}$ . Values represent means  $\pm$  SE.

<https://doi.org/10.1371/journal.pone.0193370.g003>

## Endoscopic measurement of particle transport

For *M. edulis* particle transport velocity, the one-way ANOVA showed significant differences between diets ( $F_{1,24} = 5.34$ ,  $p = 0.013$ ). When fed 100% *I. galbana* ( $2.0 \text{ mg L}^{-1}$ ), particle transport velocity had a mean of  $82.1 \pm 24.6 \mu\text{m s}^{-1}$ ; but particle velocity declined to a mean value of  $44.5 \pm 13.9 \mu\text{m s}^{-1}$  when fed the toxic diet (50% *A. catenella*: 50% *I. galbana*) (Fig 4a; see S1 File). In contrast, the one-way ANOVA showed no significant differences in particle transport velocity between diets for *M. chilensis* ( $F_{1,24} = 0.82$ ,  $p = 0.363$ ). Particle transport velocity for the control diet (mean =  $268 \pm 34.6 \mu\text{m s}^{-1}$ ) overlapped with the particle velocity for the toxic diet (mean =  $229 \pm 32.8 \mu\text{m/s}$ ) (Fig 4b; see S1 File). When results of particle transport rate for both species were included in the analysis, the two-way blocked ANOVA was significant for species ( $F_{1,63} = 43.851$ ,  $p = 0.001$ ). However, diet ( $F_{1,63} = 0.459$ ,  $p = 0.501$ ), the block ( $F_{1,5} = 1.341$ ,  $p = 0.259$ ), and the interaction ( $F_{1,63} = 0.434$ ,  $p = 0.513$ ) showed no significant effects on particle transport velocity.

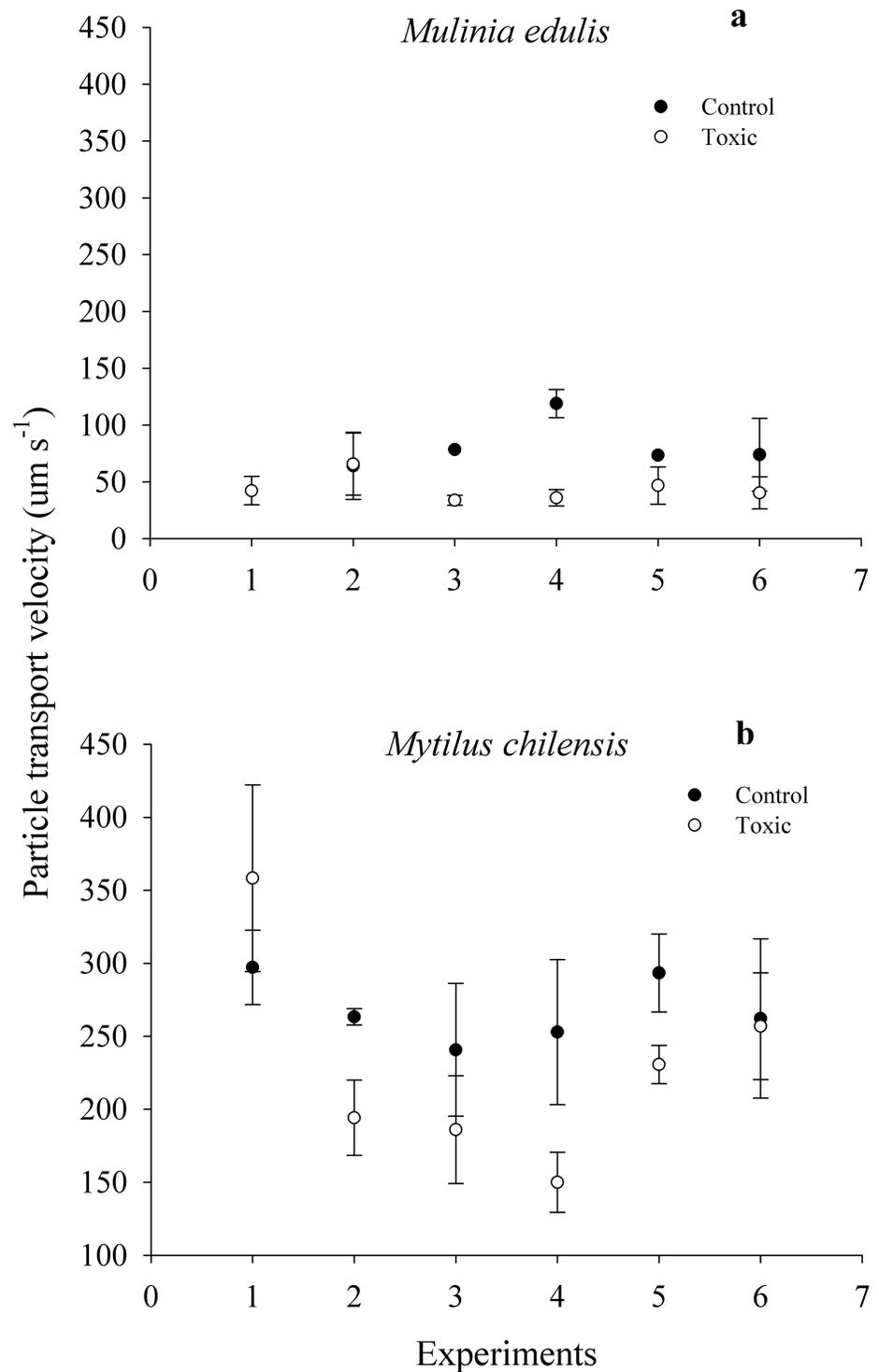
## Discussion

The clearance rate of the two bivalve species when fed on the control diet (100% *Isochrysis galbana*) was markedly different and probably reflected adaptations to their respective environments. Navarro et al. [32] found a direct relationship between clearance rate and gill area in four species of bivalves, with the higher clearance rate in the mussel *Mytilus chilensis* being consistent with a larger gill area and living in a less turbid environment. In contrast, the clam *Mulinia edulis* has a smaller gill area and showed a significantly lower clearance rate, typical of infaunal bivalves living in turbid environments [37–39].

The present study provides evidence that the clearance rates of both *M. edulis* and *M. chilensis* showed a significant reduction when fed a mixed toxic diet of 50% *Alexandrium catenella*: 50% *Isochrysis galbana*. This is comparable to results of earlier studies on the same species [26], in which *M. chilensis* had a clearance rate that was reduced 3.6 fold in response to a toxic diet ( $1.8 \text{ mg L}^{-1}$ ; 50% *A. catenella*: 10% *I. galbana*: 40% sediment) during its first day of exposure. Widdows et al. [40] observed that *Mytilus edulis*, exposed to the toxic dinoflagellate *Gyrodinium aureolum*, also reduced clearance rates. Shumway and Cucci [14] observed species-specific differences in feeding activity of seven bivalve species in the presence of the toxic dinoflagellate *Protogonyaulax tamarensis* strain GT429 (= *Alexandrium tamarensis*) and this varied with collection locality. Responses included shell valve closure and reduced clearance rates in the clam *Mya arenaria* and the mytilid *Geukensia demissa*. In the same study the bivalves *Modiolus modiolus* and *Spisula solidissima*, were not affected by the presence of the toxic dinoflagellates.

In the present study, *M. edulis* and *M. chilensis* also showed a decrease in filtration activity as food concentration increased regardless of the diet. This appears to be a generalized behaviour in bivalves, based on measurements collected under natural [7,39,41] and laboratory [42–44] conditions. This inverse relationship between clearance rate and food concentration has also been described for other species of filter-feeding bivalves, such as *Mytilus edulis* [23], *Crasostrea virginica* [45], and *Placopecten magellanicus* [14,46].

When both bivalve species were exposed to a diet ( $10 \text{ mg L}^{-1}$ ) containing the non-toxic dinoflagellate *A. affine* (50% *A. affine*: 50% *I. galbana*), clearance rates decreased by a factor of more than 5 in both *M. edulis* and *M. chilensis* in comparison to the control diet of 100% *I. galbana*. This is larger than the CR reduction induced by the toxic diet (50% *A. catenella*: 50% *I. galbana*) in *M. edulis* (factor of 1.6) and *M. chilensis* (factor of 3) in comparison to the control diet ( $10 \text{ mg L}^{-1}$ ). Although the experiments with *A. affine* lasted only 6 hours (due to difficulties in culturing this algal species), previous studies with *A. catenella* suggest that this reduction



**Fig 4. Endoscopic measurements of particle transport velocity ( $\mu\text{m s}^{-1}$ ) in (a) *Mulinia edulis* and (b) *Mytilus chilensis*, during exposure to toxic (50% *Alexandrium catenella*: 50% *Isochrysis galbana*) and control (100% *I. galbana*) diets. Values represent means  $\pm$  SE.**

<https://doi.org/10.1371/journal.pone.0193370.g004>

is only temporary, and clearance rate recovery in *M. chilensis* was apparent after 1 to 3 days [26,47]. This indicates that both algal diets containing dinoflagellates significantly reduced the CR of both bivalve species. This reduction may be a response to cell size / volume or the shape of the dinoflagellates rather than a direct toxic effect of *A. catenella*.

Both *M. edulis* and *M. chilensis* showed preferential selection for the smaller, non-toxic, *Isochrysis* cells (ca. 5–6  $\mu\text{m}$ ), resulting in an enhanced rejection of the larger and toxic, *A. catenella* cells (ca. 35  $\mu\text{m}$ ) in the pseudofaeces. The higher values of selection efficiency of *M. edulis* for diets of both toxic and the non-toxic dinoflagellates can be explained by this species' larger labial palps [32], and such preferential particle selection has been described for several bivalve species. For example, adults *Crassostrea gigas* demonstrated enhanced rejection of non-toxic *Alexandrium tamarensense* in the pseudofaeces [30]. Furthermore, the cockle *Cerastoderma edule* showed selection efficiencies as high as 77% [48], *Mytilus edulis* had selection efficiencies of around 50% [49], and *Cerastoderma edule* from the Exe estuary (southwest England) showed efficiencies between 20 and 60% when exposed to a wide range of particle concentrations (4 to 320  $\text{mg L}^{-1}$ ) [2]. In addition, the effects of the toxic dinoflagellate *Protogonyaulax tamarensense* strain GT429 (= *Alexandrium tamarensense*) on several species of bivalves demonstrated that *Mytilus edulis* showed no pre-ingestive selection behaviour, with the toxic cells filtered by the gills and subsequently presenting in both the pseudofaeces and the faeces [14]. In contrast, feeding studies with *M. arenaria* confirmed preferential rejection of the toxic dinoflagellate in the pseudofaeces and reduced ingestion of toxic cells, resulting in low toxin levels (<50  $\mu\text{g STX 100 g}$ ) during the first 10 days [1].

The preferential ingestion of *I. galbana* by *M. edulis* and *M. chilensis* would suggest mechanisms used to reject the two dinoflagellate species at the gills and/or labial palps, presumably based on cell size / shape. Although filter-feeding bivalves have the capacity to ingest particles between 2 and 100  $\mu\text{m}$  [50].

The different clearance rates of the two species were consistent with the observed differences in the transport rate of *I. galbana* and *A. catenella* cells in the food grooves of the gills. *M. chilensis* had a mean transport rate of the toxic diet ca. 3.6-fold higher than *M. edulis*, which was the same as the species difference in clearance rates prior to any adaptation to a change in diet. *M. chilensis* has previously been shown to have significantly higher particle transport velocities than *M. edulis* [32], but that study used a non-toxic diet of pure *I. galbana* at a lower particle concentration (ca. 1  $\text{mg L}^{-1}$ ). These results suggest that the lower transport particle velocity observed in the present work is related to the reduced CR, the higher particle concentration, and the presence of larger, toxic dinoflagellate cells. The lower particle transport rate on the gills of *M. edulis* reflected the high suspended particle concentrations that the clam experiences in its more turbid environment and the particle sorting and processing that are an adaptation for living in these conditions.

In conclusion, the dinoflagellate species (both toxic and non-toxic species) appear to have inhibitory effects on the feeding / ingestion rate, and therefore growth potential, of both *M. edulis* and *M. chilensis*, presumably due to the larger algal cells size / shape. In addition, the species differ in their feeding responses to diets, with and without *A. catenella* or *A. affine*, largely reflecting their adaptations to different environmental conditions. Due to the more turbid waters that characterize their natural environments, the infaunal clam, *M. edulis* shows generally lower clearance rates and slower particle transport rates on the gills as an adaptation to cope with higher particle loads. This enables them to sort particles on the gills and palps and achieve higher selection efficiency, which results in a higher rejection of large particles (e.g. sediment and *Alexandrium* spp.). In contrast the mussel, *M. chilensis*, is an epifaunal bivalve living in less turbid environments. In Yaldad Bay they are grown from rafts higher in the water

column. Consequently, they exhibit a higher clearance rate to increase the collection of food particles from their more dilute environment.

The present findings indicate that the occurrence of dinoflagellate blooms in Yaldad Bay, whether by toxic or non-toxic spp. are likely to have a greater impact on the *Mytilus chilensis* than the infaunal *Mulinia edulis* as a result of their combined effects on clearance rate, selection efficiency and particle transport velocity. The development of HABs produced by the toxic dinoflagellate *A.catenella* are occurring with greater frequency and intensity in southern Chile. Therefore the impact of HABs on the functioning of high densities of filter feeding bivalves in the coastal waters and estuaries will have a significant effect on their role in cycling large amounts of particulate matter within the ecosystem.

## Supporting information

**S1 File. Data for the different physiological variables measured are included in the file S1.** (XLSX)

## Acknowledgments

The authors are grateful to Barbara Cisternas and Verónica Garrido for their contributions to laboratory and fieldwork.

## Author Contributions

**Conceptualization:** Jorge M. Navarro.

**Data curation:** Jorge M. Navarro, Oscar R. Chaparro, Alejandro Ortíz, Carla Mellado, Paola A. Villanueva.

**Formal analysis:** Jorge M. Navarro.

**Funding acquisition:** Jorge M. Navarro, Oscar R. Chaparro.

**Investigation:** Jorge M. Navarro, John Widdows, Oscar R. Chaparro, Alejandro Ortíz, Carla Mellado, Paola A. Villanueva.

**Methodology:** Jorge M. Navarro, Oscar R. Chaparro.

**Project administration:** Jorge M. Navarro.

**Resources:** Jorge M. Navarro.

**Supervision:** Jorge M. Navarro.

**Validation:** Jorge M. Navarro.

**Writing – original draft:** Jorge M. Navarro.

**Writing – review & editing:** Jorge M. Navarro, John Widdows, Oscar R. Chaparro.

## References

1. Shumway SE, Cucci TL, Gainey LF, Yentsch CM. A preliminary study of the behavioral and physiological effects of *Gonyaulax tamarensis* on bivalve molluscs. In: Anderson DM, White AW, Bader DG, editors. Toxic Dinoflagellates. Elsevier; 1985. p. 389–94.
2. Navarro JM, Widdows J. Feeding physiology of *Cerastoderma edule* in response to a wide range of seston concentrations. Mar Ecol Prog Ser. 1997; 152(1–3):175–86.
3. Ward JE, Levinton JS, Shumway SE, Cucci TL. Particle sorting in bivalves: In vivo determination of the pallial organs of selection. Mar Biol. 1998; 131(2):283–92.

4. Zemlys P, Daunys D, Razinkovas A. Revision pre-ingestive selection efficiency definition for suspension feeding bivalves: Facilitating the material fluxes modelling. *Ecol Modell.* 2003; 166(1–2):67–74.
5. Pales Espinosa E, Allam B, Ford SE. Particle selection in the ribbed mussel *Geukensia demissa* and the Eastern oyster *Crassostrea virginica*: Effect of microalgae growth stage. *Estuar Coast Shelf Sci.* 2008; 79(1):1–6.
6. Bayne BL, Iglesias JIP, Hawkins AJS, Navarro E, Heral M, Deslous-Paoli JM. Feeding behaviour of the mussel, *Mytilus edulis*: responses to variations in quantity and organic content of the seston. *J Mar Biol Assoc United Kingdom.* 1993; 73(4):813.
7. Hawkins AJS, Smith RFM, Bayne BL, Héral M. Novel observations underlying the fast growth of suspension-feeding shellfish in turbid environments: *Mytilus edulis*. *Mar Ecol Prog Ser.* 1996; 131(1–3):179–90.
8. Rosa M, Ward JE, Shumway SE, Wikfors GH, Pales Espinosa E, Allam B. Effects of particle surface properties on feeding selectivity in the eastern oyster *Crassostrea virginica* and the blue mussel *Mytilus edulis*. *J Exp Mar Bio Ecol.* 2013; 446.
9. Villanueva PA, Navarro JM. Pre-ingestive selection efficiency in two populations of the razor clam *Tegulus dombeyi* with different histories of exposure to paralytic shellfish poisoning (PSP). *Mar Freshw Behav Physiol.* 2016; 49(4).
10. Pales Espinosa E, Mickael P, Ward JE, Shumway SE, Bassem A. Lectins associated with the feeding organs of the oyster *Crassostrea virginica* can mediate particle selection. *Biol Bull.* 2009; 217(2):130–41. <https://doi.org/10.1086/BBLv217n2p130> PMID: 19875818
11. Pales Espinosa E, Perrigault M, Ward JE, Shumway SE, Allam B. Microalgal cell surface carbohydrates as recognition sites for particle sorting in suspension-feeding bivalves. *Biol Bull.* 2010; 218(1):75–86. <https://doi.org/10.1086/BBLv218n1p75> PMID: 20203255
12. Anderson DM. Toxic Algal Blooms and Red tides: a Global Perspective. *Biology, environmental science and toxicology.* 1989. p. 11–6.
13. Anderson DM, Kulis DM, Doucette GJ, Gallagher JC, Balech E. Biogeography of toxic dinoflagellates in the genus *Alexandrium* from the northeastern United States and Canada. *Mar Biol.* 1994; 120(3):467–78.
14. Shumway SE, Cucci TL. The effects of the toxic dinoflagellate *Protogonyaulax tamarensis* on the feeding and behaviour of bivalve molluscs. *Aquat Toxicol.* 1987; 10(1):9–27.
15. Bricelj VM, Connell L, Konoki K, MacQuarrie SP, Scheuer T, Catterall WA, et al. Sodium channel mutation leading to saxitoxin resistance in clams increases risk of PSP. *Nature.* 2005; 434(7034):763–7. <https://doi.org/10.1038/nature03415> PMID: 15815630
16. Haberkorn H, Lambert C, Le Goïc N, Moal J, Suquet M, Guéguen M, et al. Effects of *Alexandrium minutum* exposure on nutrition-related processes and reproductive output in oysters *Crassostrea gigas*. *Harmful Algae.* 2010; 9(5):427–39.
17. Le Goïc N, Hégaret H, Fabioux C, Miner P, Suquet M, Lambert C, et al. Impact of the toxic dinoflagellate *Alexandrium catenella* on Pacific oyster reproductive output: application of flow cytometry assays on spermatozoa. *Aquat Living Resour.* 2013; 26(3):221–8.
18. Tran D, Ciutat A, Mat A, Massabuau JC, Hégaret H, Lambert C, et al. The toxic dinoflagellate *Alexandrium minutum* disrupts daily rhythmic activities at gene transcription, physiological and behavioral levels in the oyster *Crassostrea gigas*. *Aquat Toxicol.* 2015; 158:41–9. <https://doi.org/10.1016/j.aquatox.2014.10.023> PMID: 25461744
19. Colin SP, Dam HG. Comparison of the functional and numerical responses of resistant versus non-resistant populations of the copepod *Acartia hudsonica* fed the toxic dinoflagellate *Alexandrium tamarense*. *Harmful Algae.* 2007; 6(6):875–82.
20. Fernández-Reiriz MJ, Navarro JM, Contreras AM, Labarta U. Trophic interactions between the toxic dinoflagellate *Alexandrium catenella* and *Mytilus chilensis*: Feeding and digestive behaviour to long-term exposure. *Aquat Toxicol.* 2008; 87(4):245–51. <https://doi.org/10.1016/j.aquatox.2008.02.011> PMID: 18394727
21. Dam HG. Evolutionary Adaptation of Marine Zooplankton to Global Change. *Ann Rev Mar Sci.* 2012; (July 2012):1–22.
22. Navarro JM, Labraña W, Chaparro OR, Cisternas B, Ortiz A. Physiological Constraints in Juvenile *Ostrea chilensis* Fed the Toxic Dinoflagellate *Alexandrium catenella*. *Estuaries and Coasts.* 2016; 39(4):1133–41.
23. Bricelj VM, Lee JH, Cembella AD, Anderson DM. Uptake kinetics of paralytic shellfish toxins from the dinoflagellate *Alexandrium fundyense* in the mussel *Mytilus edulis*. *Mar Ecol Prog Ser.* 1990; 63(2–3):177–88.

24. Marsden ID, Shumway SE. Effects of the toxic dinoflagellate *Alexandrium tamarense* on the greenshell mussel *Perna canaliculus*. *New Zeal J Mar Freshw Res.* 1992; 26(3–4):371–8.
25. Ortiz A. Respuesta fisiológica y dinámica de intoxicación/detoxicación de dos poblaciones del bivalvo *Choromytilus chorus* con diferente historial de exposición al veneno paralizante de molusco (VPM). Universidad Austral de Chile; 2013.
26. Navarro JM, Contreras AM. An integrative response by *Mytilus chilensis* to the toxic dinoflagellate *Alexandrium catenella*. *Mar Biol.* 2010; 157(9):1967–74.
27. Gainey LF, Shumway SE. A compendium of the responses of bivalve mollusc to toxic dinoflagellates. *J Shellfish Res.* 1988; 7:623–8.
28. Bricelj VM, Cembella AD, Laby D, Shumway SE, Cucci TL. Comparative physiological and behavioral responses to PSP toxins in two bivalve molluscs, the softshell clam, *Mya arenaria*, and surfclam, *Spisula solidissima*. In: *Harmful and Toxic Algal Blooms.* 1996. p. 405–8.
29. Lassus P, Baron R, Garen P, Truquet P, Masselin P, Bardouil M, et al. Paralytic shellfish poison outbreaks in the Penzé estuary: Environmental factors affecting toxin uptake in the oyster, *Crassostrea gigas*. *Aquat Living Resour.* 2004; 17:207–14.
30. Bardouil M, Bohec M, Cormerais M, Bougrier S, Lassus P. Experimental study of the effects of a toxic microalgal diet on feeding of the oyster *Crassostrea gigas* Thunberg. *J Shellfish Res.* 1993; 12(2):417–22.
31. Li S-C, Wang W-X, Hsieh DPH. Effects of toxic dinoflagellate *Alexandrium tamarense* on the energy budgets and growth of two marine bivalves. *Mar Environ Res.* 2002; 53(2).
32. Navarro JM, Aguila BL, Machmar F, Chaparro OR, Contreras AM. Dynamic of intoxication and detoxification in juveniles of *Mytilus chilensis* (Bivalvia: Mytilidae) exposed to paralytic shellfish toxins. *Aquat Living Resour.* 2011; 24(1):93–8.
33. Vélez P, Sierralta J, Alcayaga C, Fonseca M, Loyola H, Johns DC, et al. A functional assay for paralytic shellfish toxins that uses recombinant sodium channels. *Toxicon.* 2001; 39(7):929–35. PMID: [11223080](https://pubmed.ncbi.nlm.nih.gov/11223080/)
34. Coughlan J. The estimation of filtering rate from the clearance of suspensions. *Mar Biol.* 1969; 2(4):356–8.
35. Bayne BL, Hawkins AJS. Filter feeding in bivalve mollusc: control on energy balance. In: Mellinger J, Truchot JP, Lahlou B, editors. *Animal nutrition and transport processes, no 1 Nutrition in wild and domestic animals Comparative physiology*, vol 5. Basel: Karger; 1990. p. 70'83.
36. Underwood AJ. *Experiments in Ecology: Their logical design and interpretation using analysis of variance.* University of Cambridge. 1997. p. 1–522.
37. Bacon GS, MacDonald BA, Ward JE. Physiological responses of infaunal (*Mya arenaria*) and epifaunal (*Placopecten magellanicus*) bivalves to variations in the concentration and quality of suspended particles I. Feeding activity and selection. *J Exp Mar Bio Ecol.* 1998; 219(1–2):105–25.
38. Velasco LA, Navarro JM. Feeding physiology of infaunal (*Mulinia edulis*) and epifaunal (*Mytilus chilensis*) bivalves under a wide range of concentrations and qualities of seston. *Mar Ecol Prog Ser.* 2002; 240:143–55.
39. Velasco LA, Navarro JM. Feeding physiology of two bivalves under laboratory and field conditions in response to variable food concentrations. *Mar Ecol Prog Ser.* 2005; 291:115–24.
40. Widdows J, Moore MN, Lowe DM, Salkeld PN. Some effects of a dinoflagellate bloom (*Gyrodinium aureolum*) on the mussel, *Mytilus edulis*. *J Mar Biol Assoc United Kingdom.* 1979 May 11; 59(2):522–4.
41. Urrutia MB, Iglesias JIP, Navarro E, Prou J. Feeding and absorption in *Cerastoderma edule* under environmental conditions in the bay of Marennes-Oleron (Western France). *J Mar Biol Assoc United Kingdom.* 1996; 76(2):431–50.
42. Navarro JM, Winter JE. Ingestion rate, assimilation efficiency and energy balance in *Mytilus chilensis* in relation to body size and different algal concentrations. *Mar Biol.* 1982; 67(3):255–66.
43. Riisgård HU. On measurement of filtration rate in bivalves—the stony road to reliable data: review and interpretation. *Mar Ecol Prog Ser.* 2001; 211:275–91.
44. Fernández-Reiriz MJ, Irisarri J, Labarta U. Flexibility of physiological traits underlying inter-individual growth differences in intertidal and subtidal mussels *Mytilus galloprovincialis*. *PLoS One.* 2016; 11(2).
45. Barille L, Prou J. Modeling Japanese oyster physiological processes under natural tidal variation in suspended particulate matter. In: *International Council for the Exploration of the Sea, Copenhagen (Denmark) Mariculture Comm.* Copenhagen (Denmark); 1993. p. 12.
46. Cranford PJ, Emerson CW, Hargrave BT, Milligan TG. In situ feeding and absorption responses of sea scallops *Placopecten magellanicus* (Gmelin) to storm-induced changes in the quantity and composition of the seston. *J Exp Mar Bio Ecol.* 1998; 219(1–2):45–70.

47. Navarro JM, Contreras AM, Chaparro OR. Short-term feeding response of the mussel *Mytilus chilensis* exposed to diets containing the toxic dinoflagellate *Alexandrium catenella*. *Rev Chil Hist Nat*. 2008; 81 (1):41–9.
48. Kiørboe T, Møhlenberg F. Particle Selection in Suspension-Feeding Bivalves. *Mar Ecol Prog Ser*. 1981; 5:291–6.
49. Bayne BL, Hawkins AJS, Navarro E, Iglesias JIP. Effects of seston concentration of feeding, digestion and growth in the mussel *Mytilus edulis*. *Mar Ecol Prog Ser*. 1989; 55:47–54.
50. Bayne BL, Newell RC. Physiological Energetics of Marine Molluscs. In: *The Mollusca*. 1983. p. 407–515.