

SHORT COMMUNICATIONS

## PCR from the CPR offers a historical perspective on marine population ecology

R.R. Kirby\* and P.C. Reid†

\*Plymouth Marine Laboratory, Citadel Hill, The Hoe, Plymouth, PL1 2PB. †The Sir Alister Hardy  
Foundation for Ocean Science, 1 Walker Terrace, The Hoe, Plymouth, PL1 3BN  
E-mail: r.kirby@pml.ac.uk

The Continuous Plankton Recorder (CPR) survey has collected plankton samples from regular tracks across the world's oceans for almost 70 y. Over 299,000 spatially extensive CPR samples are archived and stored in buffered formalin. This CPR archive offers huge potential to study changes in marine communities using molecular data from a period when marine pollution, exploitation and global anthropogenic impact were much less pronounced. However, to harness the amount of data available within the CPR archive fully, it is necessary to improve techniques of larval identification, to genus and species preferably, and to obtain genetic information for historical studies of population ecology. To increase the potential of the CPR database this paper describes the first extraction, amplification by the polymerase chain reaction and utilization of a DNA sequence (mitochondrial 16S rDNA) from a CPR sample, a formalin fixed larval sandeel.

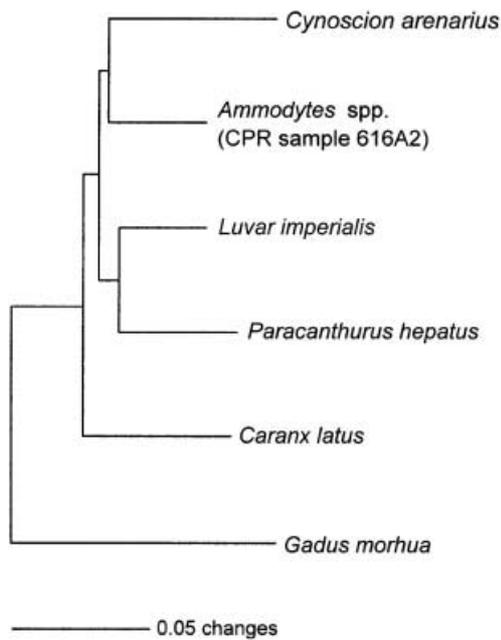
Molecular genetic analysis has already been used for the discrimination of marine invertebrate larvae (Medeiros-Bergen et al., 1995) and its use in population ecology is well known (Avisé, 1994). Amplification of DNA by polymerase chain reaction (PCR) is a powerful tool in population ecology and archival material is a potentially important source of molecular data. Since most archival material is preserved in formalin, methods have been developed to enable DNA to be amplified from this material and, to date, DNA has been successfully amplified from samples up to 85-y old (Shiozawa et al., 1992; France & Kocher, 1996; Shedlock et al., 1997; Wirgin et al., 1997; Coombs et al., 1999). However, although these methods offer great potential, few ecological questions have yet benefited from the analysis of archival specimens.

The archived database of the Continuous Plankton Recorder (CPR) survey (Warner & Hays, 1994) represents a major resource for spatial and temporal investigations of marine populations (Reid et al., 1998; Beaugrand et al., 2000). The survey has collected plankton data from the world's seas for almost 70 y (Warner & Hays, 1994). CPRs are towed at 10-m depths by volunteer ships along regular tracts at monthly intervals and plankton samples are collected onto a slow moving 280- $\mu$ m silk mesh immersed in formalin. On return to the laboratory the silk mesh is cut into sections equivalent to ten nautical miles and the phytoplankton and zooplankton analysed; the samples are then stored in borax buffered formalin ( $\sim$ pH 7.0). Since the survey began in 1931 every effort has been made to maintain the consistency and integrity of the database. Today, the archive comprises approximately 299,000 spatially extensive samples extending from a period when marine pollution, exploitation and global anthropogenic impact were much less pronounced. The ability to obtain DNA from this archival material would facilitate a major new approach to measuring long-term change in marine communities. In addition, molecular analyses will be an important aid in identifying the different larval stages of many of the planktonic species, particularly the arthropods. Therefore, to increase the potential of the CPR database, this paper describes the extraction and

utilization of a DNA sequence from a formalin-fixed CPR sample.

In this initial study two methods of DNA extraction were applied to separate 10 mg tissue samples taken from a single formalin-fixed larval sandeel (*Ammodytes* spp.) (CPR sample 616A2 obtained from a virgin uncounted silk square stored in borax-buffered formalin): (i) a PCR-Chelex extraction technique (Coombs et al., 1999); and (ii) a modification (France & Kocher, 1996) of the protocol developed by Shiozawa et al. (1992). Following extraction the DNA was resuspended in 50  $\mu$ l of sterile water. From 1  $\mu$ l of each DNA sample, partial fragments of two mitochondrial DNA (mtDNA) loci were amplified by PCR: (i) a partial fragment of the mtDNA 16S rDNA gene was amplified using 10 pmol of each of the universal amplification primers 16SAR and 16SBR (Palumbi et al., 1991); and (ii) a partial fragment on the mtDNA cytochrome-*b* gene was amplified using primers CBI-L and CB2-H (Kocher et al., 1989). *Taq* DNA polymerase (Life Technologies) was used in a reaction volume of 50  $\mu$ l. The thermal cycling conditions involved a 'hot start' with an initial denaturation of 2 min at 94°C followed by 30 cycles of 1 min at 94°C, 1 min annealing at 54°C and 30 s extension at 72°C, and a final extension of 10 min at 72°C using an MJ Research PTC-100 thermal cycler.

16S rDNA amplification products of the expected size (614 bp) were obtained from both DNA extraction methods. However, amplification products of the expected size for cytochrome-*b* (358 bp) were only obtained for the chelex based method. Only the 16S rDNA amplification products from the chelex extraction were agarose-gel purified, polished with *Pfu* polymerase (Stratagene), cloned into the plasmid pBluescript II KS- (Stratagene) and sequenced manually (eight clones were sequenced in both orientations to determine a consensus) for verification. Comparison of the sequence to Genbank indicated the 16S rDNA sequence obtained from CPR sample 616A2 (GenBank accession number: AF315121) was similar to mitochondrial 16S rDNA of the Perciformes of which the *Ammodytes* (sandeels) belong (Figure 1).



**Figure 1.** Neighbour joining tree on Jukes-Cantor distances (Jukes & Cantor, 1969) calculated using 511 nucleotides of aligned mitochondrial 16S rDNA sequence information from each marine species and constructed using PAUP (Swofford, 2001). The unidentified larval sandeel clusters among the marine Perciformes. The tree was rooted using the mitochondrial 16S rDNA sequence of the cod, *Gadus morhua*. The scale bar represents the number of changes per site.

The ability to obtain DNA sequence from CPR archive material represents an important advance creating new opportunities to exploit the extensive CPR archive to investigate change in the population ecology of the marine plankton communities. Immediate applications are to use the CPR archive to obtain data on intraspecific variation in DNA to examine geographical variability and long-term changes in the ecology and population dynamics of key planktonic and pelagic species. Pertinent questions and processes that can be addressed using the CPR database include recent change in fisheries and their planktonic food sources, fish larval recruitment, patterns of stock recovery following over-fishing and population differentiation. For example, to manage and conserve fish stocks successfully it is essential to know their population structure. An immediate and appropriate application of genetic techniques would be to examine change in the fish stocks of the North Sea. The North Sea fish stocks are subject to intense commercial fishing pressure and during the last seventy years have experienced great change. For example, the herring population is recovering from a total collapse of the population through overfishing, the cod is presently in severe decline and the fishery of sandeels, the largest single-species fishery in the North Sea, is so great that there is concern for the region's ecology. The CPR survey has sampled the North Sea particularly intensively over the last 70 y and it is hoped that the techniques described above when applied to these data will help understand the population ecology of these fisheries to better aid their conservation and management.

In addition to the examination of fisheries, applications of this work also include the molecular identification of invertebrate planktonic larvae and their developmental stages. Although an identifiable larva was used for this study, the

ability to use this approach to classify unidentifiable larvae represents an important advance in marine plankton ecology. For example, echinoderm and bivalve larvae are extremely difficult to identify to species, yet they are key members of ecosystems like the North Sea and are implicated in recent dramatic community changes attributed to trawling of the seabed by the fishery (Lindeboom & de Groot, 1998).

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