

1 **Plankton lifeforms as a biodiversity indicator for regional-scale assessment of pelagic habitats for policy**

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19
20 **Abstract**

21 Plankton are sensitive indicators of change and, at the base of marine food webs, they underpin important
22 ecosystem services such as carbon sequestration and fisheries production. In the UK and the Northeast Atlantic
23 region, change in plankton functional groups, or ‘lifeforms’, constructed based on biological traits, is the formally
24 accepted policy indicator used to assess Good Environmental Status (GES) for pelagic habitats under the Marine
25 Strategy Framework Directive (MSFD: 2008/56/EC). To identify changes in UK pelagic habitats, plankton lifeforms,
26 were used from diverse UK data sets collected by different methods, including plankton sampling by nets, water
27 bottles, integrating tube samplers, and the Continuous Plankton Recorder. A Plankton Index approach was used to
28 identify change in plankton lifeforms. This is the first time that the pelagic plankton community has been assessed
29 on a UK-wide scale and forms the foundation of the UK’s 2020 MSFD Assessment for pelagic habitat biodiversity and
30 food webs. This approach revealed that some of the plankton lifeforms used in the assessment displayed spatially-
31 variable changes during the past decade. Assessing plankton community change using a common indicator at the UK
32 scale for the first time is a significant step towards evaluating GES for European seas. Determining GES for pelagic
33 habitats, however, is a challenging process, with additional work required to interpret the assessment results and to
34 identify causation of the changes observed.

35 **Key words**

36 Functional groups, ecosystem approach, Marine Strategy Framework Directive, Good Environmental Status, plankton
37 traits

38 **1.1 Introduction**

39 The Ecosystem Approach (EA; Secretariat of the Convention on Biological Diversity, 2004) and Ecosystem-Based
40 Management (EBM; Katsanevakis et al., 2011) are high-level strategies that are increasingly influencing management
41 of marine systems for sustainability and social equity. The European Union’s Marine Strategy Framework Directive
42 (MSFD; 2008/56/EC) is a large-scale example of this holistic style of management. The MSFD requires European seas
43 to achieve Good Environmental Status (GES). An integral part of assessing GES and ensuring that it is maintained is

44 the establishment of environmental targets and indicators of ecosystem state (Claussen et al., 2011). The Directive is
45 a complex, adaptive, and ambitious policy, whose scientific and operational implementation will evolve and adapt
46 throughout its lifetime. Like all Member States, the United Kingdom (UK) is required to assess the state of pelagic
47 habitat biodiversity in its national waters, and to contribute to the MSFD regional-scale assessment, led by the
48 Convention for the Protection of the Marine Environment of the North-East Atlantic (OSPAR) in the Northeast
49 Atlantic.

50 The MSFD requires the monitoring of community-level plankton indicators in support of environmental targets for
51 criteria in its biodiversity and food web descriptors (Table 1; European Commission, 2010; European Commission,
52 2017). Plankton are the foundation of most pelagic and benthic food webs, supporting a range of key ecosystem
53 functions including carbon sequestration and energy flow to higher trophic levels, including species of commercial
54 importance to humans, such as fish (Falkowski et al., 2004). They have also been described as “beacons of climate
55 change” due to their short lifespans, temperature-dependent physiology, and high potential for dispersal (Hays et
56 al., 2005; Richardson, 2008). Furthermore, because most plankton species are not heavily exploited commercially,
57 change in plankton abundances is a direct response to environmental pressures. Because the time-series coverage of
58 plankton in the North Atlantic and fringing shelf seas is exemplary in its spatial and temporal extent (see O'Brien et
59 al., 2017), plankton time-series provide an opportunity to tease apart the prevailing footprint of climate change on
60 ecosystems from other pressures, for example, nutrient loading and fishing. Accordingly, plankton time-series are
61 increasingly used to inform marine policy and management (McQuatters-Gollop et al., 2015; McQuatters-Gollop et
62 al., 2017), as well as for fundamental understanding of marine food webs (Beaugrand and Kirby, 2018).

63 The UK has defined its MSFD target for the pelagic habitat to achieve ‘Good Environmental Status’ as *the plankton*
64 *community is not significantly adversely influenced by direct anthropogenic pressures* at the scale of the two MSFD
65 sub-regions that include UK seas. These two sub-regions are the Greater North Sea (OSPAR region II) and the Celtic
66 Seas (OSPAR region III). Detecting changes in planktonic communities, and then attributing them either to climate
67 change or to directly manageable human pressures, such as fishing or nutrient enrichment, is not a trivial task. There
68 are two reasons for this. The first relates to sample collection and analysis. Although multiple plankton time-series
69 exist in Europe (O'Brien et al., 2017), differences in sampling methods, levels of taxonomic identification, and
70 methods of taxa enumeration, even within Member States (see for example Eloire et al., 2010; Richardson et al.,
71 2006; Whyte et al., 2017; Widdicombe et al., 2010) limit the direct comparability of the data, and utilising these
72 different time-series to deliver assessments at the MSFD sub-region level represents a significant technical
73 challenge.

74 The second reason concerns the dynamic nature of the plankton. Species of plankton are adapted to the
75 ecohydrodynamic conditions of the water bodies within which they live. As a consequence, the ‘patchwork’ of
76 different hydrodynamic regimes found in north western European waters (van Leeuwen et al., 2016), gives rise to
77 spatial variation in the abundance and diversity of plankton and the species that contribute to the plankton at the
78 spatial scale of MSFD reporting regions and/or sub-regional scales (Gowen et al., 1998; Pingree et al., 1978).
79 Furthermore, the inherently variable environment experienced by the plankton, coupled with the short generation
80 time of some taxa (e.g. \leq day) influences the abundance of individual species and hence the composition of the
81 plankton over a range of temporal scales.

82 Plankton indicators have been developed and utilised under previous European environmental directives such as the
83 Urban Waste Water Treatment Directive (91/271/EEC) and the Water Framework Directive (2000/60/EC). While
84 these have explored aspects of diversity and community structure as part of indicator development, the Directives
85 focus on nutrient enrichment and eutrophication (Devlin et al., 2009; Gowen et al., 2008) and have not been used in
86 biodiversity assessments, and also do not consider zooplankton. Plankton biodiversity indicators can be constructed
87 from data at varying taxonomic scales, with each option possessing benefits and compromises (McQuatters-Gollop
88 et al., 2017). Single plankton species have long been used as indicators (Beaugrand, 2005) but tend to focus on

89 specific questions, e.g. the abundance of *Calanus finmarchicus* as an assessment of the amount of food available for
90 cod larvae. Furthermore, single species indicators do not assess the diversity of the whole plankton community.
91 There is also the problem that there are no individual species of plankton that can be used to represent the state of
92 the plankton as a whole. In contrast, diversity indices, composed of abundances of all species in a region, attempt to
93 capture the diversity of the plankton community. Diversity indices, however, were developed based on ecological
94 principles relevant to terrestrial ecology. Such indices are difficult to construct with plankton data based on light
95 microscopy due to difficulties of identification and cryptic speciation (species that look the same under a
96 microscope) within the plankton community (Appeltans et al., 2012), and are highly influenced by sampling effort
97 (Stoetaert and Heip, 1990) and the identification of rare species (Lindeque et al., 2013). Finally, Tett et al. (2013, and
98 references cited therein) point out that most meta-studies failed to find relationships between standard species
99 diversity measures and ecosystem functions that are consistent across ecosystems and concluded that functional-
100 group diversity is the key component of ecosystem structure.

101 Multiple characteristics of the plankton are required to assess the status of the plankton community. One such
102 approach (Tett et al., 2008; Tett et al., 2013) uses a more theoretically-based approach to ‘package’ the available
103 information by grouping species into lifeforms, or functional groups, analogous to the guilds of species used by
104 benthic ecologists (Bremner et al., 2004; Bremner et al., 2003). A lifeform is a group of species (not necessarily
105 taxonomically related) that carry out the same important functional role in the marine ecosystem. For example,
106 diatoms as a group of species have a functional role related to silicon cycling. Metrics based on functional traits are
107 more closely linked to ecosystem structure and functioning than those based on single species or number of species
108 (Litchman et al., 2007; Mouillot et al., 2013; Stuart-Smith et al., 2013; Villéger et al., 2008). Indicators based on a
109 functional group approach have been shown to provide a useful means of describing plankton community structure
110 and biodiversity (Estrada et al., 2004; Gallego et al., 2012; Garmendia et al., 2012; Mouillot et al., 2006) and have
111 been used to assess community response to pressures such as nutrient enrichment (Gowen et al., 2015; Tett et al.,
112 2008) and climate change (Beaugrand, 2005). Indicators based on plankton lifeforms address the above challenges
113 and can be used to examine change in plankton communities based on multiple datasets with different taxonomic
114 resolutions (Gowen et al., 2015; Tett et al., 2008). Plankton lifeform indicators have thus been developed to inform
115 the biodiversity and food webs MSFD Descriptors (Table 1).
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Table 1: MSFD Descriptors for determining GES, relevant to the pelagic habitat. *Quoted text in italics.*

| Descriptor (Annex I of the MSFD) | |
|--|--|
| Relevant criteria (European Commission, 2010, part B) | Relevant criteria (European Commission, 2017, Annex) |
| 1. Biodiversity <i>Biological diversity is maintained. The quality and occurrence of habitats and the distribution and abundance of species are in line with prevailing physiographic, geographic and climatic conditions.</i> | |
| At the habitat level, assessment includes habitat distribution and extent, plus 1.6. <i>Habitat condition</i> including condition and relative abundance of the typical species and communities, and 1.7. <i>Ecosystem structure – composition and relative proportions of ecosystem components (habitats and species)</i> | <i>D1C6 the condition of each broad habitat type, including its biotic and abiotic structure and its functions (e.g. its typical species condition and their relative abundance, absence of particularly sensitive or fragile species, providing a key function, size structure of species) is not adversely affected due to anthropogenic pressures.</i> |
| 4. Food webs <i>All elements of the marine food webs, to the extent that they are known, occur at normal abundance and diversity and levels capable of ensuring the long-term abundance of the species and the retention of their full reproductive capacity.</i> | |
| <i>This descriptor concerns important functional aspects such as energy flows and the structure of food webs (size and abundance) and the criteria include:</i> <i>4.3 Abundance/distribution of key trophic groups/species – Abundance trends of functionally important selected groups/species.</i> | The relevant ‘trophic guilds’ are phytoplankton and zooplankton (ICES, 2015); the criteria are: <i>D4C1 The diversity (species composition and their relative abundance) of the trophic guild is not adversely affected due to anthropogenic pressures.</i> <i>D4C2 The balance of total abundance between the trophic guilds is not adversely affected due to anthropogenic pressures.</i> <i>DC43 The size distribution of individuals across the trophic guild is not adversely affected due to anthropogenic pressures.</i> |

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This paper describes a preliminary and novel assessment of changes in the plankton communities found in UK waters via the OSPAR common plankton lifeform indicator (PH1/FW5: Changes in Phytoplankton and Zooplankton Communities). This is the first time that the plankton found in UK waters have been examined at a regional scale using a consistent method applied to a diverse suite of datasets. This assessment represents an important step towards determining GES for pelagic habitats and will contribute to the UK’s formal 2020 assessment for the MSFD. We explore the initial results and some of the challenges that remain and outline the additional requirements to determine whether UK pelagic habitats are in GES.

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2.1 Methods

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2.1.1 Addressing spatial variability of UK pelagic habitats

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UK waters are ecologically and physically heterogeneous and cannot be considered as one uniform system even within individual MSFD sub-regions (van Leeuwen et al., 2015; van Leeuwen et al., 2016). Furthermore, plankton taxa are adapted to live in different hydrodynamic conditions (Margalef, 1978), so that plankton community

composition, distribution, and dynamics are closely linked to environmental conditions (de Vargas et al., 2015; Jones et al., 1984; Williams et al., 1994). Using density stratification, an important large-scale physical feature in shallow shelf seas, UK waters were spatially partitioned into six “ecohydrodynamic” (EHD) regimes (Figure 1) (van Leeuwen et al., 2015). The main EHD zone types, based on a 50-year modelled hindcast of water-column structure, are:

- Permanently mixed throughout the year
- Permanently stratified throughout the year
- Regions of freshwater influence (ROFIs)
- Seasonally thermally stratified (for approximately half the year, including summer)
- Intermittently stratified
- Indeterminate regions (inconsistently alternate between the above).

UK EHD zones were divided into North Sea and Celtic Sea zones for this analysis in order to align with the OSPAR Greater North Sea (OSPAR Region II) and Celtic Seas (OSPAR Region III) sub-regions. A more highly resolved EHD model exists for the North Sea than the Celtic Seas (van Leeuwen et al., 2015; van Leeuwen et al., 2016), and therefore the zoning might be less reliable in the case of the Celtic Seas and western English Channel.

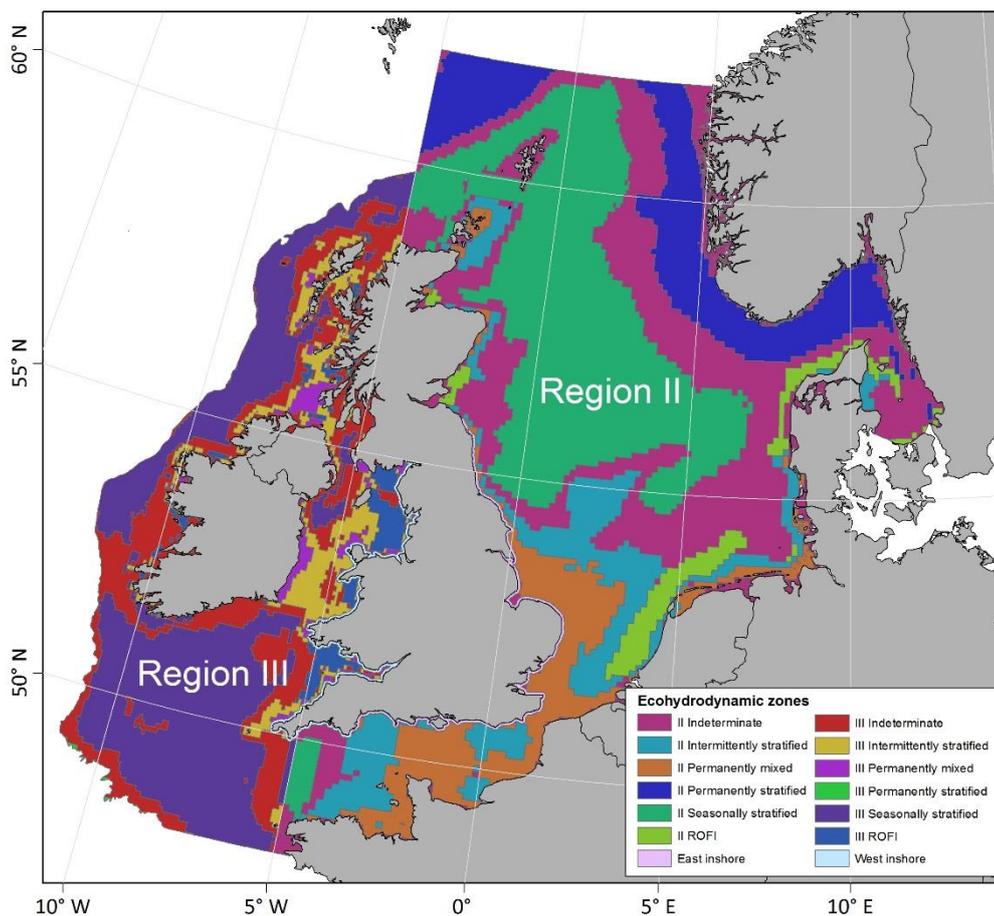


Figure 1: Map of ecohydrodynamic (EHD) zones in the Greater North Sea (OSPAR Region II) and Celtic Seas (OSPAR Region III), coloured by EHD type and region number. EHD zones were constructed based on key simulated water column features, which are important to plankton community structure and dynamics. The main EHD zone types, based on water-column structure, are 1) Permanently mixed throughout the year, 2) Permanently stratified throughout the year, 3) Regions of freshwater influence (ROFIs), 4) Seasonally thermally stratified (for about half the year, including summer), 5) Intermittently stratified and 6) Indeterminate regions (inconsistently alternate between the above levels of stratification). East and west inshore (>1 nm from shore) regions are also shown here, although they were not identified from simulations.

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160 As there is no coastal EHD type, the very near-coast (< 1nm from shore) regions have been divided into east and
161 west coastal inshore EHD zones. The hydrodynamic model Figure 1 indicates an 'indeterminate' type in the inshore
162 waters of the Scottish highlands and islands. However, observations (e.g. Inall and Gillibrand, 2010; Wood et al.,
163 1973) show that salinity-stratification and associated density-driven circulation are common here. For this reason a
164 fjordic system EHD type was used for sea lochs on the west coast of Scotland.

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166 The UK plankton monitoring programme (Figure 2) consists of coastal, fixed-point sampling stations including PML L4
167 (Atkinson et al., 2015), CEFAS SmartBuoys (Weston et al., 2008), Environment Agency (EA) Water Framework
168 Directive (WFD) monitoring stations (UKTAG, 2014), Scottish Environmental Protection Agency WFD monitoring
169 stations, Agri-Food and Biosciences Institute monitoring stations (Gowen and Stewart, 2005), the Firth of Lorne
170 Observatory (Tett, 1973; Tett and Wallis, 1978; Whyte et al., 2017), the Scottish Coastal Observatory (Bresnan et al.,
171 2016), and the offshore Continuous Plankton Recorder (CPR) survey (Richardson, 2008) (see Figure 2). These various
172 sources of data provide complementary information, with the CPR data illustrating regional and long-term change
173 and the fixed-point stations providing detailed information at higher time and depth resolution at a local scale. EHD
174 zones provide a spatial framework by which to use these two types of data together. CPR data and EA coastal
175 sampling network were thus aggregated at the EHD zone scale, allowing comparability between CPR and fixed-point
176 results in the same EHD zone. Because EHDs are constructed based on the dominant hydrodynamic features of the
177 water column, this approach also enables data from one part of an EHD zone to be used for the whole of that EHD
178 zone (Scherer et al., 2014). In other words, features of the plankton community at a fixed-point station in a particular
179 EHD zone are assumed to be representative of the plankton community throughout that EHD zone.

180 181 *2.1.2 Plankton lifeform construction*

182 The UK plankton monitoring programme consists of surveys from a variety of government agencies and research
183 organisations. They employ sampling techniques ranging from collections at fixed (buoys or moorings) time-series
184 stations using nets, tubes integrating the top 10m of the water column, and water bottles to the Continuous
185 Plankton Recorder survey, a large scale plankton monitoring programme which uses ships of opportunity (Figure 2)
186 (Bean et al., 2017). All these surveys contribute towards a large quantity of UK plankton data, however, variation in
187 sampling methods, levels of taxonomic identification, and methods of taxa enumeration provide a challenge to UK-
188 level assessments.

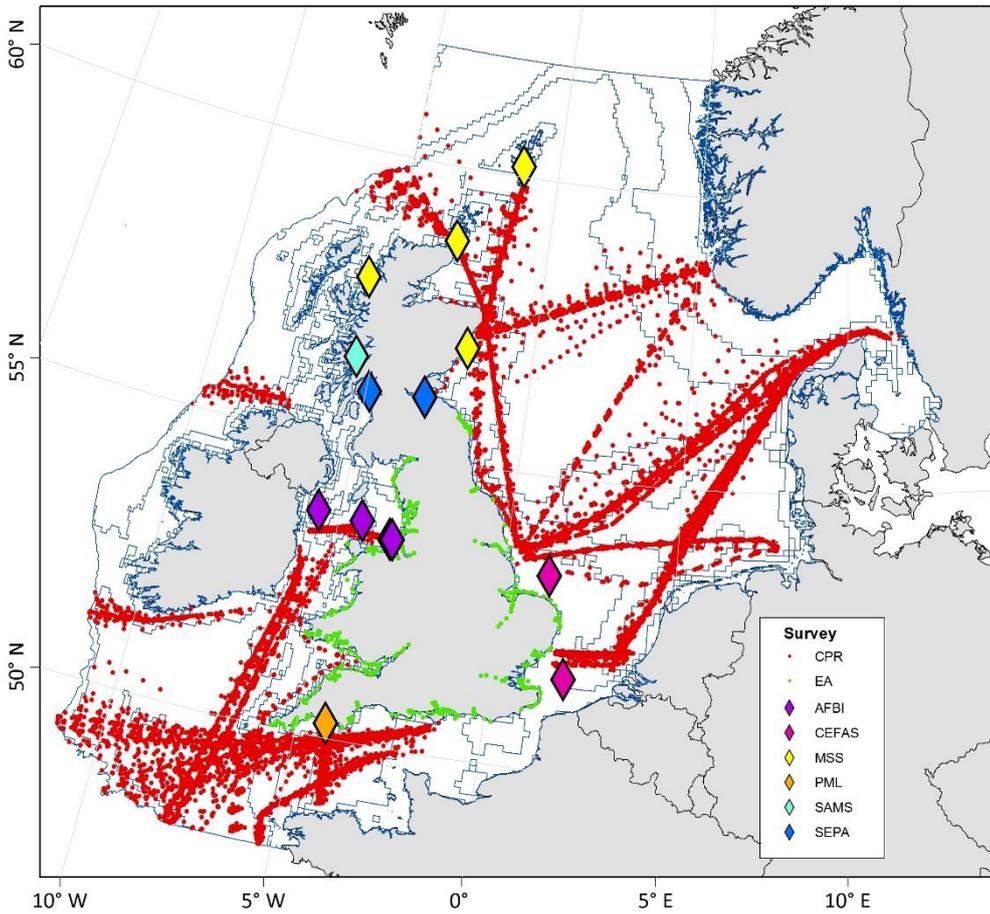


Figure 2: The UK plankton monitoring programme consists of disparate but complementary surveys. Samples from the Continuous Plankton Recorder (CPR) are displayed as red dots along routes; samples represent 10 nautical miles of water. The other surveys operate fixed-point samplings schemes. Abbreviations: AFBI – Agri-Food and Biosciences Institute; EA – Environment Agency; PML – Plymouth Marine Laboratory; MSS – Marine Scotland Science; SAMS – Scottish Association for Marine Science; Cefas - Centre for Environment, Fisheries and Aquaculture Science; and SEPA – Scottish Environmental Protection Agency.

To address this and provide a holistic view of the UK plankton, an indicator based on plankton lifeforms was developed which allows the use of all plankton datasets, regardless of differences in sampling or analysis techniques. To construct the plankton lifeform indicator, plankton taxa were grouped into lifeforms based on traits such as size, trophic, motility, and other key biological features (Table 2, 3; Litchman et al., 2012; Litchman and Klausmeier, 2008). Taxa can be assigned multiple traits, and can be included in multiple lifeforms. In instances where the trait of a taxon was unknown, the taxon was omitted from lifeforms constructed with that particular trait. Because plankton lifeforms are constructed based on traits (Table 4) rather than on species-level information, grouping plankton taxa into lifeforms allows the use of plankton data identified at different taxonomic resolutions, which suits the UK's integrated but diverse plankton monitoring programme. Additionally, plankton lifeforms are aggregations of taxa and so are less likely to experience the extreme seasonal fluctuations of single species indicators. Finally, because lifeforms consist of multiple taxa with a similar functional role, spatial intercomparability is increased, as even though the particular taxa fulfilling a functional role may vary, the corresponding lifeform is often regionally ubiquitous. When examined in ecologically-relevant plankton lifeform pairs, plankton lifeforms can provide an indication of changes in different aspects of plankton community functioning such as energy flows, benthic-pelagic coupling, and food web structure (Table 4). The eight lifeform pairs were selected according to confidence in the traits corresponding to each lifeform and to reflect multiple features of the pelagic habitat. As the knowledge base increases or policy needs change, new plankton lifeform pairs can be developed, allowing us to address additional

215 and emerging scientific and policy questions about biodiversity, food webs, eutrophication, and responses to climate
 216 change. Given the emerging importance of community functioning as a key characteristic of biodiversity, all of the
 217 lifeform pairs in Table 4 contribute to the biodiversity and food web descriptors.

218 **Table 2: Plankton taxa were assigned traits based on our simple definition based on key biological features.**

| Trait type | Trait categories |
|--------------------------|---|
| Plankton type | Phytoplankton: protista taxa that contribute to primary production Zooplankton: metazoan taxa of the kingdom Animalia |
| Zooplankton type | Fish/eggs: taxa of the subphylum Vertebrata Copepod: taxa of the subclass Copepoda Gelatinous: taxa of the phylum Cnidaria and Ctenophora Crustacean: taxa of the Subphylum Crustacea |
| Phytoplankton type | Diatom: taxa of the class Bacillariophyceae Dinoflagellate: taxa of the phylum Dinoflagellata |
| Zooplankton trophic mode | Carnivore: taxa which prey on zooplankton Herbivore: predominately suspension or filter feeders Omnivore: includes both carnivorous and herbivorous feeding Ambiguous: diet uncertain |
| Habitat | Holoplankton: taxa which spend their entire lifecycle in the plankton Meroplankton: taxa which spend part of their lifecycle in the plankton Tychopelagic: benthic diatoms which can become mixed into the water column |
| Size | Large: phytoplankton ($\geq 20 \mu\text{m}$ diameter); zooplankton ($\geq 2 \text{ mm}$ adult body length) Small: phytoplankton ($< 19 \mu\text{m}$ diameter); zooplankton ($< 1.9 \text{ mm}$ adult body length) |

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220 **Table 3: Plankton lifeforms are comprised of taxa sharing the same traits.**

| Lifeform | Traits |
|-----------------------------|---|
| Diatoms | Plankton type = 'Diatom' |
| Dinoflagellates | Plankton type = 'Dinoflagellate' |
| Gelatinous zooplankton | Plankton type = 'Gelatinous' |
| Fish larvae/eggs | Zooplankton type = 'Fish' AND 'Eggs' |
| Non-carnivorous zooplankton | Plankton type = 'Zooplankton' AND Trophic mode = either 'Herbivore', 'Omnivore', OR 'Ambiguous' |
| Crustaceans | Zooplankton type = 'Crustacean' |
| Large phytoplankton | Plankton type = 'Phytoplankton' AND Size = 'Large' |
| Small phytoplankton | Plankton type = 'Phytoplankton' AND Size = 'Small' |
| Pelagic diatoms | Phytoplankton type = 'Diatom' AND Habitat = 'Holoplankton' |
| Tychopelagic diatoms | Phytoplankton type = 'Diatom' AND Habitat = 'Tychopelagic' |
| Holoplankton | Plankton type = 'Zooplankton' and Habitat = 'Holoplankton' |
| Meroplankton | Plankton type = 'Zooplankton' and Habitat = 'Meroplankton' |
| Large copepods | Zooplankton type = 'Copepod' AND Size = 'Large' |

| Lifeform | Traits |
|----------------|---|
| Small copepods | Zooplankton type = 'Copepod' AND Size = 'Small' |
| Phytoplankton | Plankton type = 'Phytoplankton' |

Table 4: Plankton lifeform pairs consist of two contrasting and ecologically-relevant plankton lifeforms. The rationale behind their selection is also described.

| Lifeform pairs | Ecological rationale |
|--|--|
| Diatoms and dinoflagellates | Systems receiving high nutrient input are often dominated by dinoflagellates at the expense of diatoms (McQuatters-Gollop et al., 2009). In the North Atlantic, stratification plays a key role in structuring phytoplankton communities with dinoflagellate abundances connected to increased stratification while diatoms are better suited to mixed waters (Barton et al., 2015). Change in the relative abundance of the two plankton lifeforms can therefore indicate changes in nutrient and stratification regimes. |
| Pelagic diatoms and tychopelagic diatoms | Benthic disturbance, such as from development or storms, can resuspend tychopelagic (benthic) diatoms in the water column (Ubertini et al., 2012). A shift in the proportion of tychopelagic and pelagic diatoms can therefore indicate changes in the magnitude and frequency of benthic disturbance and resuspension events. |
| Large microphytoplankton ($\geq 20 \mu\text{m}$ diameter) and small microphytoplankton ($< 19 \mu\text{m}$ diameter) | Organism size is a key factor in energy transfer efficiency in pelagic habitats and may determine the system's potential to support higher trophic levels (Fox and Pitois, 2006; Thiebaut and Dickie, 1993). Changes in the relative abundance of large microphytoplankton ($\geq 20 \mu\text{m}$ diameter) and small microphytoplankton ($< 19 \mu\text{m}$ diameter) can therefore indicate alterations in energy flow to higher trophic levels. |
| Microphytoplankton and non-carnivorous zooplankton | Non-carnivorous zooplankton graze on microphytoplankton, thereby transferring energy from single-celled algae to metazoan animals. Changes in the relative abundance of the two plankton lifeforms can therefore indicate changes in energy flow through the pelagic food web. |
| Small copepods ($< 1.9 \text{ mm}$) and large copepods ($\geq 2 \text{ mm}$) adult body length | Copepods are a key food resource for higher trophic levels, including commercially important fish such as larval cod, whose survival is linked to the mean size of their prey (Beaugrand et al., 2003). A change in the proportion of large ($\geq 2 \text{ mm}$ in length) and small ($< 1.9 \text{ mm}$ in length) adult copepods can therefore indicate changes in food web structure (Capuzzo et al., 2018; Fox and Pitois, 2006). |
| Holoplankton and meroplankton | Meroplankton only spend a part of their lifecycle within the pelagic realm, and for their most part, are the larvae of benthic organisms. A change in the proportion of meroplankton and holoplankton (plankton spending their whole lifecycle within the pelagic realm) can indicate a change in the strength of benthic and/or pelagic production with consequences for pelagic-benthic coupling (Kirby et al., 2008; Lindley et al., 1995). |
| Crustaceans and gelatinous zooplankton | Gelatinous organisms within the plankton can have an important predatory effect on other crustacean plankton and fish larvae when abundant, thereby acting as a pressure on fish populations. A change in the relative abundance of crustaceans and gelatinous zooplankton can thus indicate a change from an ecosystem with numerous fish of commercial interest to an ecosystem dominated by gelatinous organisms of low commercial interest (Kirby et al., 2009; Purcell and Arai, 2001; Richardson et al., 2009). |

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| Gelatinous zooplankton and fish larvae/ eggs | Gelatinous organisms within the plankton can have an important predatory effect on other crustacean plankton and fish larvae when abundant, thereby acting as a pressure on fish populations. A change in the relative abundance of fish larvae/eggs and gelatinous zooplankton can thus indicate a change from an ecosystem with numerous fish of commercial interest to an ecosystem dominated by gelatinous organisms of low commercial interest (Kirby et al., 2009; Purcell and Arai, 2001; Richardson et al., 2009). |
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2.1.3 Identifying change in plankton lifeforms

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A 'Plankton Index' (PI) has been used to identify temporal change within plankton lifeform pairs. This approach (Gowen et al., 2011; Scherer et al., 2014; Tett et al., 2008) identifies change plankton lifeform pairs from a starting period, usually at the beginning of a time-series, although the PI has been used to hindcast (Gowen et al., 2015) and compare changes in plankton in response to human pressure in different regions of the same ecohydrodynamic regime (Scherer, 2012). Based on general systems theory (von Bertalanffy, 1972), a sample's position at any point in time is defined in "state space" by orthogonal axes of (log-transformed) lifeform abundance. For convenience and ease of visualisation, the axes are plotted two at a time, so that, for example, a sample's horizontal co-ordinate is diatom abundance and its vertical co-ordinate is dinoflagellate abundance (Figure 3).

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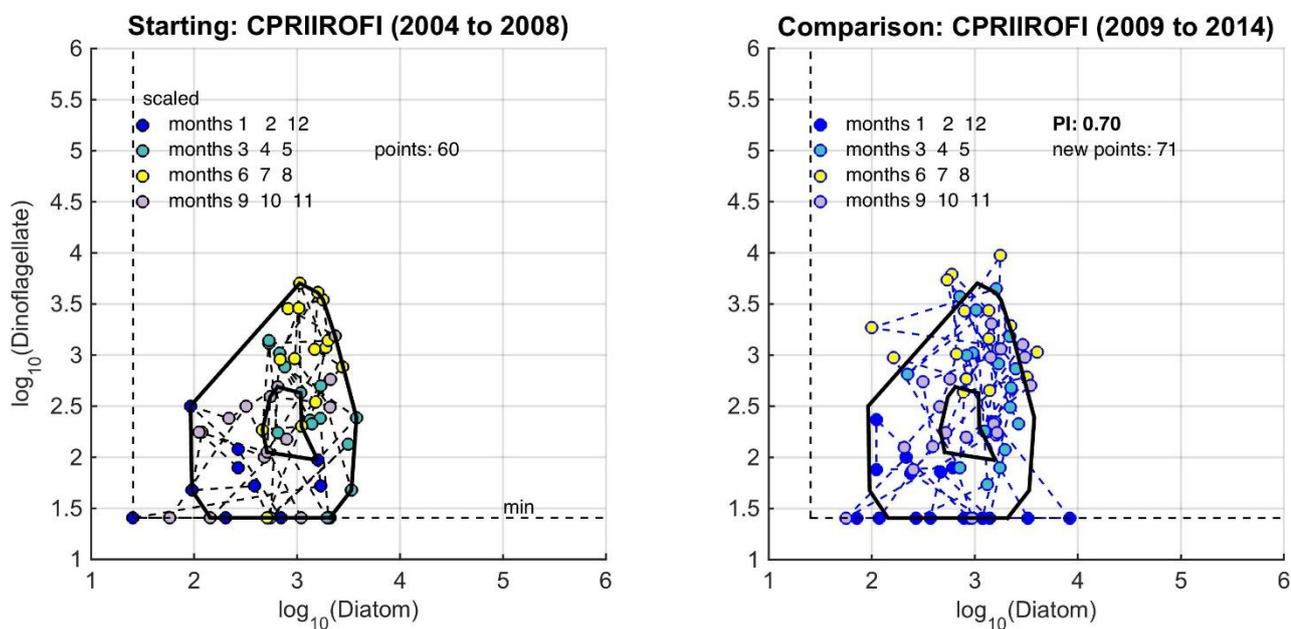
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To define the reference boundary, an envelope is drawn around several years of points representing monthly samples (Figure 3); here we used a 5-year period. Monthly averaged data from subsequent periods are then plotted in the same state space, and a Plankton Index (PI), and associated probability value, calculated as the proportion of new points falling within the reference boundaries. A PI value approaching 1 indicates no difference in plankton communities while a PI value approaching 0 indicates a complete change in plankton communities between the two time periods. Low PI values across spatially disparate datasets mean that wide scale changes in the plankton community (e.g. from climate change) can be identified. The PI approach is flexible in nature, allowing both abundance and biomass data to be used, and furthermore it is relatively robust to periods without data collection, making it ideal for identifying change in plankton communities when assessing environmental state by using multiple disparate datasets. Although originally developed to track change in phytoplankton communities (Tett et al., 2008), the PI has been adapted to also incorporate changes in zooplankton, making this a method to assess change in the plankton community more holistically.

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248 **Figure 3: An example diatoms v dinoflagellate comparison between starting and contemporary conditions for Regions of**
249 **Freshwater Influence (ROFIs) in the North Sea (OSPAR Region II).** Left: The starting conditions envelope, outlined in black, was
250 created using sampling points from 2004-2008. Right: points from the 2009-2014 UK 2020 MSFD Assessment period (n=71) are
251 overlain on the starting conditions envelope. The PI value of 0.70 suggests a statistically significant difference between the two
252 time periods (binomial $p < 0.01$), caused by 21 of the 71 assessment period points falling outside the bounds of the starting
253 conditions envelope. The distribution of the points in the assessment period suggests an increase in dinoflagellates in summer
254 months.

255 The PI value was calculated for all lifeform pairs for each fixed-point sampling station (with sufficient data) and for
256 CPR data aggregated across each EHD zone. For the UK 2020 MSFD Assessment, the period 2004 to 2008 was
257 selected to represent starting conditions to align with the starting condition period used in the OSPAR Intermediate
258 Assessment 2017. This starting period selection was therefore driven by a policy rather than scientific requirement, a
259 point we discuss later. The starting condition envelope was compared with data from the subsequent six-year MSFD
260 Assessment period (2009 to 2014), also chosen for its alignment with the MSFD assessment and reporting cycle.
261 From a policy perspective, this strategy facilitated comparability between the UK-level and OSPAR-level analyses and
262 allowed the examination of change in UK plankton with respect to regional scale plankton change, as identified
263 through the 2017 OSPAR Intermediate Assessment. Most importantly, alignment of the starting condition periods
264 allowed the examination of plankton change on the MSFD policy timescale, a key goal of the UK 2020 MSFD
265 Assessment to which this work will contribute. Here we have expanded the number of UK datasets beyond those
266 used in the UK 2020 MSFD Assessment to include all UK plankton time-series with data spanning the same 2004-
267 2014 time period. The datasets from Scottish Environmental Protection Agency (SEPA) and the Agri-Food and
268 Biosciences Institute, Northern Ireland (AFBI), however, did not cover the full duration of the starting conditions
269 period and were therefore excluded from this analysis.

270 **3.1 Results**

271 *3.1.1 A first assessment of changes in UK plankton*

272 Differences in the Plankton Index between starting conditions (2004-2008) and current conditions (2009-2014) were
273 calculated for all lifeform pairs where monthly data were available during the entire time period (Figure 4). This first
274 analysis showed similarities and differences in PI values from different EHD zones and between lifeform pairs. Of the
275 91 differences identified, 78 were statistically significant (Figure 4), suggesting alterations to the UK plankton
276 community between the starting and MSFD assessment periods. Further interpretation of these results (including
277 timing and dominance of plankton lifeforms and an investigation to the significant contributing species) were not
278 included in the MSFD assessment and are therefore beyond the scope of this current paper.

279

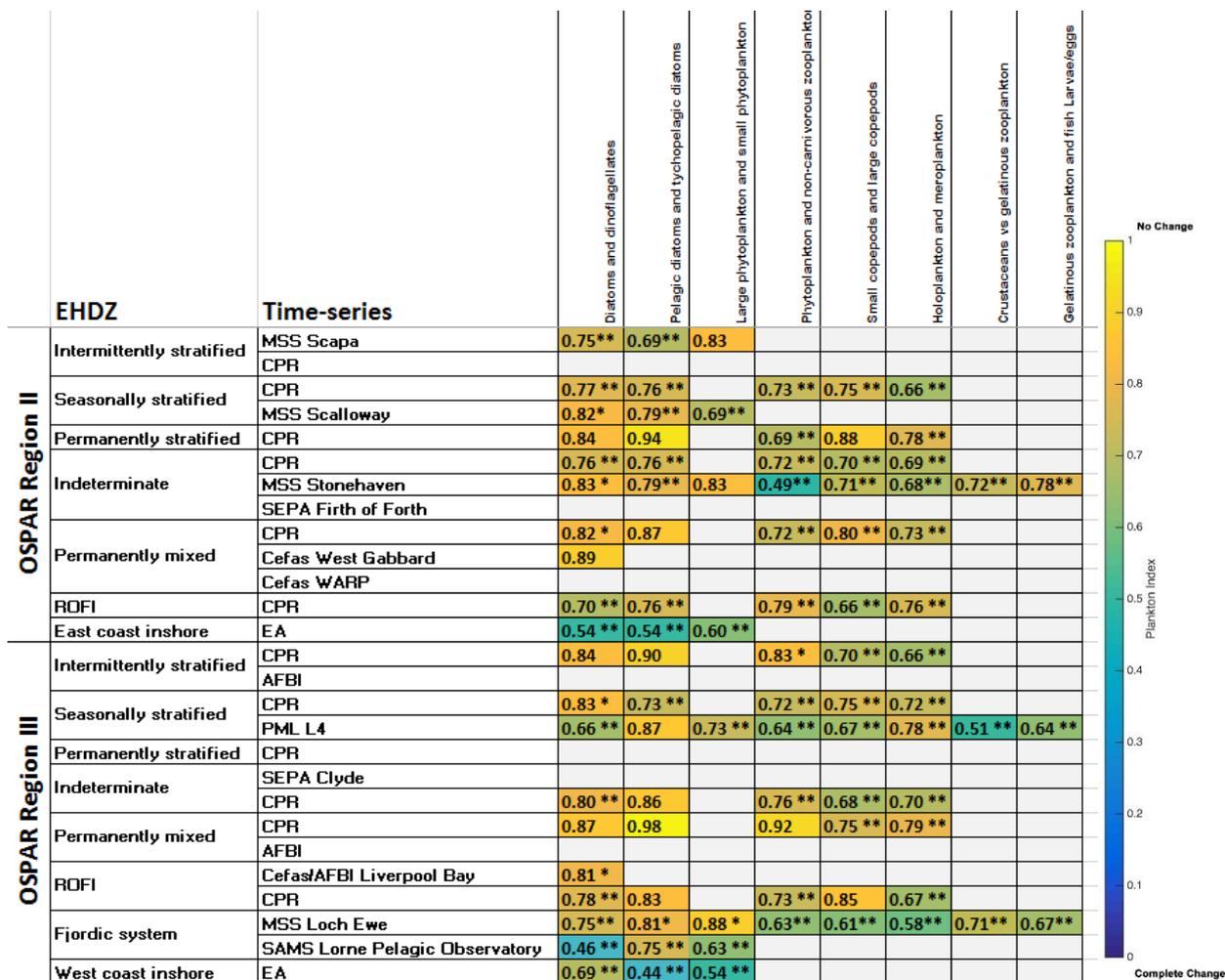
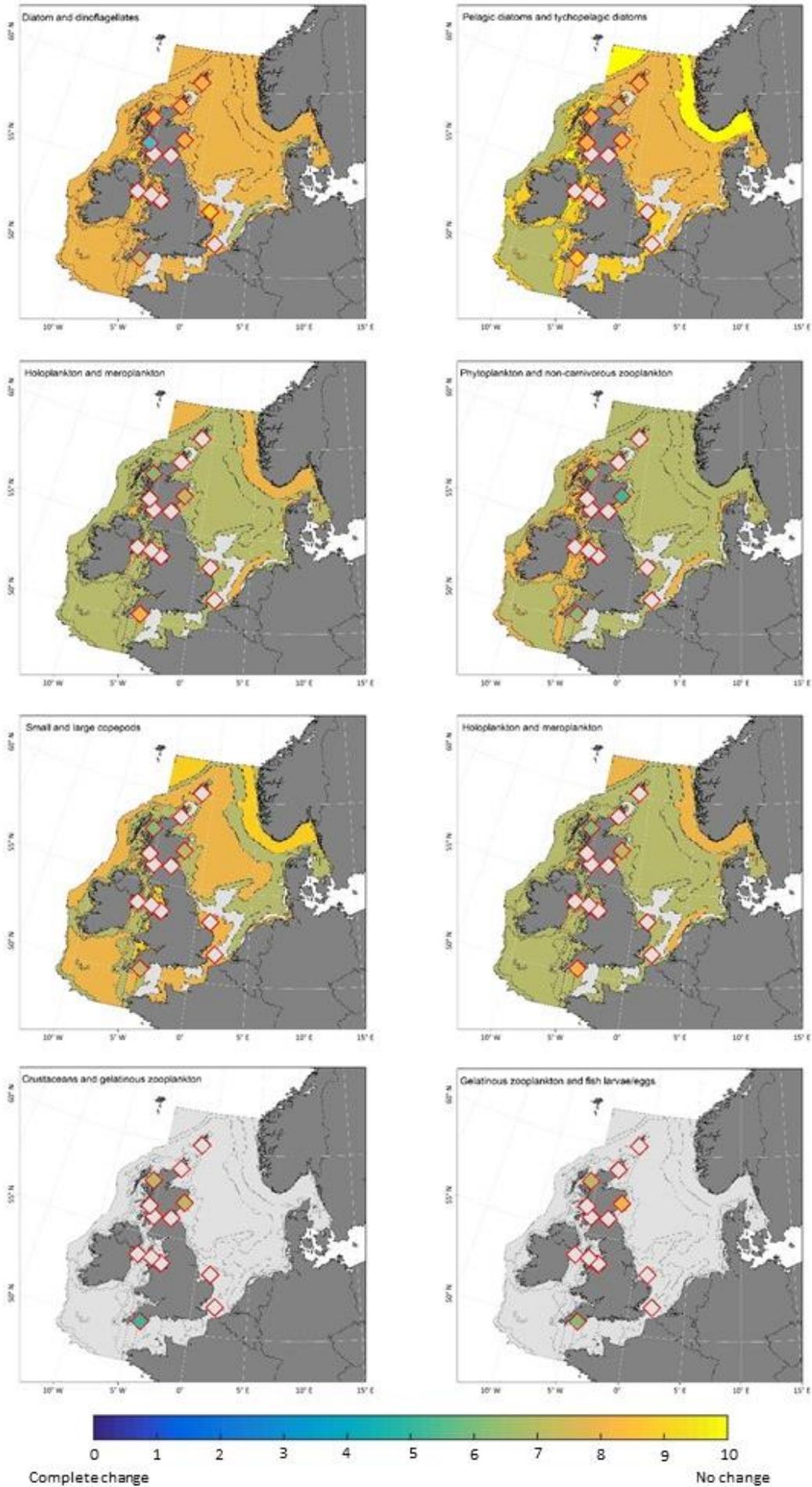


Figure 4: Plankton Indices, by OSPAR region and EHD zone, for the period 2009 - 2014 from starting conditions (2004 – 2008). Starred cells indicate theoretical significant change (*p<0.05; **p<0.01) from starting conditions. A Plankton Index approaching '1' (bright yellow cells) denotes no change from starting conditions while an Index approaching '0' (dark blue cells) represents complete change. White shading represents where data were insufficient to determine a Plankton Index. UK datasets with incomplete data during the starting conditions period were not used in the analysis, but all existing sampling stations are included to demonstrate the future potential of the monitoring program.

The degree of difference in PI value was spatially variable within each lifeform pair (Figure 5) although, in some cases, remarkable similarity between surveys exists. Of the lifeform pairs sampled for most datasets (n >12 datasets) holoplankton and meroplankton (range = 0.21) as well as small and large copepods (range = 0.27) had the smallest ranges in PI, indicating the highest levels of spatial harmony (Figure 4, Figure 5). The lifeform pair with the greatest variability was pelagic diatoms and tyhopelagic diatoms (range = 0.54), with the greatest difference between the starting and assessment period found in the west coast inshore EHD zone (PI = 0.44, p<0.01). Other than the highly dynamic west and east coast inshore zones, the most extreme differences from starting conditions of any lifeform pairs were observed in Scotland and the Western Channel, with phytoplankton and non-carnivorous zooplankton at Stonehaven (PI = 0.49, p < 0.01), and diatoms and dinoflagellates at Lorne (PI = 0.46, p < 0.01).



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301 **Figure 5: Plankton Index for each lifeform pairs in UK waters.** Changes within EHDs are based on CPR data, with fixed-point
302 stations overlain (red borders). Points or EHD zones in grey lack complete data during starting conditions and/or assessment
303 period for particular lifeform pairs. Results for non-UK EHDs are also displayed as they enable regional interpretation of UK
304 plankton dynamics.

305 Results between the near-shore fixed-point stations and CPR data in the same EHD zone were broadly consistent
306 (Figure 4, Figure 5), suggesting spatial congruence between the two survey types. For example, the results of PML L4,
307 located in the seasonally stratified Celtic Sea OSPAR Region III, were consistent with results from the CPR in the same
308 EHD zone, particularly for the pairs with zooplankton lifeforms. Similarly, the results from Stonehaven, located in the
309 indeterminate North Sea zone, matched well with CPR results from the same EHD.

310 **4.1 Discussion**

311 *4.1.1 Change in plankton lifeform indicator*

312 It has previously been established that the UK plankton community has undergone significant changes during the
313 past six decades (Beaugrand, 2004). Changes include phenological alterations (Atkinson et al., 2015; Edwards and
314 Richardson, 2004; Whyte et al., 2017), shifts in the balance of organisms (Beaugrand et al., 2002; Gregory et al.,
315 2009; Whyte et al., 2017), and spatially variable changes in phytoplankton biomass and chlorophyll (McQuatters-
316 Gollop et al., 2011; Whyte et al., 2017). Assessments of estuarine and coastal phytoplankton metrics have also been
317 carried out under the requirements of the WFD, but focussed on changes in the total taxa counts and most
318 numerous species (Devlin et al., 2009; Devlin et al., 2007). An integrated, region-wide view of plankton change
319 assessed using a common indicator and all available UK datasets, however, has been lacking. The case study
320 presented here illustrates the value of the plankton lifeform approach in connecting disparate geographic areas with
321 diverse methods of plankton sampling and analysis, using a common and comparable indicator. This is the first
322 application of this indicator across multiple plankton datasets throughout UK marine waters, illustrating change
323 between two periods examined for the UK MSFD 2020 Assessment.

324 Harmony in results between fixed-point datasets and the CPR survey highlights the complementarity of the datasets
325 comprising the UK's plankton monitoring programme (Figure 4, Figure 5). For example, results from PML L4 and the
326 CPR survey are particularly well-matched for pairs with zooplankton lifeforms and are also in line with previous work
327 showing that zooplankton seasonal cycles captured by the two time-series were similar, even though absolute
328 abundances differed (John et al., 2001; Ostle et al., 2017). The similarity in PI values between CPR and fixed-point
329 time-series suggests both are representative of EHD zones, but further validation between CPR and fixed-point data
330 from the same EHD zones are needed. Better spatial representivity exists for EHD zones which are monitored by
331 CPR routes compared to locations with only a fixed-point station, though some inshore fixed-point stations (PML,
332 MSS Stonehaven, MSS Loch Ewe) are monitored weekly and so better reflect temporal variability. Some of the EHD
333 zones are spatially large and thus averaging over such a large spatial scale may dampen or mask variability. EHD
334 zones with both CPR data and fixed-point stations have the most comprehensive and robust information. The
335 stations closest to shore, the east and west inshore EHDs and SAMS Lorne Pelagic Observatory, displayed some of
336 the most extreme differences in PI values, suggesting that coastal waters are more temporally variable than waters
337 further offshore. In the case of the east and west inshore EHDs, however, some of this variability may be caused by
338 changes to the sampling programme as mentioned above. These preliminary results show that UK plankton lifeforms
339 displayed spatially-variable changes during the past decade with greater depth of knowledge obtained by the
340 merging of many UK plankton datasets.

341 This study constitutes a first step in evaluating GES for UK waters by documenting widespread change. There is work
342 to be done in establishing the causes of change, which might include (i) the intrinsic inter-annual and decadal scale

343 cyclical variability common to many Earth systems; (ii) the longer-term effects of global change, especially that
344 associated with climate; or (iii) the superimposed effects of manageable anthropogenic pressures such as nutrient
345 enrichment, fisheries disturbance, pollution or seabed disturbance on food webs. The UK definition of GES for the
346 pelagic habitat is essentially practical: if change in lifeform absolute and relative abundances (which can be signalled
347 by the PI) is attributed to increases in manageable pressures, then the habitat is not in GES and measures need to be
348 taken to ameliorate the pressures. Thus we have referred to the 2004-2008 period as 'starting' rather than
349 'reference' conditions as these years were chosen to fit with the MSFD policy assessment cycle rather than any
350 judgement of whether the condition of the pelagic habitat was in GES or not. Ideally, the envelope used to calculate
351 a value of the PI would be drawn around a set of points from a marine ecosystem known to be in GES. Scherer et al.
352 (2016) have proposed a method for determining pelagic GES independent of the PI tool, but in default of application
353 of this method to all EHD types in UK waters, the PI only provides an indication of change. However, such change in
354 PI can be used as a 'flag' to trigger further investigation into the pressures that may be causing this change in
355 ecosystem state.

356 *4.1.2 Further development of the lifeform indicator and Plankton Index approach for assessing Good Environmental* 357 *Status*

358 As an indicator of plankton functioning and structure, the lifeform approach enables the use of multiple datasets
359 with disparate methods of sample collection and taxonomic analysis. Our results demonstrate that data collected
360 from disparate monitoring programmes established for a variety of policy drivers (e.g., Water Framework Directive,
361 investigative monitoring and research, Urban Waste Water Treatment Directive) can also be used for the
362 construction of plankton lifeforms for use as a MSFD indicator. Because plankton lifeform datasets can be populated
363 with plankton data not collected specifically for informing the MSFD indicator, the use of this single regional
364 indicator promotes synergies between disparate UK plankton monitoring surveys. This approach, whilst innovative,
365 does require several more steps to increase its robustness, enable the best use of all available plankton data, and to
366 support future use of the indicator in other geographic areas. Each of these steps is a precursor to determining GES
367 for UK pelagic habitats.

368 EHD zones provide a way to define pelagic habitats and plankton communities, but the model used to construct the
369 EHD zones was developed for use in, and validated with data from, offshore pelagic environments and as a result
370 may not accurately simulate conditions in near-shore areas (van Leeuwen et al., 2015; van Leeuwen et al., 2016).
371 Observationally-informed designations of the seasonal stratification from fixed point stations in some regions such
372 as the Western English Channel do not always agree perfectly with the EHDs defined in Figure 1. In some cases, such
373 as the Irish Sea, it is likely that numerous EHD zones occur in a relatively small region of complex hydrography
374 (Gowen et al., 1995; Scherer et al., 2016) and so may need revisiting. In addition, some EHDs (e.g. North Sea
375 seasonally stratified) are large and span a latitudinal gradient of ~ 5 degrees, and thus phytoplankton may
376 experience differing light regimes between the northern and southern regions of this EHD. Nevertheless, we have
377 used the Figure 1 map as a single and traceable regional classification for all our analysis. Further refinement of
378 modelling in hydrodynamically complex areas and improvements in coupled catchment and marine models would
379 improve the delineation of EHD zones.

380 A consequence of the different methods used in the UK plankton monitoring programme is that there is some
381 inconsistency in the elements of the plankton community sampled. As a result, the full set of lifeform pairs (Table 4)
382 could not be derived from some data sets. Although all UK stations monitor phytoplankton, only the CPR and three
383 fixed-point stations have historically collected and analysed zooplankton samples. Additionally, not all surveys
384 sample all taxa equally well. The CPR, for example, inadequately captures small phytoplankton or gelatinous taxa
385 (Richardson et al., 2006) and so did not contribute to pairs containing these plankton lifeforms. Only three 'sentinel'
386 stations, MSS Stonehaven, MSS Loch Ewe, and PML L4, can address all lifeform pairs. Adding zooplankton sampling
387 to the remaining fixed-point stations would increase the robustness and form a 'sentinel network' providing detailed

388 insight into coastal plankton dynamics which is complementary to the CPR's large-scale, regional sampling. It should
389 also be noted that the smaller size portion of the pelagic assemblage, i.e. small nanoplankton, picoplankton, marine
390 bacteria, and viruses, are poorly monitored (McQuatters-Gollop et al., 2017). Additional consideration needs to be
391 given to taxa which are difficult to monitor or enumerate routinely, such as coccolithophores and mucilage-forming
392 *Phaeocystis*. In general, there is a need for some further development of the trait-based theory (Litchman et al.,
393 2012; Litchman and Klausmeier, 2008) used to define plankton lifeforms for the present work.

394 Not all UK plankton monitoring programmes collected data during the 2004 to 2008 starting conditions period for PI
395 calculation, resulting in the exclusion of some important time-series from the UK 2020 MSFD Assessment and this
396 analysis. While the Environment Agency (EA) dataset spanned the entire time period, the sampling and analysis
397 methodology and frequency changed in 2008, as a result of implementation of the WFD. Special care must therefore
398 be taken when interpreting change from this time-series. Additionally, some plankton surveys, such as the CPR,
399 Marine Scotland Science, the SAMS Lorne Pelagic Observatory, and PML's L4, have multi-decadal databases; when
400 data from only 2004 onward are included the historical data are not used to their full potential. It is therefore clear
401 that further work into maximising the use of UK datasets is urgently required. Such investigations might test: using
402 the entire time-series as the starting condition period for calculating the PI value; varying the starting condition
403 period depending on the length of the dataset; using a more recent period for the starting conditions to include
404 newer time-series; or shortening the starting conditions period to encompass only three years of data and therefore
405 include more UK datasets. Each of these possibilities may have trade-offs. As suggested by Scherer et al. (Scherer et
406 al., 2014), for example, starting condition envelopes which encompass > 5 years will incorporate a greater amount of
407 natural variability and be less sensitive. Conversely, restricting the starting period to a single year (or two) would
408 increase sensitivity but risk detecting inter-annual variability rather than longer-term change. Similarly, using
409 different years for the starting conditions for different datasets will reduce comparability between surveys. Finally, if
410 starting conditions are set too far in the past they will not reflect prevailing conditions. Exploration of these
411 challenges will maximise the use of the UK's plankton datasets, increasing the robustness of future assessments
412 through the inclusion of all UK data.

413 The present analysis illustrates how the PI was used to identify differences in plankton lifeforms over an 11 year
414 (2004 - 2014) time span and applies this method to formal biodiversity assessment under the MSFD. This initial
415 assessment used a time frame to harmonise with the OSPAR MSFD intermediate assessment. When considering the
416 inter-annual variability that exists in the plankton community, the time period examined here is relatively short and
417 will require the inclusion of additional years before it can confidently be established if the changes observed in Figs.
418 4 and 5 are part of a long-term trend (Henson et al., 2009). As mentioned above, for many UK datasets this could be
419 a matter of adjusting the starting conditions period to be further back in time, thereby making better use of multi-
420 decadal datasets. It is therefore imperative to maintain all UK plankton time-series in their current format, as the
421 scientific and policy value of time-series increases with dataset length (Giron-Nava, 2017).

422 Notwithstanding the shortness of the assessment period, the PI value acts successfully as a flag to trigger further
423 investigation the changes that have taken place and the pressures causing change. For example, there have been
424 suggestions that increases in gelatinous zooplankton signify degraded ecosystem states due to stressors including
425 overfishing, pollution, eutrophication and anoxia (Richardson et al., 2009; Tett and Mills, 1991). The lifeform pairs
426 involving gelatinous zooplankton are instructive in this regard with a low PI value (crustaceans and gelatinous
427 zooplankton: $PI = 0.51$, $p < 0.01$) at PML's L4 station reflecting the substantial increase in gelatinous zooplankton that
428 has recently been reported here (McConville, 2018). Several publications point to multidecadal cycles of jellyfish
429 populations and even in heavily fished systems, climate change appears to be implicated in the fluctuations in
430 gelatinous taxa that have been observed (Lynam et al., 2011). This is one example of the PI 'flagging' important
431 trends that merit further analysis on causality. Particular care with interpretation, however, must be taken at the
432 boundary of significance, where $PI = 0.8$, as time-series length and starting condition envelope size may influence
433 statistical significance.

434 Another key strength of our multiple time series approach is that it allows an assessment of large-scale spatial
435 change: are the changes observed localised or widespread? As an example, long-term declines in total copepod
436 abundance have been reported in European shelf waters (Edwards, 2013). The fact that these trends are
437 widespread, and observed both in oceanic and shelf waters and in geographically separate seas (e.g. Celtic and North
438 Seas), could be argued to point more towards widespread, climate-related pressures rather than to trophic cascades
439 induced by overfishing. Impacts from the other major anthropogenic pressure, nutrient enrichment, are more likely
440 to be observed in coastal areas in the first instance. Comparison of PI values between coastal and offshore EHDs will
441 flag which plankton lifeform pairs lack coherence across these broader spatial scales and require further
442 investigation.

443 The work described here demonstrates a method to identify changes in UK plankton communities in support of the
444 2020 UK MSFD Assessment using a diverse range of datasets. To assess GES in fulfilment of the MSFD in line with the
445 Commission Decision on GES (2017/848/EU) (European Commission, 2017), and to use the lifeform approach to
446 inform policy decisions about management measures, two additional, critical steps are needed. Firstly, though the
447 present study identified change in plankton lifeforms between two time periods, identification of a trend in PI away
448 from starting conditions can identify the trajectory of change in lifeform pairs (e.g. Gowen et al., 2015). For
449 assessment purposes, this must be accomplished for each EHD zone and fixed-point time-series, though if time-
450 series are short (i.e. not multi-decadal) the statistical significance of trends and relationships may be difficult to
451 identify.

452 Secondly, change in plankton lifeforms must be interpreted with respect to environmental variation and
453 anthropogenic pressures, to identify factors responsible for plankton community change. This information is
454 required to support government policy decisions about enacting management measures, ensuring effort is applied
455 to appropriate human drivers and pressures. Causal identification is critical when assessing indicator change against
456 the agreed UK target of 'Plankton are not significantly influenced by direct anthropogenic pressure'. This target is
457 process-based, rather than linked to a threshold, which means that as long as change in the plankton is not driven by
458 direct anthropogenic pressures, such as fishing or nutrient loading, the pelagic habitat is deemed to be in GES. This
459 process-based target allows the plankton community to shift and change due to environmental and/or climate
460 change, known as 'prevailing conditions' under the Directive. The management of prevailing conditions is outside the
461 scope of the MSFD, but failing the target will trigger management action if a directly manageable anthropogenic
462 pressure causes change in the plankton community. Pressure identification will therefore help to recognise changes
463 caused by prevailing environmental conditions, a state which may be different from starting conditions but which
464 still represents GES. The pressure-state relationship in pelagic systems, however, is often unclear or non-linear and
465 discriminating between the different pressures is challenging, requiring further research (Dickey-Collas et al., 2017).
466 Despite challenges in understanding the pressure-state relationship for plankton communities, the use of plankton
467 lifeforms in a surveillance role, for example in interpreting change in other ecosystem components, also requires
468 further consideration (e.g. Bedford et al., 2018; Shephard et al., 2015).

469 The lifeform indicator is an OSPAR common indicator (PH1/FW5: Changes in Phytoplankton and Zooplankton
470 Communities) and was used for the regional OSPAR 2017 Intermediate Assessment (OSPAR, 2017); that assessment,
471 however, only considered data from PML, the CPR and one Swedish sampling station. There are a number of multi-
472 decadal plankton time-series across the OSPAR area (O'Brien et al., 2017), and as these become available to support
473 policy the lifeform indicator is flexible enough to incorporate them. This will provide an improved holistic
474 understanding of change in plankton communities, increasing the robustness of future MSFD assessments which is
475 also in line with the Commission Decision on GES (European Commission, 2017) which recognises the importance of
476 practical criteria (technical feasibility, monitoring costs, adequate time-series of data). The flexibility of the lifeform
477 approach means that the indicator can be used with data from other regional seas as long as appropriate lifeform
478 pairs are selected (Brito et al., 2015; Gowen et al., 2015; Siddons et al., 2018), and in the future could be applied at a
479 pan-European scale. Using the same indicator throughout Europe's seas would allow clear, easily comparable

480 assessments of plankton community change, enabling a consistent and coherent view of pelagic habitat status across
481 Europe.

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497

- Appeltans, W., Ah Yong, S.T., Anderson, G., Angel, M.V., Artois, T., Bailly, N., Bamber, R., Barber, A., Bartsch, I., Berta, A., Blazewicz-Paszkowycz, M., Bock, P., Boxshall, G., Boyko, C.B., Brandao, S.N., Bray, R.A., Bruce, N.L., Cairns, S.D., Chan, T.Y., Cheng, L., Collins, A.G., Cribb, T., Curini-Galletti, M., Dahdouh-Guebas, F., Davie, P.J., Dawson, M.N., De Clerck, O., Decock, W., De Grave, S., de Voogd, N.J., Domning, D.P., Emig, C.C., Erseus, C., Eschmeyer, W., Fauchald, K., Fautin, D.G., Feist, S.W., Fransen, C.H., Furuya, H., Garcia-Alvarez, O., Gerken, S., Gibson, D., Gittenberger, A., Gofas, S., Gomez-Daglio, L., Gordon, D.P., Guiry, M.D., Hernandez, F., Hoeksema, B.W., Hopcroft, R.R., Jaume, D., Kirk, P., Koedam, N., Koenemann, S., Kolb, J.B., Kristensen, R.M., Kroh, A., Lambert, G., Lazarus, D.B., Lemaitre, R., Longshaw, M., Lowry, J., Macpherson, E., Madin, L.P., Mah, C., Mapstone, G., McLaughlin, P.A., Mees, J., Meland, K., Messing, C.G., Mills, C.E., Molodtsova, T.N., Mooi, R., Neuhaus, B., Ng, P.K., Nielsen, C., Norenburg, J., Opresko, D.M., Osawa, M., Paulay, G., Perrin, W., Pilger, J.F., Poore, G.C., Pugh, P., Read, G.B., Reimer, J.D., Rius, M., Rocha, R.M., Saiz-Salinas, J.I., Scarabino, V., Schierwater, B., Schmidt-Rhaesa, A., Schnabel, K.E., Schotte, M., Schuchert, P., Schwabe, E., Segers, H., Self-Sullivan, C., Shenkar, N., Siegel, V., Sterrer, W., Stohr, S., Swalla, B., Tasker, M.L., Thuesen, E.V., Timm, T., Todaro, M.A., Turon, X., Tyler, S., Uetz, P., van der Land, J., Vanhoorne, B., van Ofwegen, L.P., van Soest, R.W., Vanaverbeke, J., Walker-Smith, G., Walter, T.C., Warren, A., Williams, G.C., Wilson, S.P., Costello, M.J., 2012. The magnitude of global marine species diversity. *Current biology* : CB 22, 2189-2202.
- Atkinson, A., Harmer, R.A., Widdicombe, C.E., McEvoy, A.J., Smyth, T.J., Cummings, D.G., Somerfield, P.J., Maud, J.L., McConville, K., 2015. Questioning the role of phenology shifts and trophic mismatching in a planktonic food web. *Progress in Oceanography* 137, 498-512.
- Barton, A.D., Lozier, M.S., Williams, R.G., 2015. Physical controls of variability in North Atlantic phytoplankton communities. *Limnology and Oceanography* 60, 181-197.
- Bean, T.P., Greenwood, N., Beckett, R., Biermann, L., Bignell, J.P., Brant, J.L., Copp, G.H., Devlin, M.J., Dye, S., Feist, S.W., Fernand, L., Foden, D., Hyder, K., Jenkins, C.M., van der Kooij, J., Kröger, S., Kupschus, S., Leech, C., Leonard, K.S., Lynam, C.P., Lyons, B.P., Maes, T., Nicolaus, E.E.M., Malcolm, S.J., McIlwaine, P., Merchant, N.D., Paltriguera, L., Pearce, D.J., Pitois, S.G., Stebbing, P.D., Townhill, B., Ware, S., Williams, O., Righton, D., 2017. A Review of the Tools Used for Marine Monitoring in the UK: Combining Historic and Contemporary Methods with Modeling and Socioeconomics to Fulfill Legislative Needs and Scientific Ambitions. *Frontiers in Marine Science* 4.
- Beaugrand, G., 2004. The North Sea regime shift: Evidence, causes, mechanisms and consequences. *Progress in Oceanography* 60, 245-262.
- Beaugrand, G., 2005. Monitoring pelagic ecosystems using plankton indicators. *ICES J. Mar. Sci.* 62, 333-338.
- Beaugrand, G., Brander, K.M., Alistair Lindley, J., Souissi, S., Reid, P.C., 2003. Plankton effect on cod recruitment in the North Sea. *Nature* 426, 661.
- Beaugrand, G., Kirby, R.R., 2018. How Do Marine Pelagic Species Respond to Climate Change? Theories and Observations. *Annual Review of Marine Science* 10, 169-197.
- Beaugrand, G., Reid, P.C., Ibanez, F., Lindley, J.A., Edwards, M., 2002. Reorganization of North Atlantic marine copepod biodiversity and climate. *Science* 296, 1692-1694.
- Bedford, J., Johns, D., Greenstreet, S., McQuatters-Gollop, A., 2018. Plankton as prevailing conditions: a surveillance role for plankton indicators within the Marine Strategy Framework Directive. *Marine Policy* 89, 109-115.
- Bremner, J., Frid, C.L.J., Rogers, S.I., 2004. Biological traits of the North Sea benthos – Does fishing affect benthic ecosystem function? , in: Barnes, P., Thomas, J. (Ed.), *Benthic habitats and the effects of fishing*. American Fisheries Society, Bethesda, MD, pp. 477-489.
- Bremner, J., Rogers, S.I., Frid, C.L.J., 2003. Assessing functional diversity in marine benthic ecosystems: a comparison of approaches. *Marine Ecology Progress Series* 254, 11-25.
- Bresnan, E., Cook, K., Hindson, J., Hughes, S., Lacaze, J.-P., Walsham, P., Webster, L., Turrell, W.R., 2016. The Scottish Coastal Observatory 1997-2013. Parts 1-3. *Scottish Marine and Freshwater Science* 7 No 26.
- Brito, A.C., Moita, T., Gameiro, C., Silva, T., Anselmo, T., Brotas, V., 2015. Changes in the Phytoplankton Composition in a Temperate Estuarine System (1960 to 2010). *Estuaries and Coasts* 38, 1678-1691.
- Capuzzo, E., Lynam, C.P., Barry, J., Stephens, D., Forster, R.M., Greenwood, N., McQuatters-Gollop, A., Silva, T., Sonja M. van Leeuwen, Engelhard, G.H., 2018. A decline in primary production in the North Sea over 25 years, associated with reductions in zooplankton abundance and fish stock recruitment. *Global Change Biology* 24, e352-e364.
- Claussen, U., Connor, D., De Vrees, L., Leppänen, J., Percelay, J., Kapari, M., Mihail, O., 2011. Common Understanding of (Initial) Assessment, Determination of Good Environmental Status (GES) and Establishment of Environmental Targets (Art. 8, 9 & 10 MSFD). Working Group on GES, p. 71.

552 de Vargas, C., Audic, S., Henry, N., Decelle, J., Mahé, F., Logares, R., Lara, E., Berney, C., Le Bescot, N., Probert, I.,
553 Carmichael, M., Poulain, J., Romac, S., Colin, S., Aury, J.-M., Bittner, L., Chaffron, S., Dunthorn, M., Engelen, S.,
554 Flegontova, O., Guidi, L., Horák, A., Jaillon, O., Lima-Mendez, G., Lukeš, J., Malviya, S., Morard, R., Mulot, M., Scalco,
555 E., Siano, R., Vincent, F., Zingone, A., Dimier, C., Picheral, M., Searson, S., Kandels-Lewis, S., Acinas, S.G., Bork, P.,
556 Bowler, C., Gorsky, G., Grimsley, N., Hingamp, P., Iudicone, D., Not, F., Ogata, H., Pesant, S., Raes, J., Sieracki, M.E.,
557 Speich, S., Stemmann, L., Sunagawa, S., Weissenbach, J., Wincker, P., Karsenti, E., Boss, E., Follows, M., Karp-Boss, L.,
558 Krzic, U., Reynaud, E.G., Sardet, C., Sullivan, M.B., Velayoudon, D., 2015. Eukaryotic plankton diversity in the sunlit
559 ocean. *Science* 348.

560 Devlin, M., Barry, J., Painting, S., Best, M., 2009. Extending the phytoplankton tool kit for the UK Water Framework
561 Directive: indicators of phytoplankton community structure. *Hydrobiologia* 633, 151-168.

562 Devlin, M., Best, M., Coates, D., Bresnan, E., O'Boyle, S., Park, R., Silke, J., Cusack, C., Skeats, J., 2007. Establishing
563 boundary classes for the classification of UK marine waters using phytoplankton communities. *Marine Pollution*
564 *Bulletin* 55, 91-103.

565 Dickey-Collas, M., McQuatters-Gollop, A., Bresnan, E., Kraberg, A.C., Manderson, J.P., Nash, R.D.M., Otto, S.A., Sell,
566 A.F., Tweddle, J.F., Trenkel, V.M., 2017. Pelagic habitat: exploring the concept of good environmental status. *ICES J.*
567 *Mar. Sci.* 74, 2333-2341.

568 Edwards, M., Bresnan, E., Cook, K., Heath, M., Helaouet, P., Lynam, C., Raine, R. and Widdicombe, C., 2013. Impacts
569 of climate change on plankton, *MCCIP Science Review*, pp. 98-112.

570 Edwards, M., Richardson, A.J., 2004. Impact of climate change on marine pelagic phenology and trophic mismatch.
571 *Nature* 430, 881-884.

572 Eloire, D., Somerfield, P.J., Conway, D.V.P., Halsband-Lenk, C., Harris, R., Bonnet, D., 2010. Temporal variability and
573 community composition of zooplankton at station L4 in the Western Channel: 20 years of sampling. *Journal of*
574 *Plankton Research* 32, 657-679.

575 Estrada, M., Henriksen, P., Gasol, J.M., Casamayor, E.O., Pedrós-Alió, C., 2004. Diversity of Planktonic
576 Photoautotrophic Microorganisms Along a Salinity Gradient as Depicted by Microscopy, Flow Cytometry, Pigment
577 Analysis and DNA-based Methods. *FEMS Microbiology Ecology* 49, 281-293.

578 European Commission, 2010. Commission Decision of 1 September 2010 on criteria and methodological standards
579 on good environmental status of marine waters 2010/477/EU.

580 European Commission, 2017. Commission Decision (EU) 2017/848 of 17 May 2017, laying down criteria and
581 methodological standards on good environmental status of marine waters and specifications and standardised
582 methods for monitoring and assessment, and repealing Decision 2010/447/EU. *Official Journal of the European*
583 *Union L 125(18.5.2017)*, 43-74.

584 Falkowski, P.G., Katz, M.E., Knoll, A.H., Quigg, A., Raven, J.A., Schofield, O., Taylor, F.J.R., 2004. The evolution of
585 modern eukaryotic phytoplankton. *Science* 305, 354– 360.

586 Fox, C.J., Pitois, S.G., 2006. Long-term changes in zooplankton biomass concentration and mean size over the
587 Northwest European shelf inferred from Continuous Plankton Recorder data. *ICES J. Mar. Sci.* 63, 785–798.

588 Gallego, I., Davidson, T.A., Jeppesen, E., Perez-Martinez, C., Sanchez-Castillo, P., Juan, M., Fuentes-Rodriguez, F.,
589 Leon, D., Penalver, P., Toja, J., Casas, J.J., 2012. Taxonomic or ecological approaches? Searching for phytoplankton
590 surrogates in the determination of richness and assemblage composition in ponds. *Ecological Indicators* 18, 575-585.

591 Garmendia, M., Borja, Á., Franco, J., Revilla, M., 2012. Phytoplankton composition indicators for the assessment of
592 eutrophication in marine waters: Present state and challenges within the European directives. *Marine Pollution*
593 *Bulletin* 66, 7-16.

594 Giron-Nava, A., James, C.C., Johnson, A.F., Dannecker, D., Johns, D.G., Kolody, B., Lee, A., Nagarkar, M., Pao, G.M.,
595 Ye, H. and Sugihara, G., 2017. A quantitative argument for long-term ecological monitoring. *Marine Ecology Progress*
596 *Series* 572, 269-274.

597 Gowen, R.J., Collos, Y., Tett, P., Scherer, C., Bec, B., Abadie, E., Allen, M., O'Brien, T., 2015. Response of diatom and
598 dinoflagellate lifeforms to reduced phosphorus loading: A case study in the Thau lagoon, France. *Estuarine, Coastal*
599 *and Shelf Science* 162, 45-52.

600 Gowen, R.J., McQuatters-Gollop, A., Tett, P., Best, M., Bresnan, E., Castellani, C., Cook, K., Forster, R., Scherer, C.,
601 Mckinney, A., 2011. The Development of UK Pelagic (Plankton) Indicators and Targets for the MSFD. *Advice to Defra*,
602 *Belfast, UK*, p. 41 pp.

603 Gowen, R.J., Raine, R., Dickey-Collas, M., White, M., 1998. Plankton distributions in relation to physical
604 oceanographic features on the southern Malin Shelf, August 1996. *ICES J. Mar. Sci.* 55, 1095-1111.

605 Gowen, R.J., Stewart, B.M., 2005. The Irish Sea: Nutrient status and phytoplankton. *Journal of Sea Research* 54, 36-
606 50.

607 Gowen, R.J., Stewart, B.M., Mills, D.K., Elliott, P., 1995. Regional differences in stratification and its effect on
608 phytoplankton production and biomass in the northwestern Irish Sea. *Journal of Plankton Research* 17, 753-769.

609 Gowen, R.J., Tett, P., Kennington, K., Mills, D.K., Shammon, T.M., Stewart, B.M., Greenwood, N., Flanagan, C., Devlin,
610 M., Wither, A., 2008. The Irish Sea: Is it eutrophic? *Estuarine, Coastal and Shelf Science* 76, 239-254.

611 Gregory, B., Christophe, L., Martin, E., 2009. Rapid biogeographical plankton shifts in the North Atlantic Ocean.
612 *Global Change Biology* 15, 1790-1803.

613 Hays, G.C., Richardson, A.J., Robinson, C., 2005. Climate change and marine plankton. *Trends in Ecology & Evolution*
614 20, 337-344.

615 Henson, S.A., Raitsos, D., Dunne, J.P., McQuatters-Gollop, A., 2009. Decadal variability in biogeochemical models:
616 Comparison with a 50-year ocean colour dataset. *Geophysical Research Letters* 36, L21601.

617 ICES, 2015. Book 1, ICES special request advice, published 20 March 2015. EU request on revisions to Marine
618 Strategy Framework Directive manuals for Descriptors 3, 4, and 6, Technical report. ICES, Copenhagen.

619 Inall, M.E., Gillibrand, P.A., 2010. The physics of mid-latitude fjords: a review. Geological Society, London, Special
620 Publications 344, 17-33.

621 John, E.H., Batten, S.D., Harris, R.P., Hays, G.C., 2001. Comparison between zooplankton data collected by the
622 Continuous Plankton Recorder survey in the English Channel and by WP-2 nets at station L4, Plymouth (UK). *Journal*
623 *of Sea Research* 46, 223-232.

624 Jones, K.J., Gowen, R.J., Tett, P., 1984. Water-column structure and summer phytoplankton distribution in the Sound
625 of Jura. *Journal of Experimental Marine Biology and Ecology* 78, 269-289.

626 Katsanevakis, S., Stelzenmüller, V., South, A., Sørensen, T.K., Jones, P.J.S., Kerr, S., Badalamenti, F., Anagnostou, C.,
627 Breen, P., Chust, G., D'Anna, G., Duijn, M., Filatova, T., Fiorentino, F., Hulsman, H., Johnson, K., Karageorgis, A.P.,
628 Kröncke, I., Mirto, S., Pipitone, C., Portelli, S., Qiu, W., Reiss, H., Sakellariou, D., Salomidi, M., van Hoof, L.,
629 Vassilopoulou, V., Vega Fernández, T., Vöge, S., Weber, A., Zenetos, A., Hofstede, R.t., 2011. Ecosystem-based
630 marine spatial management: Review of concepts, policies, tools, and critical issues. *Ocean & Coastal Management*
631 54, 807-820.

632 Kirby, R.R., Beaugrand, G., Lindley, J.A., 2008. Climate-induced effects on the meroplankton and the benthic-pelagic
633 ecology of the North Sea. *Limnology and Oceanography* 53, 1805-1815.

634 Kirby, R.R., Beaugrand, G., Lindley, J.A., 2009. Synergistic Effects of Climate and Fishing in a Marine Ecosystem.
635 *Ecosystems* 12, 548-561.

636 Lindeque, P.K., Parry, H.E., Harmer, R.A., Somerfield, P.J., Atkinson, A., 2013. Next Generation Sequencing Reveals
637 the Hidden Diversity of Zooplankton Assemblages. *PLOS ONE* 8, e81327.

638 Lindley, J.A., Gamble, J.C., Hunt, H.G., 1995. A change in the zooplankton of the central North Sea (55 to 58 N): a
639 possible consequence of changes in the benthos. *Marine Ecology Progress Series* 119, 299-303.

640 Litchman, E., Edwards, K.F., Klausmeier, C.A., Thomas, M.K., 2012. Phytoplankton niches, traits and eco-evolutionary
641 responses to global environmental change. *Marine Ecology Progress Series* 470, 235-248.

642 Litchman, E., Klausmeier, C.A., 2008. Trait-Based Community Ecology of Phytoplankton. *Annual Review of Ecology,*
643 *Evolution, and Systematics* 39, 615-639.

644 Litchman, E., Klausmeier, C.A., Schofield, O.M., Falkowski, P.G., 2007. The role of functional traits and trade-offs in
645 structuring phytoplankton communities: scaling from cellular to ecosystem level. *Ecology Letters* 10, 1170-1181.

646 Lynam, C.P., Lilley, M.K.S., Bastian, T., Doyle, T.K., Beggs, S.E., Hays, G.C., 2011. Have jellyfish in the Irish Sea
647 benefited from climate change and overfishing? *Global Change Biology* 17, 767-782.

648 Margalef, R., 1978. Life forms of phytoplankton as survival alternatives in an unstable environment. *Oceanologica*
649 *Acta* 1, 493-509.

650 McConville, K.M., 2018. Trophic and ecological implications for the gelatinous body form in zooplankton. PhD Thesis,
651 University of Plymouth, p. 223 pp.

652 McQuatters-Gollop, A., Edwards, M., Helaouët, P., Johns, D.G., Owens, N.J.P., Raitsos, D.E., Schroeder, D., Skinner, J.,
653 Stern, R.F., 2015. The Continuous Plankton Recorder survey: how can long-term phytoplankton datasets deliver
654 Good Environmental Status? *Estuarine, Coastal and Shelf Science* 162, 88-97.

655 McQuatters-Gollop, A., Gilbert, A.J., Mee, L.D., Vermaat, J.E., Artioli, Y., Humborg, C., Wulff, F., 2009. How well do
656 ecosystem indicators communicate the effects of anthropogenic eutrophication? *Estuarine, Coastal and Shelf*
657 *Science* 82, 583-596.

658 McQuatters-Gollop, A., Johns, D.G., Bresnan, E., Skinner, J., Rombouts, I., Stern, R.F., Aubert, A., Johansen, M.,
659 Knights, A., 2017. From microscope to management: the critical value of plankton taxonomy to marine policy and
660 biodiversity conservation. *Marine Policy* 83, 1-10.

661 McQuatters-Gollop, A., Reid, P.C., Edwards, M., Burkill, P., Castellani, C., Batten, S., Gieskes, W., Beare, D., Bidigare,
662 R., Head, E., Johnson, R., Kahru, M., Koslow, J., Pena, M., 2011. Is there a decline in marine phytoplankton? *Nature*
663 472, E6-E7.

664 Mouillot, D., Graham, N.A.J., Villéger, S., Mason, N.W.H., Bellwood, D.R., 2013. A functional approach reveals
665 community responses to disturbances. *Trends in Ecology & Evolution* 28, 167-177.

666 Mouillot, D., Spatharis, S., Reizopoulou, S., Laugier, T., Sabetta, L., Basset, A., Chi, T.D., 2006. Alternatives to
667 taxonomic-based approaches to assess changes in transitional water communities. *Aquatic Conservation-Marine and*
668 *Freshwater Ecosystems* 16, 469-482.

669 O'Brien, T.D., Lorenzoni, L., Isensee, K., Valdes, L., 2017. What are Marine Ecological Time Series telling us about the
670 ocean? A status report, IOC Technical Series. IOC-UNESCO, p. 297.

671 OSPAR, 2017. PH1/FW5: Changes in phytoplankton and zooplankton communities., in: OSPAR (Ed.), OSPAR
672 Intermediate Assessment 2017. OSPAR, London, UK, p. 2.

673 Ostle, C., Artigas, F., Atkinson, A., Aubert, A., Budria, A., Graham, G., Helaouët, P., Johns, D., Padegimas, B.,
674 Rombouts, I., Widdicombe, C., McQuatters-Gollop, A., 2017. Spatial Representivity of Plankton Indicators EcApRHA
675 Deliverable WP1.3. OSPAR, London, UK, p. 28.

676 Pingree, R.D., Holligan, P.M., Mardell, G.T., 1978. The effects of vertical stability on phytoplankton distributions in
677 the summer on the northwest European Shelf. *Deep Sea Research* 25, 1011-1028.

678 Purcell, J.E., Arai, M.N., 2001. Interactions of pelagic cnidarians and ctenophores with fish: a review. *Hydrobiologia*
679 451, 27-44.

680 Richardson, A.J., 2008. In hot water: zooplankton and climate change. *ICES J. Mar. Sci.* 65, 279-295.

681 Richardson, A.J., Bakun, A., Hays, G.C., Gibbons, M.J., 2009. The jellyfish joyride: causes, consequences and
682 management responses to a more gelatinous future. *Trends in Ecology & Evolution* 24, 312-322.

683 Richardson, A.J., Walne, A.W., John, A.W.G., Jonas, T.D., Lindley, J.A., Sims, D.W., Stevens, D., Witt, M., 2006. Using
684 Continuous Plankton Recorder data. *Progress in Oceanography* 68, 27-74.

685 Scherer, C., 2012. Developing and testing an index of change in microplankton community structure in temperate
686 shelf seas. A thesis in partial fulfilment of the requirements of Edinburgh Napier University, for the award of Doctor
687 of Philosophy. Napier University, p. 325.

688 Scherer, C., Gowen, R.J., Tett, P., 2016. Assessing the State of the Pelagic Habitat: A Case Study of Plankton and Its
689 Environment in the Western Irish Sea. *Frontiers in Marine Science* 3.

690 Scherer, C., Gowen, R.J., Tett, P., McQuatters-Gollop, A., Forster, R., Bresnan, E., Cook, K., Atkinson, A., Best, M.,
691 Baptie, M., Keeble, S., McCullough, G., McKinney, A., 2014. Development of a UK Integrated Plankton Monitoring
692 Programme A final report of the Lifeform and State Space project, prepared for Defra. Agri-food and Biosciences
693 Institute, Belfast, p. 450.

694 Secretariat of the Convention on Biological Diversity, 2004. The Ecosystem Approach (CBD Guidelines). Secretariat of
695 the Convention on Biological Diversity, Montreal, p. 50 p.

696 Shephard, S., Greenstreet, S.P.R., Piet, G.J., Rindorf, A., Dickey-Collas, M., 2015. Surveillance indicators and their use
697 in implementation of the Marine Strategy Framework Directive. *ICES J. Mar. Sci.*

698 Siddons, B., Glegg, G., McQuatters-Gollop, A., 2018. Inter-regional coherence: can Northeast Atlantic pelagic
699 indicators be applied to the Arctic? *Marine Policy* 96, 53-64.

700 Stoetaert, K., Heip, C., 1990. Sample-size dependence of diversity indices and the determination of sufficient sample
701 size in a high-diversity deep-sea environment. *Marine Ecology Progress Series* 59, 305-307.

702 Stuart-Smith, R.D., Bates, A.E., Lefcheck, J.S., Duffy, J.E., Baker, S.C., Thomson, R.J., Stuart-Smith, J.F., Hill, N.A.,
703 Kininmonth, S.J., Airoidi, L., Becerro, M.A., Campbell, S.J., Dawson, T.P., Navarrete, S.A., Soler, G.A., Strain, E.M.A.,
704 Willis, T.J., Edgar, G.J., 2013. Integrating abundance and functional traits reveals new global hotspots of fish diversity.
705 *Nature* 501, 539.

706 Tett, P., 1973. The use of log-normal statistics to describe phytoplankton populations from the Firth of Lorne area.
707 *Journal of experimental marine Biology and Ecology* 11, 121-136.

708 Tett, P., Carreira, C., Mills, D.K., van Leeuwen, S., Foden, J., Bresnan, E., Gowen, R.J., 2008. Use of a phytoplankton
709 community index to assess the health of coastal waters. *ICES J. Mar. Sci.* 65, 1475-1482.

710 Tett, P., Gowen, R., Painting, S., Elliott, M., Foster, R., Mills, D., Bresnan, E., Capuzzo, E., Fernandes, T., Foden, J.,
711 Geider, R., Gilpin, L., Huxham, M., McQuatters-Gollop, A., Malcolm, S., Saux-Picart, S., Platt, T., Racault, M.-F.,
712 Sathyendranath, S., Molen, J.v.d., Wilkinson, M., 2013. A framework for understanding marine ecosystem health.
713 *Marine Ecology Progress Series* 494, 1-27.

714 Tett, P., Mills, D., 1991. The plankton of the North Sea - pelagic ecosystems under stress ? *Ocean and Shoreline*
715 *Management* 16, 233-257.

716 Tett, P., Wallis, A., 1978. The general annual cycle of chlorophyll standing crop in Loch Creran. *Journal of Ecology* 66,
717 227-239.

718 Thiebaut, M.L., Dickie, L.M., 1993. Structure of the Body-Size Spectrum of the Biomass in Aquatic Ecosystems: A
719 Consequence of Allometry in Predator–Prey Interactions. *Canadian Journal of Fisheries and Aquatic Sciences* 50,
720 1308-1317.

721 Ubertini, M., Lefebvre, S., Gangnery, A., Grangeré, K., Le Gendre, R., Orvain, F., 2012. Spatial variability of benthic-
722 pelagic coupling in an estuary ecosystem: consequences for microphytobenthos resuspension phenomenon. *PLoS*
723 *one* 7, e44155-e44155.

724 UKTAG, 2014. UKTAG Coastal Water Assessment Method, Phytoplankton, Coastal Water Phytoplankton Tool. Water
725 Framework Directive - UK Advisory Group (WFD-UKTAG), Stirling, Scotland, p. 26.

726 van Leeuwen, S., Tett, P., Mills, D., Molen, J.v.d., 2015. Stratified and nonstratified areas in the North Sea: Long-term
727 variability and biological and policy implications. *Journal of Geophysical Research Oceans* 120, 4670-4686.

728 van Leeuwen, S.M., Le Quesne, W.F., Parker, E.R., 2016. Potential future fisheries yields in shelf waters: a model
729 study of the effects of climate change and ocean acidification. *Biogeosciences* 13, 441-454.

730 Villéger, S., Mason, N.W.H., Mouillot, D., 2008. New multidimensional functional diversity indices for a multifaceted
731 framework in functional ecology. *Ecology* 89, 2290-2301.

732 von Bertalanffy, L., 1972. The history and status of General Systems Theory. *The Academy of Management Journal*
733 15, 407-426.

734 Weston, K., Greenwood, N., Fernand, L., Pearce, D.J., Sivy, D.B., 2008. Environmental controls on phytoplankton
735 community composition in the Thames plume, U.K. *Journal of Sea Research* 60, 246-254.

736 Whyte, C., Davidson, K., Gilpin, L., Mitchell, E., Moschonas, G., McNeill, S., Tett, P., 2017. Tracking changes to a
737 microplankton community in a North Atlantic sea loch using the microplankton index PI(mp). *ICES J. Mar. Sci.* 74,
738 311-325.

739 Widdicombe, C.E., Eloire, D., Harbour, D., Harris, R.P., Somerfield, P.J., 2010. Long-term phytoplankton community
740 dynamics in the Western English Channel. *Journal of Plankton Research* 32, 643-655.

741 Williams, R., Conway, D.V.P., Hunt, H.G., 1994. The role of copepods in the planktonic ecosystems of mixed and
742 stratified waters of the European shelf seas. *Hydrobiologia* 292, 521-530.

743 Wood, B.J.B., Tett, P.B., Edwards, A., 1973. An Introduction to the Phytoplankton, Primary Production and Relevant
744 Hydrography of Loch Etive. *Journal of Ecology* 61, 569-585.

745