



Article

# Accuracy Assessment of Primary Production Models with and without Photoinhibition Using Ocean-Colour Climate Change Initiative Data in the North East Atlantic Ocean

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Abstract: The accuracy of three satellite models of primary production (PP) of varying complexity was assessed against 95 in situ 14C uptake measurements from the North East Atlantic Ocean (NEA). The models were run using the European Space Agency (ESA), Ocean Colour Climate Change Initiative (OC-CCI) version 3.0 data. The objectives of the study were to determine which is the most accurate PP model for the region in different provinces and seasons, what is the accuracy of the models using both high (daily) and low (weekly) temporal resolution OC-CCI data, and whether the performance of the models is improved by implementing a photoinhibition function? The Platt-Sathyendranath primary production model (PPPSM) was the most accurate over all NEA provinces and, specifically, in the Atlantic Arctic province (ARCT) and North Atlantic Drift (NADR) provinces. The implementation of a photoinhibition function in the PP<sub>PSM</sub> reduced its accuracy, especially at lower range PP. The Vertical Generalized Production Model-VGPM (PPvgpm) tended to over-estimate PP, especially in summer and in the NADR. The accuracy of PPv<sub>GPM</sub> improved with the implementation of a photoinhibition function in summer. The absorption model of primary production (PPAph), with and without photoinhibition, was the least accurate model for the NEA. Mapped images of each model showed that the PPvgpm was 150% higher in the NADR compared to PPPSM. In the North Atlantic Subtropical Gyre (NAST) province, PP<sub>Aph</sub> was 355% higher than PP<sub>PSM</sub>, whereas PP<sub>VGPM</sub> was 215% higher. A sensitivity analysis indicated that chlorophyll-a (Chl a), or the absorption of phytoplankton, at 443 nm (aph (443)) caused the largest error in the estimation of PP, followed by the photosynthetic rate terms and then the irradiance functions used for each model.

**Keywords:** phytoplankton; photosynthesis; primary production; North Atlantic Ocean; ocean colour; remote sensing

# 1. Introduction

The rate of synthesis of organic matter by marine phytoplankton through the process of photosynthesis determines the energy flow through the trophic chain in the global ocean. This process, known as PP, not only fuels biological growth and fish productivity, but also regulates carbon uptake and release by the ocean [1]. Since the 1950's, PP has been determined using radio

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labelled carbon [2] through in situ or simulated in situ incubations. Though these methods have provided great insight into the variability in PP, the number of measurements available is limited both spatially and temporally. The introduction of rapid fluorescent based and O<sub>2</sub>/Ar ratio methods using membrane inlet mass spectrometry has increased the number of measurements that can be made over time and space, but still cannot provide synoptic coverage at global, basin, or regional scales. The use of ocean colour remote sensing data has revolutionised our ability to monitor PP at higher temporal and spatial scales than can be provided by in situ techniques [3–8]. Over the past two decades, there has therefore been a concerted effort to estimate PP from the satellite ocean colour using models based on the following input parameters: (1) Phytoplankton biomass expressed as Chl *a*, carbon concentration, or phytoplankton absorption; (2) photo-physiology or photosynthetic rates; and (3) the light field. More than twenty models have been developed that are run solely using remotely sensed data. Some of these models have been validated to ascertain their relative or absolute accuracy [9–14].

Historic studies have shown that satellite derived PP accounts for 50 to 80% of the variance in measured PP depending on the input parameters used in the model [6,7,9,15,16]. Many of these historic studies are limited by a lack of in situ data to characterize the spatial, temporal, and vertical variability in photo-physiology that cannot be measured by remotely sensed parameters alone. Some parameters used in the model, including photosynthetic rates, are input as constants or as functions of other physical parameters, and their vertical distribution is assumed to be constant with depth, which can lead to significant errors in model performance [9–11,17]. In addition, some models fail in regions with atypical optical and photo-physiological properties [18–20] and therefore require a specific regional parameterization.

Early validation studies illustrated that the main cause of the difference between modelled and in situ PP was from parameters that characterize the absorption of light by phytoplankton through Chl a, phytoplankton absorption coefficients, or carbon biomass [21-23]. This resulted in an over-estimate of PP in eutrophic waters and an under-estimate in mesotrophic and oligotrophic waters [15]. A series of National Aeronautics and Space Administration (NASA) PP model inter-comparisons were conducted in the 2000's, firstly by [9], who compared twelve different PP models in nine regions of the world ocean using 89 in situ PP measurements, and found that the difference between satellite and in situ estimates of PP was highly correlated with differences between in satellite and in situ Chl a (See also [24]). Errors in the PP model can be further compounded when other input parameters are derived from Chl a. The second NASA PP inter-comparison [10] illustrated that a four-fold variation of Chl a caused a three-fold variation in PP, whereas variations in photosynthetic active radiation (PAR) and sea surface temperature (SST) had smaller effects on PP. This study also showed however, that deriving photosynthetic rates from temperature alone can cause a significant error in PP. More recently, NASA compared twenty-one PP models in ten regions of the World Ocean using 1156 in situ PP measurements, and showed that the main cause of error in PP models was in coastal, optically complex waters where satellite Chl a can be inaccurate [13]. Further to this, Milutinović and Bertino [25] conducted a comprehensive analysis of the errors associated with the parameters used in one PP model (the Vertical Generalized Production Model - VGPM [6]) and showed that up to 90% of the error is associated with the biomass specific optimum photosynthetic rate  $(P^{B_{opt}})$ . The use of regional in situ values of photosynthetic parameters in satellite derived models improves the performance of PP models [26–

In this paper, we assess the accuracy of one Wavelength-Integrated (WIM) and two Depth-Integrated (DIM) models [29] that are commonly used in the literature, in the North Atlantic. The WIM tested is the Platt and Sathyendranath model (PP<sub>PSM</sub>) [22], which uses an exponential Photosynthesis-Irradiance (*P-I*) function based on the absorption of light by the photosystems [30,31]. The most commonly used DIM is VGPM (PP<sub>VGPM</sub>), which is simple and quick to apply to satellite data. By contrast, we also assess the DIM Absorption Based Model (PP<sub>Aph</sub>) [32,33], which uses the optical absorption signature of phytoplankton as an empirical function of phytoplankton biomass to derive PP. To date, few models include a photoinhibition term that accounts for the

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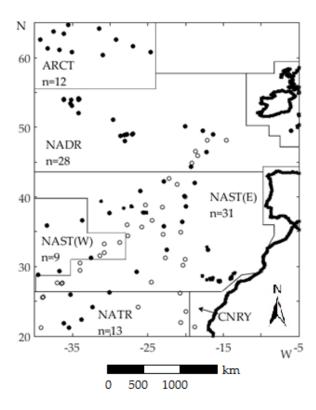
photosynthetic rate with increasing irradiance that can occur under high light conditions or when the phytoplankton community moves suddenly from low to high light, which can cause a decrease in the photosynthetic efficiency [34].

Model performance was assessed using 95 in situ measurements of daily water column primary production ( $PP_{eu}$ ) from the NEA. The models were run using the ESA, OC-CCI version 3.0 data. The paper addresses the following questions: (1) which is the most accurate model for this region? (2) Does the model performance improve when implementing a photoinhibition function? (3) Does the accuracy of each model change depending on the province and season? (4) What is the accuracy of the models using both high (daily) and low (weekly) temporal resolution data? (5) Which input parameter causes the greatest error in each model?

### 2. Materials and Methods

# 2.1. Study Region

Stations of *PP<sub>eu</sub>* were sampled in the North East Atlantic (20–65°N, 5–40°W) from 1998 to 2013, covering the four biological provinces according to [35]: the ARCT, NADR, NAST, and NATR (North Atlantic Tropical Gyre province) (Figure 1, Table 1). A total of 95 stations were analysed during summer (May–August) and in autumn (September–October). Based on OC-CCI Chl *a* during the sampling periods, 54% of the stations were in oligotrophic waters (0–0.1 mg m<sup>-3</sup>) and 42% were mesotrophic (0.1–1 mg m<sup>-3</sup>) [36].



**Figure 1.** Stations sampled for the determination of in situ integrated daily water column primary production ( $PP_{eu}$ ) in the North East Atlantic. Filled circles are stations sampled in summer; open circles are stations sampled in autumn.

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**Table 1.** Provinces and time periods for in situ integrated daily water column primary production  $(PP_{eu})$  (mg C m<sup>-2</sup> day<sup>-1</sup>) used for the accuracy assessment of the primary production satellite models.

Region	North East Atlantic (NEA): 20–65°N, 5–40°W						
Period	1998–2013						
Number of stations (n)	95						
Seasons 1							
summer, $n = 56$	June-1998, July–August 2002, June-2003, May-2004, June-2005, July–August 2007, August-2009						
autumn, $n = 38$	September-2003, September-2004, October-2008, September-2009, October-2011, October-2012, October-2013						
	ARCT $^2$ —Atlantic Arctic Province ( $n = 12$ )						
Provinces	$NADR-North\ Atlantic\ Drift\ (n=28)$						
Provinces	NAST $^3$ —North Atlantic Subtropical Gyre (East & West) ( $n = 40$ )						
	NATR $^4$ —North Atlantic Tropical Gyre ( $n = 14$ )						

<sup>&</sup>lt;sup>1</sup> When all stations were considered together, additional data from April 2002 were also included. <sup>2</sup> Biogeographical provinces were defined following [35]. <sup>3</sup> Analysis of stations in NAST(W) and NAST(E) were merged into NAST. <sup>4</sup> One station from CNRY (Canary Current Coastal province) was added to stations in NATR.

# 2.2. Simulated In Situ <sup>14</sup>C Primary Production Measurements (PP<sub>eu</sub>)

PPeu was determined on Atlantic Meridional Transects (AMT) 6, 12, 13, 14, 15, 16, 18, 19, 21, 22, and 23 [37], Ecosystem of the Mid-Atlantic Ridge (ECOMAR) cruises in the North Atlantic JC011 and JC037, and cruise D264 in the Celtic Sea [14]. Water samples were taken from 6 to 9 depths and transferred from Niskin bottles to black carboys to prevent shock to the photosynthetic lamellae of the phytoplankton cells. Water from each sample was sub sampled into three 75 mL clear polycarbonate bottles and three black polycarbonate bottles. The bottles were pre-cleaned following Joint Global Ocean Flux Study (JGOFS) protocols to reduce trace metal contamination [38]. Each sample was inoculated with between 185 and 740 kBq (5-20 μCi) NaH<sup>14</sup>CO<sub>3</sub> according to the biomass of phytoplankton and transferred to an on deck (simulated in situ) incubation system using neutral density and blue filters to simulate subsurface irradiance over depth to 97%, 55%, 33%, 20%, 14%, 1%, or 0.1% of the surface value. Incubations were conducted from dawn to dusk for 10 to 16 h. The upper four light levels were maintained at the in situ temperature by pumping water from the sea surface into the incubators, and the two lower light depths were chilled with fresh water from a water bath maintained at within ±3 °C of the in situ temperature. For all cruises, after incubating the water samples, suspended material was filtered through 0.2 µm polycarbonate filters at a vacuum pressure of <20 cm Hg. The filters were then rinsed with filtered seawater, exposed to concentrated HCl fumes, and immersed in a 5 mL scintillation cocktail. <sup>14</sup>C disintegration time per minute (DPM) was measured on board on either a Beckman LS6000 SC or a TriCarb 3900 TR liquid scintillation counter (LSC) using the external standard and the channel ratio methods to correct for quenching. The quantity of <sup>14</sup>C added to the experimental bottles was determined by adding aliquots of the stock 14C solution to Carbosorb, a CO2 absorbing scintillation cocktail, and counted immediately on the LSC.

### 2.3. Satellite Data

To estimate PP, satellite ocean colour, SST, and PAR data were used. Each model was firstly run using daily OC-CCI v.3.0 data. This was then compared using 8-day composite OC-CCI data, which is freely available to end-users through the internet. From the OC-CCI database Chl a concentration (mg m<sup>-3</sup>), the downwelling diffuse attenuation coefficient (kd) at 490 nm (m<sup>-1</sup>) and the absorption coefficient of phytoplankton ( $a_ph$ ) at 443 nm [m<sup>-1</sup>] were obtained from the merged information of four ocean colour sensors (SeaWiFS, MODIS-Aqua, MERIS, and VIIRS) at a spatial resolution of 4 × 4 km. The data was firstly filtered to exclude possible outliers related to the weather conditions and edge of cloud effects. This was performed by log transforming the data and then screening for outliers, which differed by more than four standard deviations (over a 21 × 21 pixels area) from the average at each match-up station. Additional filtering was conducted according to the type of water masses; only Case 1 waters, according to [39], were analysed in the study. All pixels classified as eutrophic

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based on a euphotic depth of <9.8 m [40], and for Case 2 waters based on  $k_d$  490 nm > 0.47 m<sup>-1</sup>, were removed.

Daily and 8-day PAR (E  $m^{-2}$  day<sup>-1</sup>) data with a spatial resolution of  $4 \times 4$  km were downloaded from the NASA's OceanColor Web. Averaged values from two ocean colour sensors, SeaWiFS and MODIS-Aqua, were used. Preliminary analysis showed that the difference between PAR values from two sensors was less than 3%. Therefore, using one sensor only, instead of the average of the two, when another sensor is not available does not cause significant errors.

Daily SST data (°C) with a spatial distribution  $1 \times 1$  km were downloaded from the Multiscale Ultra-high Resolution Sea Surface Temperature database, which is a merged data set from AVHRR, MODIS-Aqua, and AMSR, in which the influence of clouds is reduced due to the use of both infrared and microwave satellite data. As daily SST data have a  $1 \times 1$  km spatial distribution, we re-gridded them to  $4 \times 4$  km, in alignment with the ocean colour data.

### 2.4. Satellite Models of Primary Production

The models that were assessed in this study are commonly used to estimate PP, but are architecturally different. PPvGPM is forced by ocean colour Chl a and the variability in the photosynthetic rate is parameterized from satellite SST. PP<sub>Aph</sub> is based on the absorption properties of phytoplankton ( $a_{ph}$ ), which can be derived directly from the inversion of remote sensing reflectance rather than from Chl a [41]. The principal advantage of using phytoplankton absorption at 443 nm- $a_{ph}$  (443) is that it minimizes the effects of coloured dissolved organic matter and non-algal absorption, which can adversely affect PP models by modifying the light field [41,42]. PP<sub>PSM</sub> is more complex than the other two models [43], since it simulates changes in photosynthesis as a function of irradiance using a two-parameter exponential P-I function based on the theory of initial light absorption by the photosynthetic system [30,31]. By contrast, PPvGPM and PP<sub>Aph</sub> use the Michaelis-Menten equation [44].

PPv<sub>GPM</sub> retrieves  $PP_{eu}$ . It was parameterized using  $Chl_{90}$  (mg m<sup>-3</sup>), which is the average Chl a concentration in a layer of the first optical depth seen by ocean colour sensors,  $z_{eu}$ , which is the euphotic depth, (m),  $I_0$  is the daily surface PAR, (E m<sup>-2</sup> day<sup>-1</sup>), DL is the day length, (h), and an empirical coefficient, 0.66125 (see Equation (1)).  $P^B_{opt}$  (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup>), in the VGPM, was retrieved as a function of SST according to the algorithm presented in [6]. The P-I function (given in square brackets) is based on the Michaelis-Menten equation, with the half saturated constant equal to  $4.1 \text{ E m}^{-2} \text{ day}^{-1}$ :

$$PP_{eu} = Chl_{90}z_{eu}0.66125 \left[ P_{opt}^{B} \frac{I_0}{4.1 + I_0} \right] DL, \tag{1}$$

PP<sub>PSM</sub> retrieves daily PP for a specific depth, z, PP(z), based on the P-I function as an exponential curve:

$$PP(z) = Chl(z) \left[ P_m^B \left( 1 - exp\left( -\frac{\alpha^B I(z)}{P_m^B} \right) \right) \right] DL, \tag{2}$$

where, Chl(z) is Chl a at a specific depth, z, (mg m<sup>-3</sup>) and I(z) is PAR at a depth, z. The P-I function is based on two independent parameters: the biomass specific maximum rate of photosynthesis ( $P^B_m$ ) (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup>) and the initial slope ( $\alpha^B$ ) (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup> ( $\mu$  E m<sup>-2</sup> s<sup>-1</sup>)<sup>-1</sup>).

For PP<sub>Aph</sub>, daily PP(z) is calculated as follows:

$$PP(z) = a_{ph}443 \times \left[ \varphi_m \times \frac{K_{\varphi}}{K_{\varphi} + I(z)} \right] \times I(z),$$
 (3)

where  $a_{ph}$  (443) is the coefficient of light absorption by phytoplankton at 443 nm (m<sup>-1</sup>). In Equation (3), the expression in square brackets corresponds to the photosynthesis dependence on underwater illumination expressed in terms of quantum yield ( $\varphi$ ), which is the number of molecules of carbon dioxide assimilated by phytoplankton per absorbed light quantum [45]. In PP<sub>Aph</sub>, the  $\varphi$ -I curve is also presented as the Michaelis-Menten equation, where  $\varphi = \frac{P}{I}$  [46]. For the  $\varphi$ -I curve,  $\varphi$ <sub>m</sub> is the

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maximum quantum yield of photosynthesis, (mg C E<sup>-1</sup>), which is observed at low illumination, and  $K_{\varphi}$  is the half-saturation constant, (E m<sup>-2</sup> day<sup>-1</sup>). Usually, the value of  $K_{\varphi}$  is taken equal to 10 E m<sup>-2</sup> day<sup>-1</sup> [28,32,33,42,47], corresponding to  $\varphi_m = 0.06$  mole C E<sup>-1</sup>. These values of  $\varphi_m$  and  $K_{\varphi}$  were obtained based on the analysis of experimental data; the value of  $\varphi_m$  corresponds to a theoretical maximum [44,48,49].

For PP<sub>PSM</sub> and PP<sub>Aph</sub>, the photosynthetic parameters were chosen to be as close as possible to those observed in the NEA. In situ values of  $P^B_m$ ,  $\alpha^B$ , and  $\varphi_m$  were taken during different seasons (spring, summer, and autumn) and at different depths [32,43,50–55] as average values and used in the models as constant values (Table 2). For PP<sub>Aph</sub>,  $K_{\varphi}$  was determined from the  $\varphi$ -I curve for the widespread coastal phytoplankton diatom species, *Skeletonema costatum*, following [56]. Thus, for a value of  $\varphi_m$  of 0.032 mole C E<sup>-1</sup> (or 347 mg C E<sup>-1</sup> used for  $PP_{eu}$  retrieval),  $K_{\varphi}$  is equal to 138.6  $\mu$  E m<sup>-2</sup> s<sup>-1</sup>. Since, the  $\varphi$ -I function in PP<sub>Aph</sub> is expressed as daily values,  $K_{\varphi}$  is between 4.5–8.0 E m<sup>-2</sup> day<sup>-1</sup> depending on the day length.

**Table 2.** Average and range of photosynthetic parameters used in PP<sub>PSM</sub> and PP<sub>Aph</sub>:  $\alpha^B$  (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup> ( $\mu$  E m<sup>-2</sup> s<sup>-1</sup>)<sup>-1</sup>),  $P^{B_m}$  (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup>), and  $\varphi_m$  (mole C E<sup>-1</sup>).

	$\alpha^{\scriptscriptstyle B}$	$P^{B_m}$	$\varphi_m$
N	12	29	13
mean	0.049	3.316	0.032
SD	0.019	2.153	0.016
min	0.017	0.947	0.010
max	0.078	9.136	0.060

To facilitate the comparison of PP<sub>PSM</sub>, PP<sub>Aph</sub>, and PP<sub>VGPM</sub>, the P-I functions of PP<sub>PSM</sub> and PP<sub>Aph</sub> were integrated over the depth of the euphotic layer (presented in Appendix A, Table A1(a). For each model, it was assumed that the rate of photosynthesis does not depend on a wavelength of the electromagnetic spectrum and is constant during a photoperiod. The Beer–Lambert–Bouguer law, as a function of the surface PAR (I<sub>0</sub>), was used to propagate I(z) over depth, as follows:

$$I(z) = I_0 \times exp(-k_d \times z) \tag{4}$$

where  $k_d$  is the downwelling diffuse attenuation coefficient of solar radiation in water (m<sup>-1</sup>). The euphotic depth was calculated using the following equation, which is derived from Equation (4), assuming that the surface radiation corresponds to 100% and radiation at the euphotic depth is 1%:

$$z_{eu} = \frac{\ln(100)}{k_d} = \frac{4.6}{k_d'} \tag{5}$$

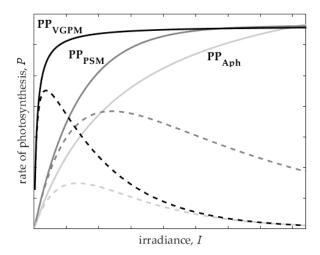
where  $k_d$  for the whole spectrum of PAR was calculated as a function of the satellite product,  $k_d$ , at 490 nm according to the algorithm given in [57].

### 2.5. Implementation of Photo-Inhibition in the Primary Production Models

PP can be modified by photoinhibition because of phytoplankton being subjected to high irradiance or from moving from low to high light conditions. A photoinhibition function was also implemented for each model to test the significance of this phenomenon in the North Atlantic. The mathematical parametrization of photoinhibition is given in Appendix A, Table A1(b), where  $\beta^B$  and  $\beta$  are the photoinhibition parameters, and the superscript,  $\beta^B$ , represents the photoinhibition normalized to biomass.

PP<sup>β</sup><sub>PSM</sub> uses  $β^B$  (mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup> (μ E m<sup>-2</sup> s <sup>-1</sup>)<sup>-1</sup>), whereas PP<sup>β</sup><sub>Aph</sub> uses β ((E m<sup>-2</sup> day<sup>-1</sup>)<sup>-1</sup>). Photoinhibition in PP<sup>β</sup><sub>VGPM</sub> is synonymous with the photoinhibition term in PP<sup>β</sup><sub>Aph</sub>, where  $\overline{I}$  is the weighted average irradiance in the water column (E m<sup>-2</sup> day<sup>-1</sup>). Values of  $β^B$  = 0.01 mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup> (μ E m<sup>-2</sup> s<sup>-1</sup>)<sup>-1</sup> [43] were used for PP<sup>β</sup><sub>PSM</sub>, and β = 0.01 (E m<sup>-2</sup> day<sup>-1</sup>)<sup>-1</sup> [32] for PP<sup>β</sup><sub>Aph</sub> and PP<sup>β</sup><sub>VGPM</sub> were used as constants at all stations. Figure 2 shows the *P-I* functions of the models used. For PP<sup>β</sup><sub>Aph</sub>, the φ-*I* function is multiplied by *I*.

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**Figure 2.** Photosynthesis-irradiance functions of the models used in the study: Solid lines—without photoinhibition, dotted lines—with photoinhibition.

### 2.6. Validation Statistics

Satellite derived estimates of  $PP_{eu}$  for each  $4 \times 4$  km pixels were compared with ship-borne in situ measurements using the statistics described in [58]. Satellite PP<sub>eu</sub> data was extracted from  $3 \times 3$  pixels and averaged over the area to compare with in situ point data. For the daily data, only satellite data with a coefficient of variation (CV) of <0.15 over the  $3 \times 3$  pixels were used. Since the OC-CCI data is from the merger of four Ocean Colour sensors (SeaWiFS, MODIS-Aqua, MERIS, and VIIRS), each of these satellites have different over pass times with respect to the in situ data. The Atlantic Meridional Transect field campaigns were designed to match the overpass of these sensors within  $\pm 3$  h of the different satellite overpasses. Firstly, Taylor [59] and scatter diagrams were used to give the pair correlation (r), Standard Deviation (SD), centre-pattern Root Mean Square Error (c.-p. RMSE), and linear regression (coefficients S and I) between the modelled and in situ  $PP_{eu}$  data. For small rows (less than 28 values), the Spearman r was calculated, while for longer ones the Pearson r was used. In addition, the bias, RMSE, and Absolute Percentage Deviation (APD) were calculated. A one-way analysis of variance (ANOVA) was used to assess whether there were significant differences between the model and in situ  $PP_{eu}$  means.

Statistics were calculated for all stations in individual provinces and by season except for the ARCT province, which were only available during summer.

For the accuracy assessment, results are only included in Table A3 (Appendix B) when the r between the modelled and in situ  $PP_{eu}$  have p-levels of <0.1, 0.05, and 0.01. For all statistics, the highest significant values are highlighted in grey to indicate the most accurate model.

# 2.7. Sensitivity Analysis

To estimate the contribution of each input parameter to the variability in satellite derived  $PP_{eu}$ , a sensitivity analysis was conducted following [14]. The average values of the satellite derived input parameters and the P-I functions were sequentially varied over their natural range.

Firstly, "reference" values of  $PP_{eu}$  were calculated based on averaged values of each of the parameters for the three models without photoinhibition. Then, each parameter was varied sequentially, whilst keeping the other parameters fixed at their average value. The results are presented in the form of box-and-whisker diagrams. Since,  $z_{eu}$  was estimated by  $k_d$  (see Equation (5)), it was not included in the sensitivity analysis, but  $k_d$  was considered instead.

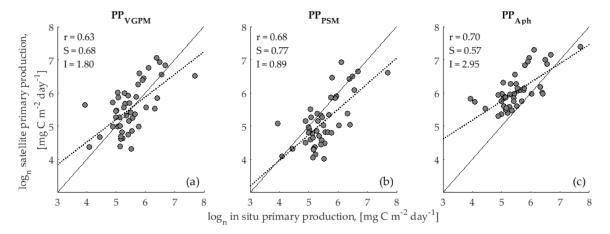
All calculations and visualization of the results were carried out using Matlab R15b.

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# 3. Results

# 3.1. Accuracy Assessment of Primary Production Models

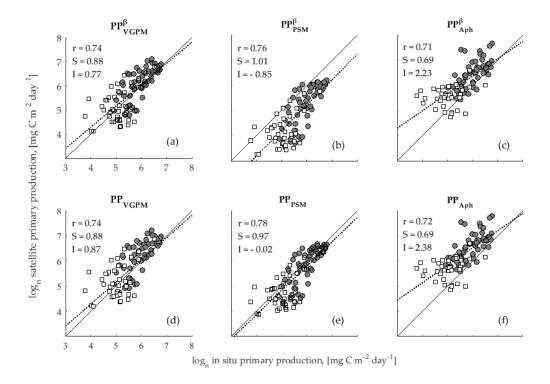
Each model was firstly run using daily OC-CCI data and compared with in situ  $PP_{eu}$  at 46 stations in the NEA (Figure 3). Since the OC-CCI data is available as eight day composites, a secondary accuracy assessment was carried out using these data at the same stations (Appendix B, Table A2) and then including more data (Appendix B, Table A3). Using N = 46, both the daily and eight day modelled data of  $PP_{eu}$  have almost identical results.



**Figure 3.** Scatter plots of log satellite and in situ  $PP_{eu}$  in the NEA using daily OC-CCI data (N = 46) for (a) PPvGPM, (b) PPPSM, and (c) PPAph. Solid line is the 1:1 line; dotted line is the linear regression.

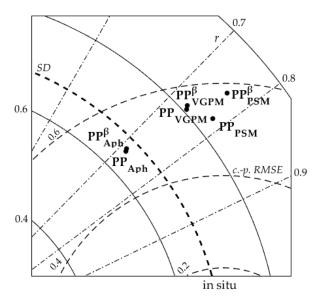
Using the eight day OC-CCI composite data, PP<sub>PSM</sub> was the most accurate model over all stations (Figure 4e). When using the photoinhibition model, PP $_{\text{PVGPM}}$  tended to over-estimate in situ  $PP_{eu}$  at higher values and under-estimate it at lower values (Figure 4a). The net result, however, was that, compared to the other models with photoinhibition, the regression line for PP $_{\text{VGPM}}$  was closest to the 1:1 line and the intercept was the smallest. By comparison, PP $_{\text{Aph}}$  tended to over-estimate in situ  $PP_{eu}$ , especially at low values, which increased the scatter, resulting in a higher intercept (Figure 4c). For PP $_{\text{PSM}}$ , all points were below the 1:1 line, indicating an under-estimate compared to in situ  $PP_{eu}$  (Figure 4b). The scatter was low, however, which increased the percentage variance explained and, though the slope was off set, it was highly correlated with the 1:1. Using no photoinhibition, the trend was the same for PP $_{\text{VGPM}}$  and PP $_{\text{Aph}}$  (Figure 4d,e), with similar statistical results to the models implemented with photoinhibition. For PP $_{\text{PSM}}$ , there was a significant improvement in the model prediction of in situ  $PP_{eu}$ , with the regression line close to the 1:1 line and the intercept was reduced.

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**Figure 4.** Scatter plots of log satellite and in situ  $PP_{eu}$  in the NEA using eight day OC-CCI composites (N = 95); (a) PP<sub>VGPM</sub> with photoinhibition, (b) PP<sub>PSM</sub> with photoinhibition, (c) PP<sub>Aph</sub> with photoinhibition, (d) PP<sub>VGPM</sub> with no photoinhibition, (e) PP<sub>PSM</sub> with no photoinhibition, and (f) PP<sub>Aph</sub> with no photoinhibition. Filled circles are data collected during summer; open squares are autumn data. Solid line is the 1:1 line; dotted line is the linear regression.

A Taylor diagram was used to illustrate the statistical relationships between modelled and in situ  $PP_{eu}$  (Figure 5). All models have a high correlation with in situ  $PP_{eu}$ , with r in the segment from 0.7–0.8. The modelled values, however, have a large c.-p. RMSE, though it did not exceed the SD of the in situ measurements. The smallest c.-p. RMSE was for  $PP_{Aph}$ ,  $PP_{Aph}$ , and  $PP_{PSM}$ .  $PP_{PSM}$  had the largest r, but its SD exceeded that of the in situ  $PP_{eu}$ , whereas the SD of  $PP_{Aph}$  and  $PP_{Aph}$  were closer to the in situ  $PP_{eu}$  SD. Photoinhibition in  $PP_{PSM}$  significantly increased the c.-p. RMSE and created a difference in SD compared to the other models.



**Figure 5.** Taylor diagram of log satellite and in situ  $PP_{eu}$  for the NEA using eight day OC-CCI composites (N = 95). SD—solid arc, centre-pattern RMSE—dotted arc, r—dashed lines with a dot.

Table A3 (Appendix B) gives the statistical results between the modelled and in situ  $PP_{eu}$ . For all stations, PP<sub>PSM</sub> was the most accurate in four out of eight statistical tests. It slightly under-estimated  $PP_{eu}$  (bias = -0.21) and explained 61% of the variance in in situ measurements (coefficient of determination ( $r^2$ ) = 0.61). There was an 8% Absolute Percentage Difference (APD) between PP<sub>PSM</sub> and in situ  $PP_{eu}$ , and this model also had the smallest RMSE. PP<sub>PSM</sub> exhibited a better performance in summer when it explained 58% of the variance in in situ  $PP_{eu}$ , compared to autumn when it was 25%. In addition, PP<sub>PSM</sub> did not always accurately capture the spatial variability in in situ  $PP_{eu}$  since the SD was 1.5 times higher than the SD of in situ  $PP_{eu}$ , except for the ARCT and NADR provinces in summer.

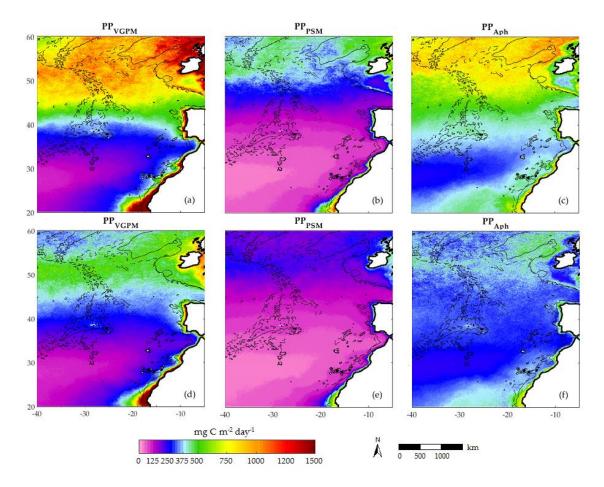
By comparison,  $PP^{\beta_{VGPM}}$  was the most accurate in just two out of eight statistical tests, though it was worse in summer (in five out of eight statistical tests). Both  $PP^{\beta_{VGPM}}$  and  $PP_{VGPM}$  slightly over-estimated in situ  $PP_{eu}$  (bias = 0.13–0.21) except in the NATR province, where it underestimated in situ values. On average,  $PP^{\beta_{VGPM}}$  and  $PP_{VGPM}$  diffed from in situ  $PP_{eu}$  by 8% and 9% (APD), respectively. For all stations,  $r^2$  for  $PP^{\beta_{VGPM}}$  and  $PP_{VGPM}$  explained 54 to 55% of the variance of in situ  $PP_{eu}$ . The  $PP^{\beta_{VGPM}}$  and  $PP_{VGPM}$  was the most accurate in the NATR province and during summer, and the statistical errors were small.

Both  $PP_{Aph}$  and  $PP_{Aph}$  over-estimated in situ  $PP_{eu}$  (bias = 0.51–0.64) compared to the other models, especially at low values (Appendix B, Table A3). APD for  $PP_{Aph}$  and  $PP_{Aph}$  were 11% and 13%, respectively.  $PP_{Aph}$  and  $PP_{Aph}$ , both captured the spatial variability in in situ  $PP_{eu}$  for all data, and the SD was similar to that of in situ  $PP_{eu}$ . In addition, the c.-p. RMSE for  $PP_{Aph}$  and  $PP_{Aph}$  was small compared to the other models, though they were less accurate in autumn compared to summer.

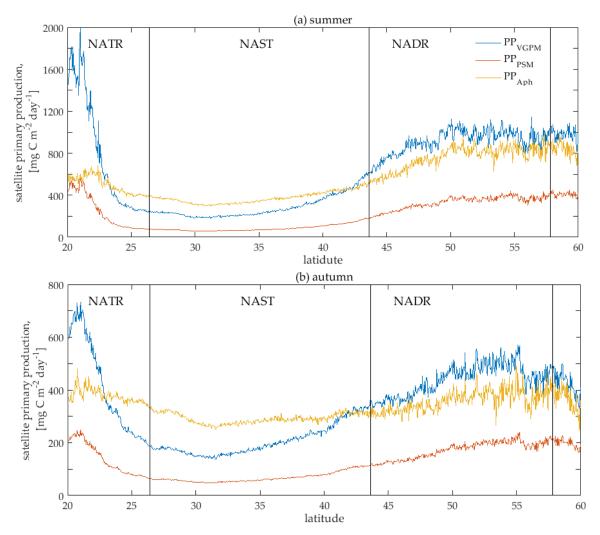
### 3.2. Satellite PPeu Images

The spatial and temporal differences between the models are illustrated in satellite images during two different periods: in summer (average June–August, 1998–2011; Figure 6a–c) and autumn (average September–November, 1998–2011; Figure 6d–f). During summer in the NADR, the PPvGPM gave the highest values (500–1500 mg C m<sup>-2</sup> d<sup>-1</sup>), whereas the PPrsM was 250–700 mg C m<sup>-2</sup> d<sup>-1</sup> and PPAph was 500–1000 mg C m<sup>-2</sup> d<sup>-1</sup>. By comparison, in the NADR autumn, the range in PPvGPM was 300–700 mg C m<sup>-2</sup> d<sup>-1</sup>, PPrsM was 100–300 mg C m<sup>-2</sup> d<sup>-1</sup>, and PPAph was 250–500 mg C m<sup>-2</sup> d<sup>-1</sup>. In contrast, in the NAST in summer, PPAph gave the highest values (250–700 mg C m<sup>-2</sup> d<sup>-1</sup>), which were lower in autumn (200–350 mg C m<sup>-2</sup> d<sup>-1</sup>). The PPvGPM returned similar values in summer and autumn, which were 125–500 mg C m<sup>-2</sup> d<sup>-1</sup>, and PPrsM was 0–350 mg C m<sup>-2</sup> d<sup>-1</sup>. Similarly, in the NATR, PPAph also returned the highest values in both summer and autumn, PPrsM yielded the lowest (<125 mg C m<sup>-2</sup> d<sup>-1</sup>), and PPvGPM were 50–250 mg C m<sup>-2</sup> d<sup>-1</sup>.

To illustrate further the spatial and temporal differences between the models, we extracted data from each model at every 4 km along a transect at 20°W from 20° to 60°N (Figure 7). In the NADR, there was a 182% and 157% Relative Percentage Deviation (RPD) between PP<sub>VGPM</sub> and PP<sub>PSM</sub> in summer and autumn, respectively, and 132% and 113% RPD between PP<sub>Aph</sub> and PP<sub>PSM</sub>, respectively. In the NAST, the differences increased to a 228% and 203% difference between PP<sub>VGPM</sub> and PP<sub>PSM</sub> and a 359% and 351% for PP<sub>Aph</sub> and PP<sub>PSM</sub> in summer and autumn, respectively. For the NATR, there was a 222% and 197% RPD between PP<sub>VGPM</sub> and PP<sub>PSM</sub> and a 208% and 224% for PP<sub>Aph</sub> and PP<sub>PSM</sub> in summer and autumn, respectively.



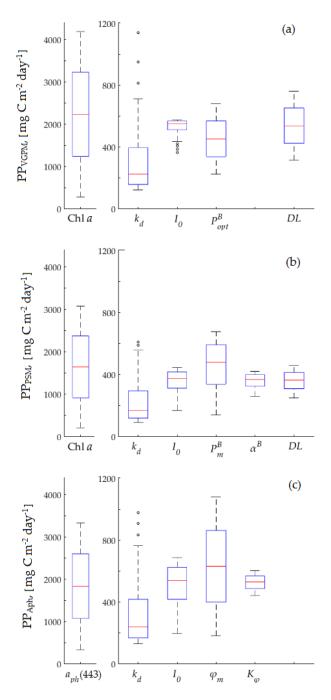
**Figure 6.** Spatial distribution of satellite  $PP_{eu}$  (mg C m<sup>-2</sup> day<sup>-1</sup>) using OC-CCI climatology for 1998–2011 for; (a) PPvgpm in summer (June–August), (b) PPpsm in summer, (c) PPaph in summer, (d) PPvgpm in autumn (September–November), (e) PPpsm in autumn, and (f) PPaph in autumn.



**Figure 7.** Spatial transects of satellite PP<sub>VGPM</sub>, PP<sub>PSM</sub>, and PP<sub>Aph</sub> (mg C m<sup>-2</sup> day<sup>-1</sup>) along 20°W from 20 to 60°N for (a) summer (June–August) and (b) autumn (September–November) using OC-CCI climatology for 1998–2011.

# 3.3. Model Sensitivity Analysis

For all three models, the largest contribution to the variability in PP was the remote sensing ocean colour product used, which for PPvgpm and PPpsm was Chl a, and for PPaph was  $a_{ph}$  (443) (Figure 8). The next most important parameter depends on the model used. For PPvgpm,  $k_d$  for these waters caused the greatest error. For PPpsm, both  $P^B_m$  and  $k_d$  produced the greatest error and for PPaph it was  $\varphi_m$ . The third most important parameter for the PPvgpm was  $P^B_{opt}$  and DL, for PPpsm it was  $I_0$  and  $I_0$ , and for PPaph it was  $I_0$  and  $I_0$ . The smallest contribution to the variability in PPvgpm was from  $I_0$ , but for PPpsm it was  $\alpha^B$ , and for PPaph it was  $K_{\Phi}$ .



**Figure 8.** Box-whisker plots for the sensitivity analysis on (a) PPv<sub>GPM</sub>, (b) PP<sub>PSM</sub>, and (c) PP<sub>Aph</sub>: For the left hand y-axis, PP is from 0–4400 mg C m<sup>-2</sup> day<sup>-1</sup>; right y-axis PP is from 0–1200 mg C m<sup>-2</sup> day<sup>-1</sup>. *Chl*<sub>90</sub> is the average Chl *a* concentration over the first optical depth,  $k_d$  is the downwelling diffuse attenuation coefficient,  $I_0$  is the daily surface PAR,  $P^B_{opt}$  is the biomass-specific optimum rate of photosynthesis, DL is the day length,  $P^B_m$  is the biomass-specific maximum rate of photosynthesis,  $\alpha^B$  is the initial slope of the P-I curve,  $a_{ph}$  (443) is the coefficient of light absorption by phytoplankton at 443 nm,  $\varphi_m$  is the maximum quantum yield of photosynthesis, and  $K_\varphi$  is the half-saturation constant of the  $\varphi$ -I curve. The rectangular boundaries are the first and third quartiles (the 25th and 75th percentiles) of the modelled  $PP_{eu}$  obtained by changing each parameter in turn. The line in the rectangle is the median (50th percentile) of the sample, the edges of the "whiskers" are the size of the sample (minimum and maximum of the sample), and the symbol, "o", represents extreme values. The larger the size of the box and whiskers, the greater the contribution of the parameter to the variability in the modelled  $PP_{eu}$ .

### 4. Discussion

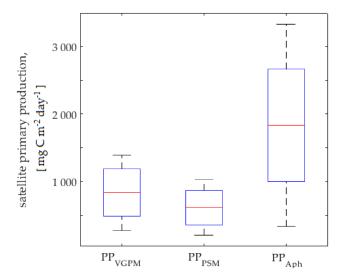
# 4.1. Validation of Primary Production Models

Validation of PPvgpm in the North East Atlantic Ocean has already been carried out by a number of independent studies in this and neighbouring seas [7,9,10,12–14,60,61]. These studies report that PPvgpm over-estimates in situ  $PP_{eu}$ , which we also observed in the NEA, especially in NADR. The only exception is in the NATR province, where PPvgpm is lower than in situ  $PP_{eu}$ . As previously highlighted [7,14], the use of  $P^{B_{opt}}$  to simulate model variations in photosynthesis as a function of SST may not consider the displacement of the layer of optimum light conditions for photosynthesis to lower depths, which can be caused by high surface PAR [62].

According to [10], PPvGPM is closer to the actual global PP than PPPSM, which tends to under-estimate global PP by approximately 13%. In our study, PPvGPM are slightly higher than PPPSM in summer and autumn in the Atlantic Ocean, especially in the NADR, NATR, and NAST (Appendix B, Table A3). Only in the ARCT province, where the waters are typically mesotrophic with a medium range Chl *a* [36], did both models converge.

Validation of PP<sub>Aph</sub> in shelf waters off Argentina [28] showed a high correlation with in situ  $PP_{eu}$  (r = 0.9), but there was a tendency for the model to under-estimate  $PP_{eu}$ , which is due to the use of the constant,  $K_{\varphi}$  = 10 E m<sup>-2</sup> day<sup>-1</sup>. In our study, we allowed  $K_{\varphi}$  to vary between 4.5–8.0 E m<sup>-2</sup> day<sup>-1</sup> depending on the day length. PP<sub>Aph</sub> slightly overestimated in situ  $PP_{eu}$ , especially in autumn, which was also observed by [28] in Argentinian shelf waters.

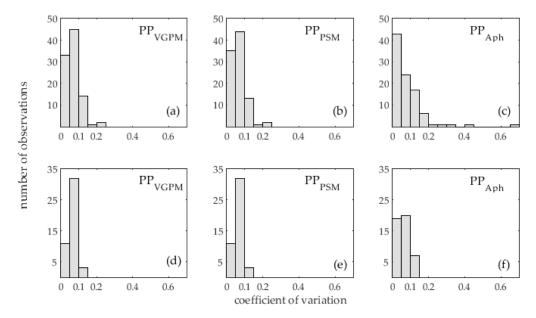
PP<sub>Aph</sub> in the open ocean waters of the South West Atlantic is also more accurate than PP<sub>VGPM</sub> ([33]; PP<sub>Aph</sub>  $r^2$  = 0.74, PP<sub>VGPM</sub>  $r^2$  = 0.44). Theoretically, this is due to the advantage of deriving  $a_{ph}$  (443) directly from  $R_{rs}$  rather than using an empirical band ratio to derive Chl a [63]. However, this depends on how accurately both  $a_{ph}$  (443),  $a_{CDOM}$  (443), and  $a_{det}$  (443) can be determined from  $R_{rs}$  (443). If there is an error in deriving  $a_{CDOM}$  (443) and  $a_{det}$  (443) it will propagate to  $a_{ph}$  (443), which can, in turn, impact PP<sub>Aph</sub>. Based on the sensitivity analysis,  $a_{ph}$  (443) causes a greater error in  $PP_{eu}$  compared to Chl a, which is used in PP<sub>PSM</sub> (Figure 8). Also for PP<sub>Aph</sub>, an accurate estimate of the photosynthetic parameters ( $\varphi_m$  and  $K_\varphi$ ) is also vital in improving the performance of this model [42]. We also observed that  $\varphi_m$  has a large impact on PP<sub>Aph</sub>, but  $K_\varphi$  has the least effect (Figure 8). In our study, PP<sub>Aph</sub> was more accurate in the NADR province ( $r^2$  = 0.65), though it still over-estimated in situ  $PP_{eu}$  and was worse in the ARCT province in summer (Appendix B, Table A3), though this was based on few points. Furthermore, Figure 9 shows that  $a_{ph}$  (443) causes a significantly greater error in PP<sub>Aph</sub> in the ARCT compared to Chl a used in the PP<sub>VGPM</sub> and PP<sub>PSM</sub>.



**Figure 9.** Box-and-whisker diagram from the sensitivity analysis for PP<sub>VGPM</sub>, PP<sub>PSM</sub>, and PP<sub>Aph</sub> in the ARCT province. Chl a and  $a_{ph}$  (443) were changed in the models based on the variability in values at stations.

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Standard satellite validation protocols were used to assess the accuracy of the PP models. The satellite data were averaged over  $3 \times 3$  pixels with CV < 0.15 for the majority of the match-ups (Figure 10), which illustrates that the quality of the satellite data was good. For the daily data, the CV for the three models was always <0.15. For the eight day composites, there were two data points for PPvgpm and PPpsm where the CV > 0.15 and six points for PPaph, which were mainly from the ARTC province.



**Figure 10.** Histograms of the coefficient of variation of satellite  $PP_{eu}$  for the area of  $12 \times 12$  km ( $3 \times 3$  pixels) for each match-up station analysed using eight day composite data (N = 95) for (a) PPvgpm; (b) PPpsm; and (c) PPaph and using daily data (N = 46) with (d) PPvgpm; (e) PPpsm; and (f) PPaph.

We found that all models performed better in summer than in autumn. Since Chl a is the main contributor to the overall error in modelled  $PP_{eu}$ , it may suggest that the error in the autumn values arise from an error in the satellite Chl a. However, numerous studies have shown for the North Atlantic that satellite Chl a is accurate during this season (e.g., [14]).

According to the sensitivity analysis,  $P^B_m$  and  $\varphi_m$  are the second parameters that determine the overall error of  $PP_{eu}$  retrieval. The values of  $P^B_m$  and  $\alpha^B$  used in the study for the NEA (Table 2) correspond with values from a global database of in situ photosynthesis parameters presented in [64]. Mean values of  $P^B_m$  and  $\alpha^B$  averaged over four provinces (ARCT, NADR, NAST, and NATR) and over spring, summer, and autumn from this database are 3.25 mg C (mg Ch)<sup>-1</sup> h<sup>-1</sup> and 0.042 mg C (mg Chl)<sup>-1</sup> h<sup>-1</sup> ( $\mu$  E m<sup>-2</sup> s<sup>-1</sup>)<sup>-1</sup>, respectively. A value of  $\varphi_m$  = 0.03 mol C E<sup>-1</sup> used in our study is characteristic of sunlit regions and was also used by [9] for retrieving  $PP_{eu}$  in different parts of the global ocean. An identical value of  $\varphi_m$  was obtained in waters off the southern California coast (30–34°N) [65]. Errors in PP<sub>PSM</sub> may arise from inaccurate derivation of the photosynthetic parameters,  $P^B_m$  and  $\alpha^B$ , which can vary over space, time, and depth depending on the dominant phytoplankton community.

To calculate PP<sub>PSM</sub>, we used 29 values of  $P^B_m$ , and 20 values during autumn. When a  $P^B_m$  value, typical for autumn, of 2.36 mg C (mg Ch)<sup>-1</sup> h<sup>-1</sup> was used, it did not improve the correlation between in situ and modelled  $PP_{eu}$  ( $r^2 = 0.23$ ). Spatial changes in the species composition of phytoplankton communities and the related changes in photosynthetic rates may also cause a weak correlation between the modelled and in situ  $PP_{eu}$  [66]. In addition, we input the Chl a and photosynthetic parameters as constant values over depth, which may not be representative of the vertical variability of these parameters in autumn and may therefore contribute to the error in the model. Kahru et al. [42] validated the  $PP_{Aph}$  algorithm in the California Current, and showed that accounting for the vertical profile of  $a_{ph}$  (443) within the euphotic zone improved the correlation between  $PP_{Aph}$  and in situ  $PP_{eu}$ . The coefficient of determination ( $r^2$ ) increased from 0.28 to 0.56. Errors in  $R_{rs}$ , which is used

to calculate  $a_{ph}$  (443), could be the main cause of error in the retrieval of  $PP_{eu}$ . These estimates can vary widely between radiometers (15–18% for  $R_{rs}$  at 443 nm).

The tendency in the scatter plots is that PPvgpm over-estimates PPeu in summer and under- and over-estimates  $PP_{eu}$  in autumn,  $PP_{Aph}$  over-estimates  $PP_{eu}$  in summer, especially in autumn, and that PP<sub>PSM</sub> shows good agreement in summer, but has a tendency to under-estimate PP<sub>eu</sub> in autumn (Figure 4). Visual examination of the resulting mapped images provides a qualitative analysis of whether the tendencies shown in the scatter plots correspond over wider spatial and temporal scales. The extent of the differences between the models in the mapped images and data extracted from these over a transect from 20 to 60°N illustrate that, of the three models, PPvgpm gives the highest PPeu in summer and autumn in the NATR and NADR, whereas the PPAph has the highest PPeu in the NAST, especially in autumn (Figure 7). The PPPSM consistently has the lowest PPeu, but is closest to the in situ  $PP_{eu}$ , but the differences between  $PP_{PSM}$  and the other models are less in autumn. The outliers in the scatter plot for the PP<sub>Aph</sub> model (Figure 4f) may suggest that, in the higher PP waters of the NADR, this model would return the highest  $PP_{eu}$ . The images and transect, however, show that PPvgpm gives the highest PPeu, which is not reflected in the corresponding scatter plot (Figure 4d). The tendency for the PPvgpm to over-estimate  $PP_{eu}$  at higher values in the scatter plot represents large areas of the NADR and NATR during summer in the corresponding mapped image (Figure 6a).

The results suggest that the PP<sub>PSM</sub> model with the OC-CCI data is accurate for the NEA. The model is available as an OC-CCI product, but requires further testing to assess its accuracy over other basins and the entire global ocean.

# 4.2. Effect of Photoinhibition in PP Models

Photoinhibition describes the decrease in the photosynthetic rate with increasing irradiance [67]. This phenomenon is dependent on the phytoplankton community and its adaptation to the light conditions in the water column. It is usually greatest when phytoplankton is confined to surface high light conditions or when it moves quickly from low to high light [34]. Under such high light conditions, photorespiration can be activated, which is accompanied by utilization of assimilated carbon and, therefore, a decrease in the efficiency of photosynthesis [68]. For different phytoplankton communities, it has been shown experimentally that this effect varies as a function of depth and solar radiation at the surface and photo-adaptation of phytoplankton [69,70]. For example, shade-adapted communities saturate quickly and reach photoinhibition faster than high-light adapted communities.

Since the total amount of incoming solar radiation decreases with the distance from the equator and is dependent on the time of the year, the difference in the estimates of  $PP_{eu}$ , both with and without photoinhibition, also decrease with increasing latitude, and has a distinctive seasonal variation associated with the variability of incoming solar radiation during the year. The maximum difference in the estimates of  $PP_{eu}$  is in the second half of June, when solar radiation is at a maximum. The smallest difference is at the end of December or beginning of January, when solar radiation reaches a minimum. Since both states can occur in the tropical waters of the NAST and NATR provinces where weak wind stress and density gradient can prevent the migration of the phytoplankton community and, trap it at high light conditions for a significant proportion of the day, including photoinhibition may improve model performance. We found that the  $PP_{VGPM}$ , for example, performed better in the NAST during summer.

Table 3 illustrates the mathematical representation of photoinhibition in the models used, which does not differ significantly between each model. Photoinhibition had a greater effect on PP<sub>PSM</sub> compared to the other two models, however. Usually, photoinhibition in the models is presented as an exponentially decreasing term [70]. Photoinhibition in PP<sup> $\beta$ </sup><sub>PSM</sub> is implemented as the *P-I* curve increases at low light [43]. After integration over the euphotic depth, its mathematical representation can cause large differences between PP<sup> $\beta$ </sup><sub>VGPM</sub> and PP<sup> $\beta$ </sup><sub>Aph</sub>, even though the photoinhibition functions are identical. According to [32], the photoinhibition term in PP<sup> $\beta$ </sup><sub>Aph</sub> can reduce the values of the modelled *PP*<sub>eu</sub> from 10% to 55% depending on the light intensity.

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**Table 3.** Relative percentage difference between modelled  $PP_{eu}$  with and without photoinhibition. The negative sign "-" indicates how much photoinhibition reduces  $PP_{eu}$ .

Model	Mean	Min	Max
PPvgpm	-7	-2	-10
$PP_{\text{PSM}}$	-42	-20	-50
$\operatorname{PP}_{Aph}$	-11	-4	-15

### 5. Conclusions

Three satellite models of PP were processed using OC-CCI data, with and without photoinhibition, and compared against ship-borne in situ 14C measurements from the NEA. Validation of the models using daily OC-CCI data (N = 46) showed that the PPrsM and PPvgPM had a similar accuracy, whereas the PPAph was not suitable for the region. Using eight day OC-CCI composite data, the number of match-ups was doubled (N = 95) and PPPSM was more accurate over all provinces. For individual provinces, the PPPSM was more accurate in the ARCT and NADR regions whereas the PPvgpm was more accurate in the NAST and NATR regions. The PPaph was the least accurate model for the NEA, which was due to errors in the  $a_{ph}$  (443) OC-CCI product. The use of a photoinhibition function in the PPvgpm and PPaph had little effect on the model performance, whereas, in the PPPSM, photoinhibition reduced the accuracy of the model, especially at lower range of the values. The performance of the models also varied seasonally; all models were more accurate during summer and less accurate during autumn. Over all provinces, the PPvgpm with photoinhibition was the most accurate during summer, and the PPPSM with no photoinhibition was more accurate during autumn. Mapped images illustrated that differences between models were far greater than indicated by the scatter plots, especially for PPAph and PPvGPM in the NATR and NADR provinces. A sensitivity analysis indicated that the primary ocean colour product (Chl a for PPPSM and PPvgpm; aph (443) for PPaph) caused the greatest variability in model performance, followed by the photosynthetic terms,  $P^{B}_{m}$  and  $\varphi_{m}$ , in the PP<sub>PSM</sub> and PP<sub>Aph</sub> models, respectively, and  $P^{B}_{opt}$  and  $k_{d}$  in the PPvgpm. Improvement in the accuracy of these input parameters will ultimately lead to an improvement in satellite PP algorithms for this region.

**Author Contributions:** Conceptualization was derived by P.L., I.B. and V.B.; methodology was derived and implemented by P.L. and G.T.; software was developed by P.L. and I.B.; validation was performed by P.L. and G.T.; formal analysis was conducted by P.L. and G.T.; investigation was performed by P.L., G.T., I.B. and V.B.; resources to support the research were provided by P.L., G.T. and I.B.; writing—original draft preparation was done by P.L. and G.T.; writing—review & editing, was done by P.L., G.T., I.B. and V.B.; visualization (data, tables, & figures) was done by P.L.; supervision was from I.B., G.T. and V.B.; project administration was from P.L.; funding acquisition to support the research were sought by P.L. and I.B.

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**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

# Appendix A

**Table A1.** Mathematical equations for PP<sub>PSM</sub> and PP<sub>Aph</sub> after integrating their *P-I* functions over the euphotic depth; (a)  $E_1$  is an exponential integral defined as:  $E_1(x) = \int_x^\infty \frac{e^{-t}}{t} dt$ ; (b) mathematical representation of photoinhibition in the models tested.

	(a) Mathematical Equations for the Models									
Model	With Photoinhibition	Without Photoinhibition								
PP <sub>PSM</sub>	$PP_{eu} = \frac{Chl_{90} \times DL \times P_m^B}{k_d} \left[ \left( E_1 \left( \frac{I_0 \beta^B}{P_m^B} exp(-k_d z_{eu}) \right) - E_1 \left( \frac{I_0 \beta^B}{P_m^B} \right) \right) - \left( E_1 \left( \frac{I_0 (\alpha^B + \beta^B)}{P_m^B} exp(-k_d z_{eu}) \right) - E_1 \left( \frac{I_0 (\alpha^B + \beta^B)}{P_m^B} \right) \right) \right]$	$\begin{aligned} & PP_{eu} \\ & = \frac{Chl_{90} \times DL \times P_m^B}{k_d} \\ & \times \left[ E_1 \left( \frac{\alpha^B \times I_0}{P_m^B} \right) + (k_d \times z_{eu}) \right. \\ & \left E_1 \left( \frac{\alpha^B \times I_0}{P_m^B} \times exp(-k_d \times z_{eu}) \right) \right] \end{aligned}$								
$PP_{Aph}$	$PP_{eu} = \frac{aph443 \times \varphi_m \times K_{\varphi}}{k_d} \times exp(\beta K_{\varphi}) \times \left[ E_1 \left( \beta \left( K_{\varphi} + I_0 exp(-k_d z_{eu}) \right) \right) - E_1 \left( \beta \left( K_{\varphi} + I_0 \right) \right) \right]$	$PP_{eu} = \frac{aph443 \times \varphi_m \times K_{\varphi}}{k_d} \times \left[ \ln(K_{\varphi} + I_0) - \ln(K_{\varphi} + I_0 \times \exp(-k_d \times z_{eu})) \right]$								
	(b) Mathematical Representation of Pho	otoinhibition								
Model	Photoinhibition Term	Reference								
PP <sub>PSM</sub>	$exp\left(-rac{eta^B}{P_m^B}I(z) ight)$	[43]								
PPAph	$exp(-\beta \times I(z))$	[32]								
PPvgpm	$\bar{I} = \frac{exp(-\beta \times \bar{I})}{\sum_{z=0}^{z=u} I(z)dz} = 0.215I_0$	this paper								

# Appendix B

**Table A2.** Validation statistics for each model using (a) daily and (b) eight day composite OC-CCI data (N = 46). The numbers at the head of the table indicate statistics for one test. The most significant statistical values are highlighted in grey. r is the Pearson pair correlation coefficient, S is the Slope and S is the Intercept of the linear regression coefficients, RMSE is the Root Mean Square Error, and APD is the Absolute Percentage Deviation.

(a) Dalla M. 46		1	2	3		4	5	6
(a) Daily, N = 46	r	p-Level	Centre-Pattern RMSE	S	I	Bias	RMSE	APD, %
$PP^{\beta_{VGPM}}$	0.63	p < 0.001	0.60	0.68	1.73	-0.04	0.60	9.2
$PP_{VGPM}$	0.63	<i>p</i> < 0.001	0.60	0.68	1.80	0.05	0.61	9.3
$\mathrm{PP}^{eta}_{PSM}$	0.69	<i>p</i> < 0.001	0.57	0.79	0.16	-0.99	1.15	18.8
$PP_{PSM}$	0.68	<i>p</i> < 0.001	0.57	0.77	0.89	-0.38	0.68	9.9
$\mathrm{PP}^{eta}_{\mathrm{Aph}}$	0.71	<i>p</i> < 0.001	0.48	0.56	2.86	0.42	0.63	9.8
$PP_{Aph}$	0.70	<i>p</i> < 0.001	0.48	0.57	2.95	0.56	0.74	11.8
(b) Eight day composites, $N = 46$								
$PP^{\beta_{VGPM}}$	0.65	<i>p</i> < 0.001	0.56	0.66	1.80	-0.06	0.57	8.6
$PP_{VGPM}$	0.66	p < 0.001	0.56	0.67	1.86	0.03	0.56	8.6
$\mathrm{PP}^{eta}_{PSM}$	0.69	p < 0.001	0.57	0.79	0.16	-0.99	1.15	18.8
$PP_{PSM}$	0.73	p < 0.001	0.49	0.75	0.98	-0.40	0.64	9.5
$\mathrm{PP}^{eta}_{Aph}$	0.75	p < 0.001	0.45	0.53	3.00	0.40	0.60	9.4
$PP_{Aph}$	0.73	p < 0.001	0.46	0.54	3.09	0.53	0.70	11.4

**Table A3.** Validation statistics for each model for the whole region and for individual biogeographical provinces using eight day composite OC-CCI data. The numbers at the head of the table indicate statistics for one test. The most significant statistical values are highlighted in grey. r is the Pearson or Spearman pair correlation coefficient (For N > 28, Pearson was used; for N < 28 Spearman was used, but  $r^2$  was not computed),  $r^2$  is the coefficient of determination, SD is the Standard Deviation, RMSE is the Root Mean Square Error, S is the Slope, I is the Intercept of the linear regression coefficients, APD is the Absolute Percentage Deviation, and F and P are ANOVA (Analysis of variance) coefficients. Those models, for which the significance of P was more than 0.1, were not included into the analysis and are not presented in the Table for summer stations in NAST (N = 16) and ARCT (N = 12), autumn stations in NADR (N = 6) and NAST (N = 24).

All Ct. C. N. OF	1		2		3	4	5	6	7		8
All Stations, $N = 95$	SD	r	<b>r</b> <sup>2</sup>	S	I	bias	RMSE	Centre-Pattern RMSE	APD, %	ANOVA, F	ANOVA, p
In situ PPeu	0.68										
$PP^{\beta_{VGPM}}$	0.81	0.74	0.54	0.88	0.77	0.13	0.57	0.56	9	1.4	0.237
$PP_{VGPM}$	0.81	0.74	0.55	0.88	0.87	0.21	0.59	0.55	9	3.9	0.051
$\mathrm{PP}^{eta_{\mathrm{PSM}}}$	0.90	0.76	0.58	1.01	-0.85	-0.78	0.97	0.58	15	45.6	0.000
$PP_{PSM}$	0.84	0.78	0.61	0.97	-0.02	-0.21	0.56	0.52	8	3.6	0.060
$\mathrm{PP}^{eta_{\mathrm{Aph}}}$	0.66	0.71	0.51	0.69	2.23	0.51	0.71	0.50	11	27.0	0.000
$PP_{Aph}$	0.65	0.72	0.52	0.69	2.38	0.64	0.81	0.50	13	44.2	0.000
NADR, $N = 28$											
In situ <i>PPeu</i>	0.54										
$PP^{\beta_{VGPM}}$	0.35	0.73	0.53	0.47	3.70	0.48	0.61	0.38	9	15.1	0.000
PPvgpm	0.35	0.74	0.54	0.48	3.69	0.55	0.66	0.37	10	19.5	0.000
$\mathrm{PP}^{eta_{\mathrm{PSM}}}$	0.43	0.79	0.62	0.62	1.91	-0.40	0.52	0.34	7	9.1	0.004
$PP_{PSM}$	0.43	0.82	0.67	0.64	2.25	0.10	0.33	0.32	5	0.5	0.468
$\mathrm{PP}^{eta}_{\mathrm{Aph}}$	0.37	0.80	0.63	0.54	3.18	0.41	0.53	0.34	8	10.4	0.002
$\operatorname{PP}_{Aph}$	0.38	0.80	0.65	0.56	3.18	0.52	0.62	0.33	9	16.7	0.000
NAST, $N = 40$											
In situ <i>PPeu</i>	0.57										
$PP^{\beta_{VGPM}}$	0.55	0.52	0.27	0.51	2.59	0.07	0.55	0.55	9	0.32	0.572
$PP_{VGPM}$	0.56	0.54	0.29	0.53	2.57	0.17	0.57	0.54	10	1.68	0.199
$\mathrm{PP}^{eta_{\mathrm{PSM}}}$	0.49	0.51	0.26	0.44	1.88	-0.97	1.10	0.52	19	65.16	0.000
$PP_{PSM}$	0.50	0.55	0.30	0.48	2.29	-0.34	0.61	0.51	10	7.76	0.008
$\mathrm{PP}^{eta_{\mathrm{Aph}}}$	0.42	0.42	0.17	0.31	4.06	0.55	0.78	0.55	14	23.80	0.000
$\operatorname{PP}_{Aph}$	0.44	0.43	0.19	0.34	4.06	0.70	0.89	0.55	16	36.57	0.000
NATR, N = 14											
In situ PPeu	0.33										
$PP^{\beta_{VGPM}}$	0.66	0.51		1.49	-3.22	-0.49	0.68	0.46	11	5.78	0.024
$PP_{VGPM}$	0.66	0.51		1.49	-3.13	-0.39	0.61	0.47	10	3.58	0.069
$\mathrm{PP}^{eta}_{\mathrm{PSM}}$	0.62	0.57		1.44	-3.89	-1.44	1.50	0.42	26	54.11	0.000
$PP_{PSM}$	0.63	0.54		1.44	-3.22	-0.77	0.88	0.43	14	15.14	0.001
All Summer Stations, $N = 56$											
In situ PPeu	0.49										
$PP^{\beta_{VGPM}}$	0.68	0.73	0.54	1.01	0.07	0.14	0.48	0.46	7	1.5	0.218
$PP_{VGPM}$	0.66	0.73	0.53	0.98	0.35	0.23	0.51	0.45	7	4.3	0.041
$\mathrm{PP}^{eta_{\mathrm{PSM}}}$	0.80	0.76	0.58	1.25	-2.20	-0.74	0.91	0.54	13	33.9	0.000
PPpsm	0.72	0.76	0.58	1.11	-0.84	-0.17	0.50	0.47	7	2.1	0.146

1	0.53	0.66									0.000
		0.00	0.44	0.72	2.32	0.64	0.77	0.42	11	43.0	0.000
ARCT, $N = 12$											
In situ PPeu	0.40										
$\mathrm{PP}^{eta}_{\mathrm{PSM}}$	0.28	0.56		0.45	3.01	-0.37	0.47	0.30	7	6.3	0.020
PP <sub>PSM</sub>	0.28	0.55		0.46	3.37	0.09	0.31	0.30	4	0.4	0.547
NADR, N = 22											
In situ PPeu	0.39										
$PP^{\beta_{VGPM}}$	0.30	0.57		0.40	4.09	0.39	0.52	0.34	7	13.3	0.001
PPvgpm	0.29	0.55		0.40	4.22	0.46	0.58	0.34	8	18.9	0.000
$\mathrm{PP}^{\beta_{\mathrm{PSM}}}$	0.34	0.53		0.53	2.45	-0.47	0.57	0.33	8	16.9	0.000
$PP_{PSM}$	0.32	0.59		0.50	3.14	0.05	0.32	0.31	5	0.2	0.669
$\mathrm{PP}^{eta_{\mathrm{Aph}}}$	0.28	0.58		0.46	3.68	0.33	0.45	0.30	6	9.7	0.003
$PP_{Aph}$	0.28	0.62		0.45	3.88	0.45	0.54	0.30	8	18.6	0.000
NATR, $N = 6$											
In situ PPeu	0.19										
$PP^{\beta}_{VGPM}$	0.21	0.81		0.80	0.57	-0.50	0.52	0.15	9	15.4	0.003
PPvgpm	0.21	0.81		0.76	0.93	-0.39	0.42	0.16	7	9.2	0.012
All Autumn Stations, N = 38											
In situ PPeu	0.55										
$PP^{\beta}_{VGPM}$	0.68	0.41	0.16	0.50	2.64	0.10	0.69	0.68	12	0.5	0.477
PPvgpm	0.67	0.41	0.17	0.50	2.70	0.18	0.69	0.67	12	1.6	0.213
$\mathrm{PP}^{eta_{\mathrm{PSM}}}$	0.65	0.47	0.22	0.54	1.42	-0.87	1.07	0.62	18	38.6	0.000
PPpsm	0.61	0.50	0.25	0.55	1.97	-0.28	0.65	0.58	11	4.4	0.039
NATR, $N = 8$											
In situ PPeu	0.38										
$PP^{\beta_{VGPM}}$	0.84	0.69		1.67	-4.29	-0.48	0.77	0.60	12	1.9	0.190
PPvgpm	0.85	0.69		1.69	-4.29	-0.39	0.72	0.60	12	1.2	0.286
$\mathrm{PP}^{\beta_{\mathrm{PSM}}}$	0.77	0.64		1.53	-4.38	-1.38	1.48	0.53	25	18.0	0.001
PPpsm	0.78	0.69		1.58	-4.00	-0.72	0.90	0.54	13	4.8	0.046

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### References

- 1. Falkowski, P. Ocean productivity from space. *Nature* **1988**, 335, 205.
- 2. Steemann Nielsen, E. The use of radio-active carbon (C<sup>14</sup>) for measuring organic production in the sea. *ICES J. Mar. Sci.* **1952**, *18*, 117–140.
- 3. Smith, R.C.; Eppley, R.W.; Baker, K.S. Correlation of primary production as measured aboard ship in southern California coastal waters and as estimated from satellite chlorophyll images. *Mar. Biol.* **1982**, *66*, 281–288.
- 4. Eppley, R.W.; Stewart, E.; Abbott, M.R.; Owen, R.W. Estimating ocean production from satellite-derived chlorophyll: Insights from the Eastropac data set. *Oceanol. Acta Spec. Issue* **1987**, *SP*, 109–113.
- 5. Antoine, D.; André, J.M.; Morel, A. Oceanic primary production: 2. Estimation at global scale from satellite (coastal zone color scanner) chlorophyll. *Glob. Biogeochem. Cycles* **1996**, *10*, 57–69.
- 6. Behrenfeld, M.J.; Falkowski, P.G. Photosynthetic rates derived from satellite-based chlorophyll concentration. *Limnol. Oceanogr.* **1997**, 42, 1–20.
- 7. Tilstone, G.; Smyth, T.; Poulton, A.; Hutson, R. Measured and remotely sensed estimates of primary production in the Atlantic Ocean from 1998 to 2005. *Deep Sea Res. Part II Top. Stud. Oceanogr.* **2009**, *56*, 918–930.
- 8. Joo, H.; Son, S.; Park, J.W.; Kang, J.J.; Jeong, J.Y.; Lee, C.I.; Kang, C.K.; Lee, S.H. Long-Term pattern of primary productivity in the East/Japan sea based on ocean color data derived from MODIS-Aqua. *Remote Sens.* **2016**, *8*, 25, doi:10.3390/rs8010025
- 9. Campbell, J.; Antoine, D.; Armstrong, R.; Arrigo, K.; Balch, W.; Barber, R.; Behrenfeld, M.; Bidigare, R.; Bishop, J.; Carr, M.-E.; et al. Comparison of algorithms for estimating ocean primary production from surface chlorophyll, temperature, and irradiance. *Glob. Biogeochem. Cycles* **2002**, *16*, doi:10.1029/2001GB001444.
- Carr, M.E.; Friedrichs, M.A.; Schmeltz, M.; Aita, M.N.; Antoine, D.; Arrigo, K.R.; Asanuma, I.; Aumont, O.; Barber, R.; Behrenfeld, M.; et al. A comparison of global estimates of marine primary production from ocean color. *Deep Sea Res. Part II Top. Stud. Oceanogr.* 2006, 53, 741–770.
- 11. Friedrichs, M.A.; Carr, M.E.; Barber, R.T.; Scardi, M.; Antoine, D.; Armstrong, R.A.; Asanuma, I.; Behrenfeld, M.J.; Buitenhuis, E.T.; Chai, F.; et al. Assessing the uncertainties of model estimates of primary productivity in the tropical Pacific Ocean. *J. Mar. Syst.* **2009**, *76*, 113–133.
- 12. Saba, V.S.; Friedrichs, M.A.M.; Carr, M.-E.; Antoine, D.; Armstrong, R.A.; Asanuma, I.; Aumont, O.; Bates, N.R.; Behrenfeld, M.J.; Bennington, V.; et al. Challenges of modeling depth-integrated marine primary productivity over multiple decades: A case study at BATS and HOT. *Glob. Biogeochem. Cycles* **2010**, 24, doi:10.1029/2009GB003655.
- 13. Saba, V.S.; Friedrichs, M.A.M.; Antoine, D.; Armstrong, R.A.; Asanuma, I.; Behrenfeld, M.J.; Ciotti, A.M.; Dowell, M.; Hoepffner, N.; Hyde, K.J.W.; et al. An evaluation of ocean color model estimates of marine primary productivity in coastal and pelagic regions across the globe. *Biogeosciences* **2011**, *8*, 489–503.
- 14. Tilstone, G.H.; Taylor, B.H.; Blondeau-Patissier, D.; Powell, T.; Groom, S.B.; Rees, A.P.; Lucas, M.I. Comparison of new and primary production models using SeaWiFS data in contrasting hydrographic zones of the northern North Atlantic. *Remote Sens. Environ.* **2015**, *156*, 473–489.
- 15. Balch, W.; Evans, R.; Brown, J.; Feldman, G.; McClain, C.; Esaias, W. The remote sensing of ocean primary productivity: Use of a new data compilation to test satellite algorithms. *J. Geophys. Res. Oceans* **1992**, *97*, 2279–2293.
- 16. Berthon, J.F.; Morel, A. Validation of a spectral light-photosynthesis model and use of the model in conjunction with remotely sensed pigment observations. *Limnol. Oceanogr.* **1992**, *37*, 781–796.
- 17. Petrenko, D.; Pozdnyakov, D.; Johannessen, J.; Counillon, F.; Sychov, V. Satellite-derived multi-year trend in primary production in the Arctic Ocean. *Int. J. Remote Sens.* **2013**, *34*, 3903–3937.
- 18. Dierssen, H.M.; Smith, R.C. Bio-optical properties and remote sensing ocean color algorithms for Antarctic Peninsula waters. *J. Geophys. Res. Oceans* **2000**, *105*, 26301–26312.
- 19. IOCCG. Remote Sensing of Ocean Colour in Coastal, and Other Optically-Complex Waters; Sathyendranath, S., Ed.; Reports of the International Ocean-Colour Coordinating; IOCCG: Dartmouth, NS, Canada, 2000.
- 20. Volpe, G.; Santoleri, R.; Vellucci, V.; d'Alcalà, M.R.; Marullo, S.; d'Ortenzio, F. The colour of the Mediterranean Sea: Global versus regional bio-optical algorithms evaluation and implication for satellite chlorophyll estimates. *Remote Sens. Environ.* **2007**, 107, 625–638.

Remote Sens. 2018, 10, 1116 23 of 24

21. Campbell, J.W.; O'Reilly, J.E. Role of satellites in estimating primary productivity on the northwest Atlantic continental shelf. *Cont. Shelf Res.* **1988**, *8*, 179–204.

- 22. Platt, T.; Sathyendranath, S. Oceanic primary production: Estimation by remote sensing at local and regional scales. *Science* **1988**, 241, 1613–1620.
- 23. Platt, T.; Caverhill, C.; Sathyendranath, S. Basin-scale estimates of primary production by remote sensing: The North Atlantic. *J. Geophys. Res.* **1991**, *96*, 15147–15159.
- 24. Gregg, W.W.; Conkright, M.E. Global seasonal climatologies of ocean chlorophyll: Blending in situ and satellite data for the Coastal Zone Color Scanner era. *J. Geophys. Res. Oceans* **2001**, *106*, 2499–2515.
- 25. Milutinović, S.; Bertino, L. Assessment and propagation of uncertainties in input terms through an ocean-color-based model of primary productivity. *Remote Sens. Environ.* **2011**, *115*, 1906–1917.
- 26. Balch, W.M.; Byrne, C.F. Factors affecting the estimate of primary production from space. *J. Geophys. Res. Oceans* **1994**, 99, 7555–7570.
- 27. Finenko, Z.Z.; Suslin, V.V.; Churilova, T.Y. The regional model to calculate the Black Sea primary production using satellite color scanner SeaWiFS. *Mar. Ecol. J.* **2009**, *8*, 81–106. (In Russian)
- 28. Dogliotti, A.I.; Lutz, V.A.; Segura, V. Estimation of primary production in the southern Argentine continental shelf and shelf-break regions using field and remote sensing data. *Remote Sens. Environ.* **2014**, 140, 497–508.
- 29. Behrenfeld, M.J.; Falkowski, P.G. A consumer's guide to phytoplankton primary productivity models. *Limnol. Oceanogr.* **1997**, 42, 1479–1491.
- 30. Peterson, D.H.; Perry, M.J.; Bencala, K.E.; Talbot, M.C. Phytoplankton productivity in relation to light intensity: A simple equation. *Estuar. Coast. Shelf Sci.* **1987**, 24, 813–832.
- 31. Platt, T.; Sathyendranath, S. Modelling Marine Primary Production; IOCCG: Halifax, NS, Canada, 2002; 280p.
- 32. Lee, Z.P.; Carder, K.L.; Marra, J.; Steward, R.G.; Perry, M.J. Estimating primary production at depth from remote sensing. *Appl. Opt.* **1996**, *35*, 463–474.
- 33. Lee, Z.; Lance, V.P.; Shang, S.; Vaillancourt, R.; Freeman, S.; Lubac, B.; Hargreaves, B.R.; Castillo, C.D.; Miller, R.; Twardowski, M.; et al. An assessment of optical properties and primary production derived from remote sensing in the Southern Ocean (SO GasEx). *J. Geophys. Res. Oceans* **2011**, *116*, doi:10.1029/2010JC006747.
- 34. Kirk, J.T.O. *Light and Photosynthesis in Aquatic Ecosystems*, 3rd ed.; Cambridge University Press: Cambridge, UK, 2011; 649p.
- 35. Longhurst, A. Seasonal cycles of pelagic production and consumption. Prog. Oceanogr. 1995, 36, 77–167.
- 36. Vedernikov, V.I.; Gagarin, V.I.; Demidov, A.B.; Burenkov, V.I.; Stunzhas, P.A. Primary production and chlorophyll distributions in the subtropical and tropical waters of the Atlantic ocean in the autumn of 2002. *Oceanology* **2007**, *47*, 386–399.
- 37. Tilstone, G.H.; Lange, P.K.; Misra, A.; Brewin, R.J.W.; Cain, T. Microphytoplankton photosynthesis, primary production and potential export production in the Atlantic Ocean. *Prog. Oceanogr.* **2017**, *158*, 109–129.
- 38. IOC. Protocols for the Joint Global Ocean Flux Study (JGOFS) Core Measurements; UNESCO: Paris, France, 1994.
- 39. Morel, A.; Prieur, L. Analysis of variations in ocean color. Limnol. Oceanogr. 1977, 22, 709–722.
- 40. Lizon, F.; Seuront, L.; Lagadeuc, Y. Photoadaptation and primary production study in tidally mixed coastal waters using a Lagrangian model. *Mar. Ecol. Prog. Ser.* **1998**, *169*, 43–54.
- 41. Lee, Z.; Marra, J.; Perry, M.J.; Kahru, M. Estimating oceanic primary productivity from ocean color remote sensing: A strategic assessment. *J. Mar. Syst.* **2015**, *149*, 50–59.
- 42. Kahru, M.; Jacox, M.G.; Lee, Z.; Kudela, R.M.; Manzano-Sarabia, M.; Mitchell, B.G. Optimized multi-satellite merger of primary production estimates in the California Current using inherent optical properties. *J. Mar. Syst.* **2015**, *147*, 94–102.
- 43. Platt, T.G.; Gallegos, C.L.; Harrison, W.G. Photoinhibition of photosynthesis in natural assemblages of marine phytoplankton. *J. Mar. Res.* **1980**, *38*, 687–701.
- 44. Kiefer, D.A.; Mitchell, B.G. A simple, steady state description of phytoplankton growth based on absorption cross section and quantum efficiency. *Limnol. Oceanogr.* **1983**, *28*, 770–776.
- 45. Morel, A. Available, usable, and stored radiant energy in relation to marine photosynthesis. *Deep Sea Res.* **1978**, 25, 673–688.
- 46. Rabinovich, E. Photosynthesis; Foreign Lit: Moscow, Russia, 1953; Volume 2, 652p. (In Russian)

Remote Sens. 2018, 10, 1116 24 of 24

47. Marra, J.; Ho, C.; Trees, C. An Alternative Algorithm for the Calculation of Primary Productivity from Remote Sensing Data; Lamont Doherty Earth Observatory Technical Report (LDEO-2003-1); LDEO: Palisades, NY, USA, 2003.

- 48. Bannister, T.T. Production equations in terms of chlorophyll concentration, quantum yield, and upper limit to production. *Limnol. Oceanogr.* **1974**, *19*, 1–12.
- 49. Laws, E.A.; Bannister, T.T. Nutrient-and light-limited growth of *Thalassiosira fluviatilis* in continuous culture, with implications for phytoplankton growth in the ocean. *Limnol. Oceanogr.* **1980**, *25*, 457–473.
- 50. Babin, M.; Morel, A.; Claustre, H.; Bricaud, A.; Kolber, Z.; Falkowski, P.G. Nitrogen-and irradiance-dependent variations of the maximum quantum yield of carbon fixation in eutrophic, mesotrophic and oligotrophic marine systems. *Deep Sea Res. Part I Oceanogr. Res. Pap.* **1996**, 43, 1241–1272.
- 51. Morel, A.; Antoine, D.; Babin, M.; Dandonneau, Y. Measured and modeled primary production in the northeast Atlantic (EUMELI JGOFS program): The impact of natural variations in photosynthetic parameters on model predictive skill. *Deep Sea Res. Part I Oceanogr. Res. Pap.* **1996**, 43, 1273–1304.
- 52. Kyewalyanga, M.N.; Platt, T.; Sathyendranath, S.; Lutz, V.A.; Stuart, V. Seasonal variations in physiological parameters of phytoplankton across the North Atlantic. *J. Plankton Res.* **1998**, *20*, 17–42.
- 53. Suggett, D.; Kraay, G.; Holligan, P.; Davey, M.; Aiken, J.; Geider, R. Assessment of photosynthesis in a spring cyanobacterial bloom by use of a fast repetition rate fluorometer. *Limnol. Oceanogr.* **2001**, *46*, 802–810
- 54. Smyth, T.J.; Tilstone, G.H.; Groom, S.B. Integration of radiative transfer into satellite models of ocean primary production. *J. Geophys. Res. Oceans* **2005**, *110*, doi:10.1029/2004JC002784.
- 55. Picart, S.S.; Sathyendranath, S.; Dowell, M.; Moore, T.; Platt, T. Remote sensing of assimilation number for marine phytoplankton. *Remote Sens. Environ.* **2014**, *146*, 87–96.
- 56. Johnson, Z. Regulation of Marine Photosynthetic Efficiency by Photosystem II. Ph.D. Thesis, Graduate School of Duke University, Durham, NC, USA, 2000; 189p.
- 57. Morel, A.; Huot, Y.; Gentili, B.; Werdell, P.J.; Hooker, S.B.; Franz, B.A. Examining the consistency of products derived from various ocean color sensors in open ocean (Case 1) waters in the perspective of a multi-sensor approach. *Remote Sens. Environ.* **2007**, *111*, 69–88.
- 58. Emery, W.J.; Tomson, R.E. *Data Analysis Methods in Physical Oceanography*; Gulf Professional Publishing: Houston, TX, USA, 2001; 638p.
- 59. Taylor, K.E. Summarizing multiple aspects of model performance in a single diagram. *J. Geophys. Res. Atmos.* **2001**, *106*, 7183–7192.
- 60. Barnes, M.; Tilstone, G.H.; Smyth, T.J.; Suggett, D.; Kromkamp, J.; Astoreca, R.; Lancelot, C. Absorption based algorithm of primary production for total and size-fractionated phytoplankton in coastal waters. *Mar. Ecol. Prog. Ser.* **2014**, *504*, 73–89.
- 61. Tilstone, G.H.; Smyth, T.J.; Gowen, R.J.; Martinez-Vicente, V.; Groom, S.B. Inherent optical properties of the Irish Sea and their effect on satellite primary production algorithms. *J. Plankton Res.* **2005**, *27*, 1–22.
- 62. Curl, H.; Small, L.F. Variations in photosynthetic assimilation ratios in natural, marine phytoplankton communities. *Limnol. Oceanogr.* **1965**, *10*, R67–R73.
- 63. O'Reilly, J.E.; Maritorena, S.; Siegel, D.; O'Brien, M.C. Ocean color chlorophyll a algorithms for SeaWiFs, OC2, and OC4: Technical report. In *SeaWiFS Postlaunch Calibration and Validation Analyses, Part 3*; Postlaunch Technical Report Series; NASA Goddard Space Flight Centre: Greenbelt, MD, USA, 2000; Volume 11, pp. 9–23.
- 64. Bouman, H.A.; Platt, T.; Doblin, M.; Figueiras, F.G.; Gudmudsson, K.; Gudfinnsson, H.G.; Huang, B.; Hickman, A.; Hiscock, M.; Jackson, T.; et al. Photosynthesis-irradiance parameters of marine phytoplankton: Synthesis of a global data set. *Earth Syst. Sci. Data* **2018**, *10*, 251–266.
- 65. Sosik, H.M. Bio-optical modeling of primary production: Consequences of variability in quantum yield and specific absorption. *Mar. Ecol. Prog. Ser.* **1996**, *143*, 225–238.
- 66. Uitz, J.; Huot, Y.; Bruyant, F.; Babin, M.; Claustre, H. Relating phytoplankton photophysiological properties to community structure on large scales. *Limnol. Oceanogr.* **2008**, *53*, 614–630.
- 67. Kok, B. On the inhibition of photosynthesis by intense light. Biochim. Biophys. Acta 1956, 21, 234-244.
- 68. Miller, C.B.; Wheeler, P.A. Biological Oceanography; John Wiley & Sons: Hoboken, NJ, USA, 2012; 504p.
- 69. Ryther, J.H.; Menzel, D.W. Light adaptation by marine phytoplankton. *Limnol. Oceanogr.* 1959, 4, 492–497.
- 70. Steele, J.H. Environmental control of photosynthesis in the sea. Limnol. Oceanogr. 1962, 7, 137–150.



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