

Journal of the
MARINE BIOLOGICAL ASSOCIATION
of the United Kingdom



CAMBRIDGE
 UNIVERSITY PRESS

The role of cryptic dispersal in shaping connectivity patterns of marine populations in a changing world.



Journal:	<i>Journal of the Marine Biological Association of the United Kingdom</i>
Manuscript ID	JMBA-07-16-RE-0203.R1
Manuscript Type:	Review
Date Submitted by the Author:	n/a
Complete List of Authors:	David, Andrew; Clarkson University, Biology Loveday , Benjamin ; Plymouth Marine Laboratory
Keywords:	oceanography, modelling, genetics, larvae, introduction, invasive, species, aquaculture
Abstract:	Genetic connectivity directly shapes the demographic profile of marine species, and has become one of the most intensely researched areas in marine ecology. More importantly, it has changed the way we design and describe Marine Protected Areas across the world. Population genetics is the preferred tool when measuring connectivity patterns, however, these methods often assume that dispersal patterns are 1) natural and 2) follow traditional metapopulation models. In this short review, we formally introduce the phenomenon of cryptic dispersal, where multiple introductory events can undermine these assumptions, resulting in grossly inaccurate connectivity estimates. We also discuss the evolutionary consequences of cryptic dispersal and advocate for a cross-disciplinary approach that incorporates larval transport models into population genetic studies to provide a level of oceanographic realism that will result in more accurate estimates of dispersal. As globalized trade continues to expand, the rate of anthropogenic movement of marine organisms is also expected to increase and as such, integrated methods will be required to meet the inevitable conservation challenges that will arise from it.

SCHOLARONE™
 Manuscripts

1 **Title: The role of cryptic dispersal in shaping connectivity patterns of marine populations in a**
2 **changing world.**

3

4 Running Title: Cryptic dispersal in marine populations

5

6 Short Review Article

7

8 **Authors:** Andrew A. David^{a*}, Benjamin R. Loveday^b

9 ^aDepartment of Biology, Clarkson University, Potsdam, NY – 13699, USA

10 ^bPlymouth Marine Laboratory, Prospect Place, Plymouth, PL1 3DH, United Kingdom

11

12 ***Corresponding author:** Department of Biology, 8 Clarkson Avenue, Clarkson University, Potsdam NY

13 13699, Box 5805. Tel: (+1)3152684355, email: adavid@clarkson.edu

14

15

16

17

18

19

20

21

22

23

24

25

26

27 **Abstract**

28 *Genetic connectivity directly shapes the demographic profile of marine species, and has become one of*
29 *the most intensely researched areas in marine ecology. More importantly, it has changed the way we*
30 *design and describe Marine Protected Areas across the world. Population genetics is the preferred tool*
31 *when measuring connectivity patterns, however, these methods often assume that dispersal patterns are*
32 *1) natural and 2) follow traditional metapopulation models. In this short review, we formally introduce*
33 *the phenomenon of cryptic dispersal, where multiple introductory events can undermine these*
34 *assumptions, resulting in grossly inaccurate connectivity estimates. We also discuss the evolutionary*
35 *consequences of cryptic dispersal and advocate for a cross-disciplinary approach that incorporates*
36 *larval transport models into population genetic studies to provide a level of oceanographic realism that*
37 *will result in more accurate estimates of dispersal. As globalized trade continues to expand, the rate of*
38 *anthropogenic movement of marine organisms is also expected to increase and as such, integrated*
39 *methods will be required to meet the inevitable conservation challenges that will arise from it.*

40

41

42

43 **Keywords:** oceanography, modelling, genetics, larvae, introduction, invasive, species, aquaculture, vector,
44 ballast

45

46

47

48

49

50

51

52

53 Introduction

54 Understanding the dispersal capacity of marine organisms is the most widely researched, albeit least
55 understood area in marine ecology (Cowen & Sponaugle, 2009; Hellberg, 2009; Buston & D'Aloia, 2013;
56 Crook et al. 2015). Dispersal patterns govern population connectivity, which in turn influences important
57 ecological and evolutionary processes (Levin, 2006). As such, the study of dispersal dynamics is
58 fundamental to marine biodiversity and conservation research where it can help distinguish distinct
59 genetic lineages which are of evolutionary importance when designing marine protected areas (MPAs)
60 (Palumbi, 2003; Von der Heyden, 2009). Understanding the dispersal capacity of an organism is also
61 crucial for assessing the invasion potential of non-indigenous species (NIS) as it can serve as a reliable
62 proxy for measuring connectivity and genetic diversity in recently introduced populations (Roman &
63 Darling, 2007). In many fish and invertebrates, larval movement is ultimately responsible for dispersal on
64 both local and regional scales and for sessile animals such as tunicates, barnacles and sponges, to name a
65 few, it is the sole means of natural dispersal. However, due to the large numbers and minute sizes of
66 larvae, along with the vast expanse of the world's oceans, tracking and quantifying dispersal has been
67 notoriously difficult and some would argue, impossible (Metaxas & Saunders, 2009; Cowen &
68 Sponaugle, 2009; Selkoe & Toonen, 2011). As a result, alternative approaches have been developed
69 which offers indirect but pragmatic estimates of connectivity.

70 Population genetics has emerged as one of these alternatives and has proven to be a powerful tool
71 for measuring dispersal in the marine realm (Levin, 2006). When estimating dispersal capacity,
72 population genetics assumes that larval dispersal patterns follow traditional metapopulation models (e.g.
73 island, stepping stone, etc) and that the movement of alleles can be traced back to the natural movement
74 of individuals. The use of mitochondrial genetic markers (mtDNA) and nuclear genetic markers (nDNA)
75 are often employed. MtDNA markers such as the cytochrome b (Cyt b) and cytochrome c oxidase I (COI)
76 genes have high rates of sequence evolution and are often used to gain insights into past events that have
77 helped shape current genetic patterns (Avice, 2009). These markers are often integrated into a 'molecular
78 clock' which can provide divergence estimates that parallels important geological events such as sea level

79 rise and glacial retreats. In contrast, nDNA markers such as SNPs (single nucleotide polymorphisms)
80 show more variability when used in large quantities and are used to gain insights into the contemporary
81 movement of organisms. Higher resolution nuclear markers such as AFLPs (Amplified Fragment Length
82 Polymorphisms) and microsatellites are much more variable than both mitochondrial genes and SNPs and
83 usually provide deeper insights into recent dispersal events. While there is no 'ideal' marker, utilizing
84 both mtDNA in combination with nDNA markers provides a more holistic understanding of the genetic
85 architecture and connectivity patterns of populations (Karl et al. 2012).

86 The results of population genetic studies are often interpreted within the context of the species'
87 larval developmental strategy. For example, traditional life history theory posits that organisms producing
88 larvae with long planktonic larval duration (PLD) phases will be able to disperse to far distances and
89 therefore be expected to show high levels of connectivity among spatially separated populations. This was
90 based on the assumption that larvae act as passive particles and are at the mercy of the diffusive forces of
91 the pelagic environment (Selkoe & Toonen, 2011). In contrast organisms that exhibit abbreviated larval
92 development (short PLDs) or direct development (no planktonic phase) are expected to show high
93 recruitment rates and hence low levels of population connectivity. A meta-analysis by Shanks (2009)
94 provides the most comprehensive dataset thus far showing an acceptable correlation ($R^2 = 0.48$) between
95 PLD and dispersal distance. While genetic studies have found this to be true for many cases, recent
96 studies have shown increasing numbers of exceptions. These exceptions are important because they allude
97 to a more complex picture of dispersal in the marine realm. For example, environmental heterogeneity
98 such as haloclines, thermoclines, strong current systems, vertical stratification of the water column,
99 bathymetry and upwelling cells are all oceanographic features that can act as dispersal barriers, limiting
100 connectivity of a species regardless of PLD (Robinson et al. 2011). These barriers are sometimes known
101 as phylogeographic breaks because they are usually associated with known biogeographic boundaries that
102 can limit gene flow, thereby facilitating the formation of distinct genetic lineages (Figure 1). On the other
103 end, unorthodox dispersal vectors such as rafting has been shown to significantly increase population
104 connectivity of species that produces larvae with short PLDs phases or are direct developers (Nikula et al.

105 2013; Cumming et al. 2014; Donald et al. 2015). Independent of the aforementioned factors, genetic
106 estimates of dispersal are further complicated by issues such as inadequate and inaccurate taxon sampling
107 (the latter refers to potential cryptic species) (Wysor et al. 2002; Wrangle et al. 2016), chaotic genetic
108 patchiness, where significant genetic structure is observed in the absence of dispersal barriers (Kesaniemi
109 et al. 2014) and asymmetric dispersal patterns, where diversity across sites is wholly driven by diversity at
110 upstream locations thereby masking true patterns of connectivity and demography (Pringle & Wares,
111 2007; Wares & Pringle, 2008).

112 A recent study by David et al. (2016) coined the term, *cryptic dispersal* – a phenomenon where
113 the anthropogenic movement of organisms via vectors such as the aquaculture trade and transfer of ballast
114 water may erode phylogeographic signal, thereby reducing the power of genetic markers and in doing so
115 render gene flow and genetic connectivity estimates inaccurate. Cryptic dispersal therefore adds another
116 dimension of complexity to dispersal dynamics in the marine realm. The primary aim of this review is to
117 formally introduce the concept of cryptic dispersal, highlight some of the more recent studies that are
118 either potential or definitive cases of cryptic dispersal and briefly discuss the evolutionary consequences
119 of this phenomenon. This review does not aim to exhaustively review the effects of introductions on the
120 genetic architecture of populations, as this general topic has received considerable coverage in the
121 literature, but rather to hone in on the least understood and most inconspicuous effect of human-mediated
122 introductions. In this review, we distinguish ‘intentional’ vectors such as aquaculture and shipping from
123 rafting and attachment to floating structures, which are often inconsistent and have both a biological (e.g.
124 floating kelp bodies) and human (e.g. driftwood) component.

125

126

127

128

129

130

131 **Anthropogenic Movement as a Powerful Agent of Gene Flow**

132 Large scale human-mediated movement of marine organisms has occurred for centuries with the
133 emergence of the first wooden ships capable of harboring communities of fouling organisms such as
134 bryozoans, sponges, algae, barnacles, molluscs and tunicates (Carlton, 1989). After the twentieth century,
135 the 'dry' ballast of ships was later replaced with water which allowed planktonic organisms including the
136 larval stages of a variety of species to be pumped in and transported to sites located thousands of
137 kilometers away from their native habitats. Surveys by Carlton (1989) and colleagues at the Oregon
138 Institute of Marine Sciences had found over 200 species in ship ballast destined for Oregon from Japan;
139 all of which survived the trip. Also, a report by Chu et al. (1997) found a total of 81 species distributed
140 among five cargo containers in the Pacific destined for Hong Kong. In the last twenty years however,
141 there has been a fourfold increase in the growth of transoceanic shipping, partly driven by technological
142 advancements that have produced larger and faster ships and partly by the rapid pace of globalization that
143 has opened up new international trade routes (Tournadre, 2014; Cope *et al.*, 2015) (Figure 2A). For
144 example, a recent survey of hull fouling by Ashton et al. (2016) estimated 680,000 arrivals per year of
145 barnacle species at ports distributed across the Atlantic and the Pacific coasts. Considering that only 15
146 commercial vessels were sampled, it is likely that this number was an underestimate. The most important
147 ecological consequence of transoceanic shipping is the increased introductions of NIS which has
148 subsequently resulted in higher rates of invasion events (Roman & Darling, 2007).

149 In addition to shipping, the aquaculture trade has also been an important vector for the movement
150 of organisms both regionally and globally (Elton 1995; Grosholz *et al.* 2015). Commercial shellfish such
151 as oysters, abalone and mussels are often transported across long distances for transplantation purposes
152 (Figure 2B). These shellfish may harbor a variety of organisms which can reside within or inside crevices
153 of the shells, in mudpacks that accompany brood stocks or even within the organism itself. For example,
154 the introduction of the Pacific oyster *Crassostrea gigas* to Europe resulted in the arrival of more than a
155 dozen NIS, with about five or six eventually becoming established (Wolff & Reise, 2002). In a more

156 recent episode, the polychaete *Diopatra biscayensis* in France was able to expand its range across a
157 phylogeographic break due to anthropogenic transport on mussel seed ropes (Woodin *et al.*, 2014).

158 While marine invasions are an important consequence of anthropogenic movement of NIS, a
159 more conspicuous phenomenon is the erosion of phylogeographic signal due to continuous and consistent
160 movement of migrants (Wares *et al.*, 2002; Dawson *et al.*, 2005; David *et al.*, 2016; Wrange *et al.* 2016).
161 This phenomenon is coined as ‘cryptic dispersal’ since the anthropogenic effect cannot be definitively
162 detected by genetic patterns alone (David *et al.*, 2016). Cryptic dispersal is primarily driven by propagule
163 pressure and also by the coastal environment, specifically the strength of phylogeographic breaks. If two
164 distinct populations of a species are separated by a strong break, isolated introductory events that
165 exchanges propagules from both populations will probably not significantly alter genetic structure and
166 such introductions could be easily detected by genetic markers (Darling *et al.*, 2008; Reitzel *et al.*, 2008;
167 Reusch *et al.*, 2010). However, if these introductory events become continuous and consistent, closely
168 mimicking metapopulation migration models (e.g. stepping stone and island models), then
169 phylogeographic signal may become eroded, driving down Wright’s fixation index (F_{ST} values) and
170 giving the illusion of low genetic structure and high connectivity. Furthermore, if cryptic dispersal has
171 been occurring across longer timescales, even genetic patterns inferred from mtDNA may be obscured via
172 reshuffling of ancient haplotypes due to past translocation events (Wrange *et al.* 2015; David *et al.*, 2016;
173 Williams *et al.* 2016).

174 Cryptic dispersal highlights an important limit to population genetics, which is that the movement
175 of genes does not necessarily correlate with natural movement. This is an important point because it
176 opens up the possibility of drawing grossly inaccurate interpretations of dispersal patterns from genetic
177 data in regions where distinct barriers exist. Perhaps the most vulnerable population genetic studies are
178 those that ‘detect’ a panmictic population, which is defined as naturally dispersed endemic populations
179 that freely interbreed due to the absence of dispersal barriers. In these studies, introductory events may be
180 suggested as an after-thought or never at all and the lack of structure is usually attributed to the species’
181 ‘strong dispersal capabilities’. For example, studies by Wrange *et al.* (2016) found high frequency of

182 shared mtDNA and microsatellite haplotypes in globally separated populations of the barnacle *Balanus*
183 *improvisus* which produces planktonic larvae. This genetic pattern was primarily attributed to
184 anthropogenic dispersal mechanisms, despite the fact that the authors were unable to definitively
185 distinguish between oceanographic connectivity and anthropogenic dispersal. Another recent genetic
186 study by Hudson et al. (2016) found little genetic differentiation in the tunicate *Ciona intestinalis* which
187 exhibits abbreviated development. Interestingly, this study was carried out in *C. intestinalis*' native range
188 but again, the authors were unable to determine whether the observed genetic pattern was attributed to
189 anthropogenic or natural dispersal. An interesting phylogenetic and phylogeographic study by Ciotir &
190 Freeland (2016) on invasive cattails recently described the process of 'cryptic intercontinental dispersal'
191 where the horticulture trade was responsible for the widespread dispersal of a variety of cattail species.
192 However, like the previous two studies, much of the data on phylogeographic signal was inconclusive.

193 The most obvious solution to the cryptic dispersal problem will be the development of a tool that
194 can discern the relative contribution of both natural and anthropogenic dispersal types to the observed
195 genetic patterns of a population. To accomplish this from the anthropogenic side one would need to be
196 able to quantify the number of migrants of the study species being carried in each ship's ballast per route.
197 However, considering the sheer amount of shipping traffic that occurs along a typical coastal system
198 combined with the millions of tons of ballast water that are pumped in and out per trip – even with
199 environmental DNA (eDNA) as a monitoring tool for identification, such a task would be logistically
200 impossible. An interesting study by Darling et al. (2012) attempted to investigate possible correlations
201 between vector patterns and genetic connectivity of an invasive tunicate (*Styela clava*) in the northeastern
202 Pacific. The authors compiled shipping data (specifically vessel routes) for the northeastern Pacific which
203 was then used to create a shipping connectivity matrix of the region. Their results showed that the genetic
204 data failed to capture the anthropogenic effects of shipping, which supports the aforementioned view that
205 such an approach for evaluating cryptic dispersal is problematic and in many cases impractical. With
206 respects to aquaculture, the task of tracking shellfish movement is considerably less onerous than large
207 transoceanic shipping vessels. In addition, the shellfish in a brooding stock that were transplanted can be

208 examined individually and the target hitchhiker species can be quantified and processed for genetic
209 studies. However, there are currently no known studies that have carried out experimental transplants to
210 this extent and is therefore an area ripe for future research.

211

212 **Evolutionary Consequences of Cryptic Dispersal**

213 Populations that are separated by phylogeographic breaks are genetically differentiated units that are
214 locally adapted to their environment (Irwin, 2012). While these distinct units may show some level of
215 phenotypic divergence such as size or colour variation, gene-flow 'leakage' across dispersal barriers is
216 enough to prevent speciation events. In a system where cryptic dispersal is occurring, we would expect
217 that these dispersal barriers will be weakened. This weakening would occur as human-mediated transport
218 (e.g. ballast water transfer) deliver a sufficient number of propagules to overcome local adaptation. If
219 propagules are being transported in this manner, then populations can be homogenized via some form of
220 reverse speciation which was defined as "a reversal of the processes that lead to the diversification of
221 species pairs" (Taylor et al. 2006). This is important to consider because genetic homogenization
222 ultimately results in a loss of genetic diversity. For example, in many population genetic studies, a source
223 population is often the one that has the highest haplotype or nucleotide diversity. However, if cryptic
224 dispersal is occurring then not only is phylogeographic signal being diluted but 'original' haplotypes of
225 the source population are being distributed and re-distributed across multiple sink populations at a high
226 enough frequency to obfuscate the detectability of a distinct source. If genetic variation supplies the raw
227 material for evolution, then it follows that cryptic dispersal could reduce the evolutionary potential of an
228 entire species. For example, a review of the aquaculture industry with regards to introductions highlighted
229 the dangers of repeated translocations in fish stocks where such activities can reduce genetic diversity of
230 commercially important species (Johnson, 2000).

231 Here, we would like to emphasize that the reduction in the evolutionary potential of a species due
232 to cryptic dispersal is a phenomenon that is expected to occur largely in introduced species, where a
233 history of vector transport has already been established. While dispersal in the native habitats could also

234 be candidates, detection may be more difficult due to the longer evolutionary histories of these species.
235 Interestingly, if multiple introductions are the driving force behind the homogenization process, there is
236 the possibility that the expected reduction in genetic diversity could be buffered by individuals arriving
237 and carrying unique haplotypes from a completely different source. A recent study by Lejeusne et al.
238 (2014) illustrated such a scenario where high levels of gene flow were detected in a Palaemonid shrimp
239 using the COI genetic marker. The authors also found high genetic diversity which was attributed to
240 multiple introductions with international shipping being the culprit vector. Another recent study by David
241 et al. (2016) used the cytochrome b gene and a single nuclear locus to detect high genetic connectivity
242 among populations (no geographic patterning of haplotypes) of a shell-boring polychaete distributed
243 across three phylogeographic breaks in South Africa. The movement of oysters among aquaculture farms
244 distributed along the country's ~2000 km coastline was identified as the main driver of this high
245 connectivity (David et al. 2016; Williams et al. 2016). Despite the high connectivity levels, genetic
246 diversity was low which was probably due to the homogenizing effect of cryptic dispersal along with a
247 lack of individuals arriving from genetically distinct sources.

248 Adaptability also plays an important role in the cryptic dispersal capacity of a species since
249 propagules would not only have to be consistently transported across phylogeographic breaks, but would
250 also have to survive and thrive in the different biogeographic regions. For example, in the case of *P.*
251 *hoplura*, experimental studies found that the species was capable of surviving in temperatures as low as
252 12°C and as high as 24°C, with both temperatures characteristic of the Atlantic Ocean on the west coast
253 of the country and the Indian Ocean on the east coast respectively (David & Simon, 2014). It is therefore
254 not surprising that the high genetic connectivity observed in this species could have been mistaken for
255 panmixia.

256

257 **Integrating Ocean Models into Population Genetic Studies to Detect Cryptic Dispersal**

258 The most practical method for assessing cryptic dispersal will involve focusing on natural
259 movement - a process which can be numerically modeled. Studies integrating high resolution larval

260 transport models (LTM) into population genetic studies to measure dispersal have been on the rise in the
261 past few years, partly due to advances in computing capabilities and the costs associated with accessing
262 them (Viard *et al.* 2006; Galindo *et al.*, 2006; Selkoe *et al.*, 2008; Baums *et al.*, 2006; White *et al.* 2010).
263 Because LTMs incorporate the prevailing hydrographic conditions of the study area, they add a high
264 degree of oceanographic realism to dispersal studies, which is especially important for understanding
265 contemporary movement of larvae (Selkoe *et al.*, 2008). LTMs coupled with population genetics offer a
266 powerful means of assessing cryptic dispersal since a larval transport model can act as a control, depicting
267 what connectivity patterns should look like in the absence of anthropogenic movement. Once connectivity
268 patterns are determined based on the model, they can be cross validated with genetic patterns. One of the
269 first comprehensive studies to utilize this approach was conducted by Dawson *et al.* (2005) who assessed
270 the population structure of a supposedly highly dispersed cosmopolitan jellyfish, *Aurelia* sp. The authors,
271 using mtDNA and a single nDNA loci, found high levels of genetic connectivity among global
272 subpopulations. However, their larval transport model showed limited connectivity that coincided with
273 known phylogeographic breaks, indicating that multiple introductory events over a longer time scale,
274 possibly via shipping vectors, could have eroded the phylogeographic signal, giving the illusion of a
275 panmictic population (Dawson *et al.*, 2005). In a similar but more recent study, David *et al.* (2016) found
276 that the aquaculture trade in South Africa was facilitating genetic connectivity in the invasive polychaete
277 *Polydora hoplura*, which is notorious for burrowing and residing in oyster and abalone shells. The
278 authors found a lack of any clear geographic patterning of haplotypes and low F_{ST} despite the fact that
279 populations were distributed across multiple phylogeographic breaks. However, a high-resolution
280 transport model found limited connectivity that coincided with these breaks. It was known at the time that
281 oyster farmers frequently transported their stock among farms that are widely distributed along the
282 country's coast and in a non-directional manner (Simon *et al.*, 2006; Haupt *et al.*, 2010). This movement
283 resulted in the polychaete being moved with the oysters, across the breaks, consequentially resulting in a
284 reduction in signal and an elimination of any geographic clustering of haplotypes (Williams *et al.* 2016)
285 (Figure 3).

286 While the aforementioned studies used discordance between the LTMs and genetics to propose
287 the existence of cryptic dispersal, others have ruled out cryptic dispersal when both approaches show
288 congruent results. For example, Viard *et al.* (2006) assessed the dispersive capacity of the introduced
289 gastropod, *Crepidula fornicata* along the French coast in the Bay of Biscayne using microsattellites and a
290 simple 2D hydrographic model. The authors found that low F_{ST} values (high genetic connectivity)
291 correlated with the model's estimate of extensive dispersal along the coast and assumed that the pattern
292 was a direct result of the larva's dispersal capabilities. However, this study explicitly assumed that there
293 was no anthropogenic transport occurring and it was conducted using a 2D model on a regional scale.

294 High resolution LTMs are based on Lagrangian mechanics, which presents a 3D numerical
295 representation of velocities at different depths (Siegel *et al.*, 2003). Larvae are represented by virtual
296 floats, which are deployed at specific localities in the model. Dispersal simulations are then carried out
297 and repeated for a number of years using the available ocean circulation data for each year (Figure 4).
298 Valuable data concerning connectivity patterns include dispersal trajectory and density maps along with
299 particle capture data which can be analyzed both qualitatively and quantitatively. The complexity of the
300 model can be increased by incorporating specific biological characteristics into the floats, such as
301 duration in the plankton (which determines how far the floats will be carried by surface currents),
302 mortality rates (which will determine the number of floats that would be 'captured' at a pre-determined
303 site) and fecundity (which determines the number of floats per simulation run). The most recent
304 generation of transport models that are often used in conjunction with population genetics is the Regional
305 Oceanic Modeling System (ROMS) (Shchepetkin & McWilliams, 2005; Baums *et al.* 2006; Selkoe *et al.*
306 2008). While model predictions can offer valuable insights into the 'pure' movement of larvae, it is
307 important to note that ocean models, like all computer models, do possess limitations. For example,
308 LTMs are limited by the knowledge of important ecological processes involved in dispersal. In other
309 words, how well do we know our study species? Many species, especially fishes can produce larvae that
310 do not act as passive floaters and are capable of counteracting the advective effects of currents by actively
311 adjusting their orientation in the water column or exhibiting diel vertical migrations (Levin, 2006). In

312 such cases, incorporating appropriate life history parameters into the virtual floats along with adding
313 drag-drift effects into the simulations is essential for accurately modeling dispersal in such a species.
314 Perhaps the biggest limitation is that for models to be as accurate as possible, they need to be able to fully
315 capture coastal processes, especially nearshore circulation patterns which are responsible for determining
316 particle trajectory and supply/recruitment results.

317 The strength of using seascape genetics to detect cryptic dispersal lies in the power of cross-
318 validation. If populations show limited dispersal based on LTM estimates but show high connectivity
319 based on the genetic data (e.g. low non-significant F_{ST} , non-significant isolation by distance and mixed
320 haplotypes), it is likely that cryptic dispersal is occurring. However, this approach is only useful if
321 connectivity patterns are discordant. In scenarios where high connectivity is estimated by both population
322 genetic studies and LTMs, it would be virtually impossible to discern the contributions of anthropogenic
323 transport to the observed genetic pattern. One possible solution would be to integrate both physical
324 oceanography and population genetic data into a time-step model that includes an estimate of the number
325 of propagules being transported in a vessel at any given time. As computing power continues to increase,
326 we expect the development of these types of complex predictive models to emerge within the field of
327 marine invasion biology, which would greatly aid in providing informative data that can be used to
328 mitigate the loss of diversity caused by cryptic dispersal.

329

330 Conclusions

331 Over the last few decades, genetics has provided crucial data on the dispersal potential and
332 connectivity patterns of a great number of species. This has given us novel insights into important marine
333 ecological process and has challenged us to re-evaluate conservation methodologies such as the way
334 marine reserves are designed. However, as humans continue to affect every aspect of the marine
335 environment, especially through biological invasions, the need for cross-disciplinary collaboration is
336 crucial in order to respond to these new challenges. Here we highlighted the phenomenon of cryptic
337 dispersal, where multiple introductory events can mimic traditional migration models, thereby diluting or

338 eroding phylogeographic signal which gives the illusion of a naturally dispersing species. Such a
339 phenomenon is problematic since the erosion of dispersal barriers can allow contact between spatially
340 separated populations, thereby initiating the homogenization process. In order to solve this problem, we
341 outlined the importance of utilizing both population genetics and larval transport models (LTMs). These
342 LTMS will not only be able to act as a control to detect cryptic dispersal but has also been shown to
343 elucidate fine scale ocean processes that can be related back to genetic patterns (Gilg & Hilbish, 2003).
344 While previous studies have focused on the novelty of using this integrated approach (White *et al.*, 2010),
345 here we call for such an approach to be regarded as the gold standard for evaluating connectivity patterns
346 on large and complex coastal systems.

347

348

349 **Acknowledgements**

350 We would like to thanks Dr. Jon Wares and another anonymous reviewer for greatly improving the final
351 version of this manuscript. We would also like to thank MarineTraffic for their assistance in developing
352 the shipping density map and Dr. Tamara Robinson for valuable input.

353

354 **References**

355 Buston P.M., D'Aloia C.C. (2013) Marine ecology: reaping the benefits of local dispersal. *Current*
356 *Biology* 23: R351–R353.

357 Chu K.H, Tam P.F, Fung C.H and Chen Q.C. (1997) A biological survey of ballast water in container
358 ships entering Hong Kong. *Hydrobiologia* 352: 201–206.

359 Ciotir C. and Freeland J. (2016) Cryptic intercontinental dispersal, commercial retailers, and the genetic
360 diversity of native and non-native cattails (*Typha* spp.) in North America. *Hydrobiologia* 768:

361 137 – 150.

- 362 Cope R.C., Prowse T.A.A., Ross J.V, Wittmann T.A. and Cassey P. (2015) Temporal modelling of ballast
363 water discharge and ship-mediated invasion risk to Australia. *Royal Society Open Science* 2:
364 150039.
- 365 Cowen R.K and Sponaugle S. (2009) Larval dispersal and marine population connectivity. *Annual*
366 *Review of Marine Science* 1: 443–466.
- 367 Crook D.A., Lowe W.H., Allendorf F.W., Eros T., Finn D.S., Gillanders B.M., Hadwen W.L., Harrod C.,
368 Hermoso V., Jennings S., Kilada R.W., Nagelkerken I., Hansen M.M., Page T.J., Riginos C., Fry
369 B., Hughes J.M. (2015) Human effects on ecological connectivity in aquatic ecosystems:
370 integrating scientific approaches to support management and mitigation. *Science of the Total*
371 *Environment* 534: 52 – 64.
- 372 Cumming R.A., Nikula R., Spencer H.G., Waters J.M. (2014) Transoceanic genetic similarities of kelp-
373 associated sea slug populations: long-distance dispersal via rafting? *Journal of Biogeography* 41:
374 2357 – 2370.
- 375 dos Santos A., Santos M.P., Conway D.V.P., Bartilotti C., Lourenco P. and Queiroga H. (2008) Diel
376 vertical migration of decapod larvae in the Portuguese coastal upwelling ecosystem: implications
377 for offshore transport. *Marine Ecology Progress Series* 359: 171–183
- 378 Darling J.A., Bagley M.J., Roman J., Tepolt C.K. and Geller G.B. (2008). Genetic patterns across
379 multiple introductions of the globally invasive crab genus *Carcinus*. *Molecular Ecology* 17:
380 4992–5007.
- 381 Darling J.A., Herborg L.M. and Davidson I.C. (2012) Intracoastal shipping drives patterns of regional
382 population expansion by and invasive marine invertebrate. *Ecology and Evolution* 2: 2557–2566.
- 383 David A.A., Matthee C.A., Loveday B.R. and Simon C.A. (2016) Predicting the dispersal potential of an
384 invasive polychaete pest along a complex coastal biome. *Integrative and Comparative Biology*
385 56: 600 – 610.
- 386 David A.A. and Simon C.A. (2014) The effect of temperature on larval development of two non-
387 indigenous poecilogonous polychaetes (Annelida: Spionidae) with implications for life-history

- 388 theory, establishment and range expansion. *Journal of Experimental Marine Biology and Ecology*
389 461: 20 – 30.
- 390 Dawson M.N., Gupta A.S. and England M.H. (2005) Coupled biophysical global ocean model and
391 molecular genetic analyses identify multiple introductions of cryptogenic species. *Proceedings of*
392 *the National Academy of Sciences of the United States of America* 102: 11968–11973.
- 393 Elton C.S. (1958) *The ecology of invasions by animals and plants*. Methuen, London, England. 181 pp.
- 394 Galindo H.M., Olson D.B. and Palumbi S.R. (2006) Seascape genetics: a coupled oceanographic-genetic
395 model predicts population structure of Caribbean corals. *Current Biology* 1622–1626.
- 396 Gilg M.R. and Hilbish T.J. (2003) The geography of marine larval dispersal: coupling genetics with fine-
397 scale physical oceanography. *Ecology* 84: 2989–2998.
- 398 Grosholz E.D., Crafton R.E., Fontana R.E., Pasari J.R., Williams S.L. and Zabin CJ (2015) Aquaculture as
399 a vector for marine invasions in California. *Biological Invasions* 17: 1471–1484.
- 400 Haupt T.M., Griffiths C.L., Robinson T.B. and Tonin A.F.G. (2010) Oysters as vectors of marine alines,
401 with notes on four introduced species associated with oyster farming in South Africa. *African*
402 *Zoology* 45: 52–62.
- 403 Hellberg M.E. (2009) Gene flow and isolation among populations of marine animals. *Annual Review of*
404 *Ecology, Evolution and Systematics* 40: 291–310.
- 405 Hellberg M.E., Burton R.S., Neigel J.E. and Palumbi S.R. (2002) Genetic assessment of connectivity
406 among marine populations. *Bulletin of Marine Science* 70: 273 – 290.
- 407 Hudson J., Viard F., Roby C. and Rius M. (2016) Anthropogenic transport of species across native
408 ranges: unpredictable genetic and evolutionary consequences. *Biology Letters* 12: 20160620.
- 409 Karl S.A., Toonen R.J., Grant W.S. and Bowen B.W. (2012) Common misconceptions in molecular
410 ecology: echoes of the modern synthesis. *Molecular Ecology* 21: 4171 – 4189.
- 411 Kesaniemi J.E., Hansen B.W., Banta G.T. and Knott K.E. (2014) Chaotic genetic patchiness and high
412 relatedness of a poecilogonous polychaete in a heterogenous estuarine landscape. *Marine Biology*
413 161: 2631 – 2644.

- 414 Levin LA. (2006) Recent progress in understanding larval dispersal: new directions and digressions.
415 Integrative and Comparative Biology 46: 282–297.
- 416 McDonald K.M., Winter D.J., Ashcroft A.L. and Spencer H.G. (2015) Phylogeography of the whelk
417 genus *Cominella* (Gastropoda: Buccinidae) suggests long-distance counter-current dispersal of a
418 direct developer. Biological Journal of the Linnean Society 115: 315–332.
- 419 Metaxas A. and Saunders M. (2009) Quantifying the “Bio-” components in biophysical models of larval
420 transport in marine benthic invertebrates: advances and pitfalls. Biological Bulletin 216: 257–272.
- 421 Nikula R., Spencer H.G. and Waters J.M. (2013) Passive rafting is a powerful driver of transoceanic
422 gene-flow. Biology Letters 9: 20120821.
- 423 Palumbi S.R. (2003) Population genetics, demographic connectivity, and the design of marine reserves.
424 Ecological Applications 13: 146–158.
- 425 Pfeiffer-Herbert A.S., McManus M.A., Raimondi P.T., Chao Y. and Chai F. (2007) Dispersal of barnacle
426 larvae along the central California coast: a modeling study. Limnology and Oceanography 52:
427 1559–1569.
- 428 Pringle J.M. and Wares J.P. (2007) Going against the flow: maintenance of alongshore variation in allele
429 frequency in a coastal ocean. Marine Ecology Progress Series 335: 69 – 84.
- 430 Reitzel A.M., Darling J.A., Sullivan J.C. and Finnerty J.R. (2008) Global population genetic structure of
431 the starlet anemone *Nematostella vectensis*: multiple introductions and implications for
432 conservation policy. Biological Invasions 10: 1197–1213.
- 433 Reusch T.B.H., Bolte S., Sparwel M., Moss A.G. and Javidpour J. (2010) Microsatellites reveal origin and
434 genetic diversity of Eurasian invasions by one of the world’s most notorious marine invader,
435 *Mnemiopsis leidyi* (Ctenophora). Molecular Ecology 19: 2690–2699.
- 436 Shchepetkin A. and McWilliams J. (2005) The Regional Oceanic Modeling System (ROMS): a split-
437 explicit, free-surface, topography-following-coordinate ocean model. Ocean Model 9: 347–404.
- 438 Selkoe K.A., Henzler C.M. and Gaines S.D. (2008) Seascape genetics and the spatial ecology of marine
439 populations. Fish and Fisheries 9: 363–377.

- 440 Selkoe K.A. and Toonen R.J. (2011) Marine connectivity: a new look at pelagic larval duration and
441 genetic metrics of dispersal. *Marine Ecology Progress Series* 436: 291 – 305.
- 442 Siegel D.A., Kinland B.P., Gaylord B. and Gaines S.D. (2003) Lagrangian descriptions of marine larval
443 dispersion. *Marine Ecology Progress Series* 260: 83–96.
- 444 Simon C.A., Ludford A. and Wynne S. (2006) Spionid polychaetes infesting cultured abalone, *Haliotis*
445 *midiae*, in South Africa. *African Journal of Marine Science* 28: 167–171.
- 446 Teske P.R., Von der Heyden S., McQuaid C.D. and Barker N.P. (2011) A review of marine
447 phylogeography in southern Africa. *South African Journal of Science* 107: 43–53.
- 448 Viard F., Ellien C. and Dupont L. (2006) Dispersal ability and invasion success of *Crepidula fornicata* in
449 a single gulf: insights from genetic markers and larval-dispersal model. *Helgoland Marine*
450 *Research* 60: 144–152.
- 451 Von der Heyden S. (2009) Why do we need to integrate population genetics into South African marine
452 protected area planning? *African Journal of Marine Science* 31: 263–269.
- 453 Wares J.P. and Pringle J.M. (2008) Drift by drift: effective population size is limited by advection. *BMC*
454 *Evolutionary Biology* 8: 235
- 455 White C., Selkoe K.A., Watson J., Siegel D.A., Zacherl D.C. and Toonen R.J. (2010) Ocean currents help
456 explain population genetic structure. *Proceedings of the Royal Society of London B* DOI:
457 10.1098/rspb.2009.2214.
- 458 Wrange A.L., Charrier G., Thonig A., Alm Rosenblad M., Blomberg A., Havenhand J.N., Jonsson P.R.
459 and Andre C. (2016) The story of a hitchhiker: population genetic patterns in the invasive
460 barnacle *Balanus* (*Amphibalanus*) *improvisus* Darwin 1854. *PLoS One* 11: e0147082.
- 461 Wysor B., Kooistra H.C.F. Wiebe, Fredericq S. (2002) Comparative phylogeography of reticulate
462 cladophorean algae. *Journal of Phycology* 38: 38 – 39.
- 463
- 464
- 465

466 Figure Legends

467 Fig 1. Examples of phylogeographic breaks on the southern African coast. Breaks coincide with major the
468 biogeographic boundaries that separate cool-temperate waters of the Atlantic coast from the warm-
469 temperate south coast and the warm-temperate south coast from the sub-tropical and tropical coasts of the
470 Indian Ocean. Map modified from Teske *et al.* (2011)

471

472 Fig. 2. A) Overall Density Map showing global vessel traffic for the year 2015 based on AIS satellite data
473 and B) Batch of farmed oysters from offshore cultivation beds in Saldanha Bay, South Africa.

474

475 Fig 3. Cryptic dispersal of the shell-boring polychaete *Polydora hoplura* in South Africa via the
476 aquaculture trade. South Africa. Step 1: planktotrophic larvae enters shellfish farm through the inflow,
477 step 2: larvae settles, undergo metamorphosis and burrows into farmed oysters, step 3: infested oysters are
478 transported to geographically distant farm, step 4: brooding females release larvae into the water column,
479 step 5: fraction of larvae escapes into the wild via the outflow. Dashed arrows refer to spat or adult
480 oysters imported into the farm (locally and internationally) and exported to other farms in the region.

481

482 Fig. 4. Ocean circulation model built using the Regional Oceanic Modeling System (ROMS). Model
483 shows 289,788 possible trajectories of virtual floats that were recovered after being deployed at three sites
484 along the southern African coast (Jacobsbaai, Hermanus and Haga Haga) with the Cape Point
485 phylogeographic break highlighted. Total of 1271 floats were deployed each month from 1991-2010.

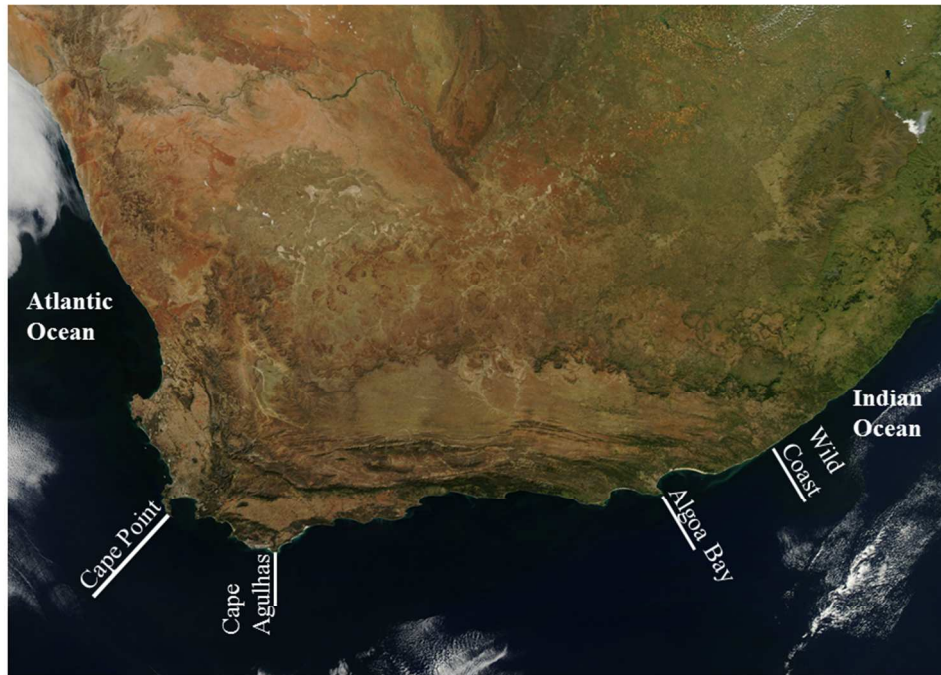


Fig 1. Examples of phylogeographic breaks on the southern African coast. Breaks coincide with major the biogeographic boundaries that separate cool-temperate waters of the Atlantic coast from the warm-temperate south coast and the warm-temperate south coast from the sub-tropical and tropical coasts of the Indian Ocean. Map modified from Teske et al. (2011)

254x190mm (96 x 96 DPI)

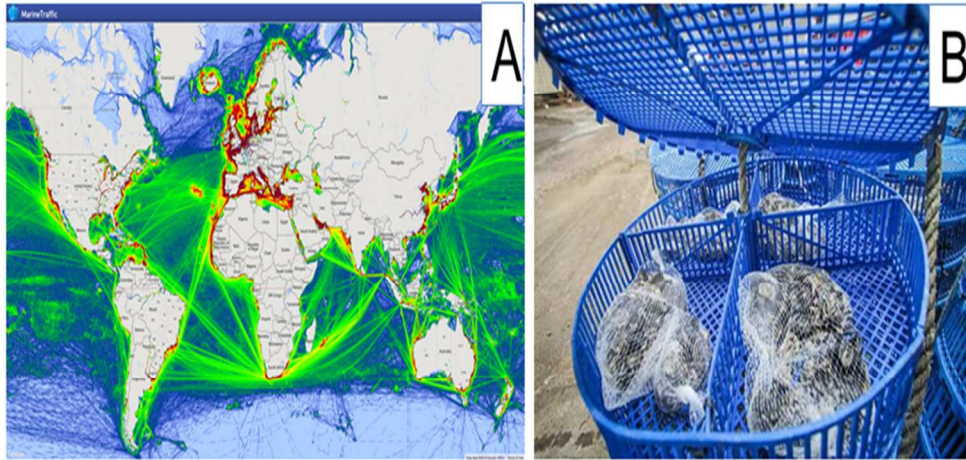


Fig. 2. A) Overall Density Map showing global vessel traffic for the year 2015 based on AIS satellite data and B) Batch of farmed oysters from offshore cultivation beds in Saldanha Bay, South Africa.

254x77mm (96 x 150 DPI)

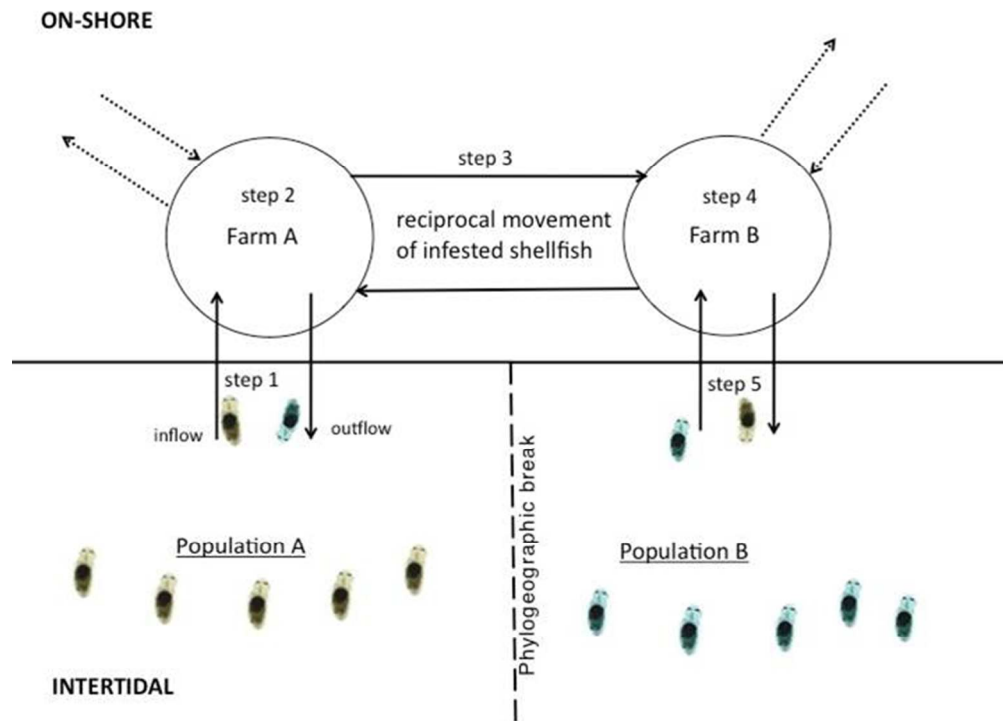


Fig 3. Cryptic dispersal of the shell-boring polychaete *Polydora hoplura* in South Africa via the aquaculture trade. South Africa. Step 1: planktotrophic larvae enters shellfish farm through the inflow, step 2: larvae settles, undergo metamorphosis and burrows into farmed oysters, step 3: infested oysters are transported to geographically distant farm, step 4: brooding females release larvae into the water column, step 5: fraction of larvae escapes into the wild via the outflow. Dashed arrows refer to spat or adult oysters imported into the farm (locally and internationally) and exported to other farms in the region.

143x108mm (127 x 127 DPI)

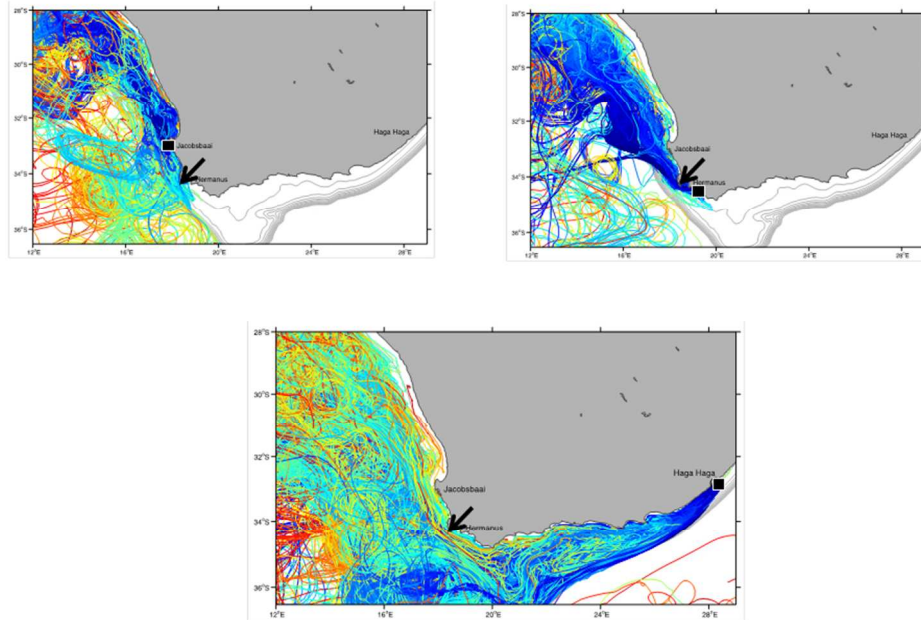


Fig. 4. Ocean circulation model built using the Regional Oceanic Modeling System (ROMS). Model shows 289,788 possible trajectories of virtual floats that were recovered after being deployed at three sites along the southern African coast (Jacobsbaai, Hermanus and Haga Haga) with the Cape Point phylogeographic break highlighted. Total of 1271 floats were deployed each month from 1991-2010.

254x190mm (96 x 96 DPI)